

Factors Influencing the Survival of Pathogenic Microbes in the Built (i.e. hospitals) and Natural Environments

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بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

In the name of Allah, the Most Merciful, the Most Kind

Dedication

I dedicated this work to my parents (Nahlah Khawajh, Abdulrahim Alaeq) who continue to learn, grow and develop and who has been a source of encouragement and inspiration to me throughout my life. Also, I am grateful to my sisters: Roaa, Rola, Rawan, and Reman and my brothers Ali and Rakan and my friends for their spiritual support and encouragement.

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ABSTRACT

Healthcare associated infections, i.e. nosocomial infections, occur in patients under medical care. These infections occur during stays in hospital and cause prolonged hospitalisation, disability, and an economic burden. Nosocomial pathogens include bacteria, viruses and fungal parasites. Pathogens living in the healthcare environment and equipment can be an obvious source of pathogen transmission. These pathogens can be transmitted by person to person contact or via contaminated water and food, infected individuals, contaminated healthcare personnel's skin or contact via shared items and surfaces. Pathogens can survive in the hospital environment for long periods and, in some cases, resist disinfection. Bacteria are the most common pathogens responsible for nosocomial infections. The Thesis begins with a description of the aims of the work, followed by studies the distribution of bacteria in the environment and their survival in the healthcare settings, the determination of the number of bacteria on hands after washing and drying normally and following the use of a warm air dryer. The factors which influence the survival of bacteria and *Candida* in the built environment were also determined on ceramic tiles, copper and plastic plumbing surfaces, and on new toothbrushes. Pathogenic bacteria were isolated from used toothbrushes and the effect of toothpastes on the growth of pathogenic bacteria was determined, as was the effectiveness of antibacterial cloths in inhibiting the growth of bacteria and yeast. The metabolic diversity of the bacterial isolates was also determined. Bacteria were isolated from a range of surfaces commonly found in hospitals and health care settings. A wide variety of bacteria were isolated from sinks, computer keyboards and computer mice, taps and the surface of mobile phones and toilet mirrors. Species of *Bacillus* were exclusively isolated from vacuum cleaner dust and from books and dust obtained from a library. Bacteria other than just *Bacillus* were isolated from the soles of shoes. Lift buttons were found to be contaminated with bacteria, not surprisingly

with species which are typically skin commensals, with the number being highest on the ground floor-call button. Bacteria were found to be spread by hot-air hand dryers, both into the toilet environment and onto previously washed hands. It is provisionally recommended that disposable paper towels are used in preference to such machines. Bacteria and the yeast, *Candida rugosa* survived when inoculated onto both rough and smooth tile surfaces similar to those used in health care settings. A wide range of potentially pathogenic bacteria were isolated from used toothbrushes and the survival of inoculated bacteria on tooth brushes was determined. The bacteria were shown to survive for varying periods, a finding of some concern in relation to dental hygiene. A range of toothpastes were also shown to be antibacterial. The survival of inoculated bacteria on copper and plastic surfaces typically used as piping in health care settings was determined. Copper surfaces were shown to be antibacterial, while plastic surfaces were not. It is therefore suggested that in critical healthcare areas, copper piping should be given preference over the plastic variety. Proprietary antibacterial clothes were tested for their antibacterial properties. Despite being marketed for this purpose, the cloths showed no obvious, marked antibacterial activity.

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CHAPTER 1

DISTRIBUTION AND SURVIVAL OF BACTERIA IN HEALTHCARE SETTINGS

1.1 Introduction

A wide range of microorganisms can be isolated from the environment, including bacteria, viruses, filamentous fungi and yeasts. Particular emphasis has been placed on the role of bacteria in medicine, but fungi (including yeasts) are also causal agents of many plant and animal diseases although their role as human pathogens is often underplayed. In a recent study setting (Perlroth *et al.*, 2007), for example, fungi were estimated to kill at least as many people as tuberculosis or malaria, over the past decades. The pathogenicity of both bacteria and yeasts is particularly potentially dangerous in the case of immunocompromised patients, in which they cause potentially life-threatening diseases (Sullivan *et al.*, 1997).

A hospital-acquired infections (HAIs), also known as a **nosocomial infection**, are acquired in health care facilities. Health care staff can spread infection, as can contaminated equipment, bed linens, or air droplets (McBryde *et al.*, 2004). The infection can originate from the outside environment, infected patients or staff or in some cases the source of the infection is unknown. The microorganism may originate from the patient's own skin microflora, becoming opportunistic after surgery or other procedures that compromise the protective barrier of the skin. It is estimated that in The US around 1.7 million hospital-associated infections occur

and contribute to 99,000 deaths each year. Nosocomial infections can cause severe pneumonia and infections of the bloodstream, urinary tract and other areas of the body. Many of these infections are difficult to treat with antibiotics (Klebens *et al.*, 2007).

1.2 Organisms involved

Methicillin resistant *Staphylococcus aureus*, *Candida albicans*, *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Stenotrophomonas maltophilia*, *Clostridium difficile*, *Escherichia coli*, and Vancomycin-resistant *Enterococcus*.

1.3 Sterilization

Sterilization kills all microbes on equipment and surfaces through exposure to chemicals, ionizing radiation, dry heat, or steam under pressure.

1.4 Isolation

Isolation precautions are designed to prevent transmission of microorganisms by common routes in hospitals. It involves the isolation of infectious cases in special hospitals and patients with infected wounds and joint transplantation patients in specific rooms. The following are important considerations regarding hygiene in health-care settings:

1.4.1 Handwashing

Handwashing is frequently the single most important act which reduces the risks of transmitting skin microorganisms between people or from one site to another on the same patient (Hugonnet *et al.*, 2002). Prompt, thorough washing of hands between patient contacts and after contact with blood, body fluids, secretions, excretions, and equipment or articles contaminated by them is a major component of infection control and isolation (Katz, 2004). The spread of nosocomial infections, among immunocompromised patients is linked to health care workers' hand contamination in nearly half of all hands requires correct hand-hygiene procedures flora. The first relates to microbes picked up by workers from the environment which can survive on human skin and sometimes to grow (Katz, 2004). The second group is represented by the permanent microbes living on the skin surface which have low pathogenicity and infection rate, and can prevent colonization by more pathogenic bacteria; microbes making up the resident flora include: *Staphylococcus epidermidis*, *S. hominis*, and *Micrococcus*, *Propionibacterium*, *Corynebacterium*, *Dermobacterium*, and *Pitosporum spp.*, while transient organisms comprise, *S. aureus*, and *Klebsiella pneumoniae*, and *Acinetobacter*, *Enterobacter* and *Candida spp.* Hand hygiene is aimed at eliminating the transient flora using a careful and dedicated hand washing with different kinds of soap, (normal and antiseptic), and alcohol-based gels. The lack of available sinks and time-consuming performance of hand washing presents problems related to its effectiveness (Langley, 2002).

1.4.2 Use of gloves

As well as hand washing, gloves play an major role in reducing the risks of microbial transmission. Gloves are worn to reduce the chance that microorganisms present on the hands of personnel can be transmitted to patients during invasive or other patient-care procedures and are also worn to limit the likelihood that the hands of personnel contaminated with microbes from a patient transmits the to another patient. Gloves must be changed between patient contacts, and hands should be washed after glove removal.

The wearing of gloves does not replace the need for handwashing, because gloves may have undetectable defects and hands can also become contaminated during glove removal; failure to change gloves between patient contacts is an obvious infection control hazard.

1.4.3 Antimicrobial surfaces

Microbes can survive on inanimate ‘touch’ surfaces for long periods, a fact which is troublesome in hospital environments in which patients with immunodeficiency are at increased risk of contracting nosocomial infections.

Common touch surfaces include bed rails, call buttons, touch plates, chairs, door handles, light switches, grab rails, intravenous poles, dispensers (alcohol gel, paper towel, soap), dressing trolleys, and counter and table tops are known to be contaminated with *Staphylococcus*, MRSA and vancomycin resistant *Enterococcus* (VRE) (Chemaly *et al.*, 2014). A number of compounds can

decrease the risk of bacteria growing on surfaces including: copper, silver, and germicides such as hydrogen peroxide vapour (Otter and French, 2009).

Nosocomial infections in the blood can cause serious medical problem and can result in major health care expenditure and death. The onset of infections in the bloodstream can be exacerbated by the previous use of antibiotics, corticosteroids, and a variety of chemotherapeutic agents. Major medical problems such cancers; neutropenia are also associated with increased problems from blood infections as is surgery and haemodialysis (Khan *et al.*, 2015).

The hospital environment is a potential reservoir of bacterial pathogens which are able to infect patients with a diverse variety of pathogenic microbes and thereby expose a large number of susceptible individuals to significant morbidity and mortality. Bacterial pathogens have an innate ability to survive for long periods on surfaces in the hospital environment (Dancer, 2009). Bacterial pathogens, isolated from hospital environment, are also beginning to develop resistance to multiple antimicrobial agents; these then cause difficulty in the treatment of nosocomial infections. The environment of patients is, as a result, heavily contaminated by infectious multidrug resistant organisms including methicillin-resistant *S. aureus* (MRSA), vancomycin-resistant Enterococci (VRE), *Clostridium difficile*, *Acinetobacter baumannii* and *Pseudomonas aeruginosa*, all of which are emerging to produce a major problem for healthcare systems (Dancer, 2009).

Infections in hospitals are, in the main, caused by factors such as cross-contamination between medical personnel, patients and visitors. Hospital surfaces

and equipment can also act as major reservoirs and sources of such cross infection (Cesar-Pastor *et al.*, 2012). Ubiquitous pathogens such as bacteria (*Escherichia coli*, *Enterococcus spp*, *Acinetobacter spp*, species of *Pseudomonas* and *Staphylococcus aureus*), viruses (e.g. noroviruses) and fungi (notably *Candida*) are particularly troublesome in the hospital environment because of the way they can survive for long periods on surfaces or humans (Thomas *et al.*, 2004). The uniforms of medical staff are often contaminated with multi-resistant *Staphylococcus aureus* and thereby provide a source of cross contamination (Boyce *et al.*, 1997). Air conditioning systems also act as source of airborne fungi such as species of *Aspergillus*, which can cause lung problems, especially amongst immunocompromised patients (Thomas *et al.*, 2004). Temperature is also a factor in the spread and development of hospital disease, especially in over-heated and under-ventilated environments which are often associated with the hospital environment, notably in winter (Ribera *et al.*, 1994).

1.5 Pathogen transmission

The transmission of contaminated pathogens on environmental surfaces is dependent on many factors including: the ability of pathogens to remain viable on dry surfaces; their ability to remain virulent after environmental exposure; the ability to colonize patients; the ability to transiently colonize the hands of health care workers and finally, the ability to develop resistance to disinfectants used on environmental surfaces (Weber *et al.*, 2010). Bacteria are transmitted by direct

contact between hands, body fluids such as saliva and mucus droplets; dust contaminated by bacteria in the air; via patient carers and via contaminated objects or equipment (Ortega *et al.*, 2010)

1.6 Survival of bacteria

Pathogens are able to survive in the hospital environment for long periods and, in some cases, resist disinfection. The frequency of environmental contamination with multi-resistant bacteria is greater for patients with infected wounds or were supplied with urinary drainage systems. Gastmeier *et al.* (2006) showed a correlation between survival time and diversity of important nosocomial pathogens. Most Gram-positive bacteria can survive for months on dry surfaces; *Enterococcus spp.* (including VRE) for example, can survive for 5 days to 4 months, *Staphylococcus aureus* remains viable for from 7 days to 7 months and *Streptococcus pyogenes* survives for 3 days to 6.5 months. Many Gram-negative species can also survive for several months. *Acinetobacter spp.* for example, can survive for 3 days to 5 months on environmental surfaces, *Escherichia coli* persist for 16 months, while *Klebsiella spp.* remain viable for more than 30 months (Kramer *et al.*, 2006). Vancomycin-resistant *Enterococcus* (VRE) can remain viable on inanimate surfaces from seven days to two-four months (Burke, 2010). The duration of bacterial survival depends on the bacteria concerned and the nature of the contaminated surfaces. For example, Enterococci survive for more than 24h on bed-frames, 18h on cotton, 1h on telephones, 30 min on the diaphragm of a stethoscope, and 1 hour on gloves and hands (Talon, 1999).

Vancomycin-resistant *Enterococcus* have also been recovered from 3 of 10 seat cushions located in a room occupied by a VRE patient.

1.7 Responses of microorganisms to stress

Microbial cells sense stress in order to protect themselves against the deleterious effects of heat (56°C), freeze–thaw injury, chemical or osmotic shock, exposure to reactive oxygen and nitrogen and to antimicrobial peptides or proteins. These responses involve physiological adaptations that counterbalance damage and allow the cells to continue to survive and grow (Griffiths, 2005). Entry into the stationary phase produces a general stress response, resulting in microbial resistance to multiple stresses. For example, when *Escherichia coli* is exposed to a sub-lethal stress, some of the cells die while some can be recovered on growth media. Growth of the recovered cell is dependent on a number of factors, including the level of stress, the nature of the recovery conditions and the growth phase of cell when the stress was imposed (Griffiths, 2005). Stationary phase cells show a greater tolerance to stress than exponentially growing ones. *E. coli* cells become shorter and rounder, the mode of metabolism alters and changes in membrane and cell wall structure occur (Griffiths, 2005). This response is seen in facultative anaerobes such as *Salmonella* and *Staphylococcus aureus* but generally not in strictly fermentative organisms such as *Streptococcus mutans*. *Campylobacter jejuni* also shows a reduced resistance during the stationary phase (Griffiths, 2005).

1.8 Responses of microorganisms to stress include:

- 1- Synthesis of protective proteins that repair cell damage, cell maintenance and bring about the eradication of stress agents.
- 2- Increases in resistance to lethal factors.
- 3- Transformation of cells to a latent state, e.g. the formation of spores.
- 4- Evasion of the host's defence mechanisms.
- 5- The production of adaptive mutations.

1.8.1 Types of microbial stress adaptation

The mechanisms of bacterial defence against environmental conditions are divided into two classes:

- 1- Limited or specific adaptive response resulting from microbial exposure to physical, chemical or biological stress, which protects cells against the subsequent lethal effects following the same stress (Griffiths, 2005).
- 2- Multiple adaptive responses (cross-protection) which occur when bacterial cells adapt to an inherent physiological condition or to an environmental factor, which results in microbial protection against subsequent lethal treatments, including stresses to which the microorganism had not been previously exposed. This type of protection is produced by a variety of stress conditions such as cell starvation,

exposure to high or low temperatures, high osmolality, and low pH (De Angelis and Gobbetti, 2004).

1.8.2 Starvation-stress response

When *E. coli*, *Salmonella*, and many other microorganisms are starved, they respond by inducing the expression of up to 50 new proteins or pre-existing proteins. The genetic and physiologic reprogramming that occurs is termed the starvation-stress response (SSR). The SSR allows for long term starvation survival of bacteria and provides generalized cross-resistance to a variety of other environmental stresses. In order to protect bacteria from damage, physiologic changes that occur during SSR include; the degradation of cellular RNA, proteins, and fatty acids; the reduction in the number of ribosomes; alterations in the amounts and type of nine lipids in the cytoplasmic membrane; an increase amounts of lipopolysaccharide in the outer membrane of Gram-negative bacteria; and increase the concentration of chromosomal DNA (Griffiths, 2005).

1.9 Epidemiologically significant pathogens

1.9.1 Bacteria

The main reservoirs for MRSA are infected patients and personnel in hospitals, and not surprisingly, the degree of pathogen contamination directly relates to the degree of localised infection in patients and on bandages, urine or blood. Both *Staphylococcus aureus* and MRSA can survive for up to nine weeks

(Popovich *et al.*, 2008). *Staphylococcus aureus* causes a variety of infections including skin, soft tissue infections, BSIs (blood stream infections), pneumonia, meningitis, endocarditis, and toxic shock syndrome (Popovich *et al.*, 2008).

Coagulase-Negative Staphylococci (CoNS) are able to form biofilms on foreign devices located within patients, including prosthetic joints, pacemakers, intravenous catheters and shunts (Cervera *et al.*, 2009). Hidron *et al.* (2008) found that CoNS are easily the most frequent cause of central line-linked blood system infections (CLABSI) and the second most common cause of surgical site infections (SSIs). Less commonly, CoNS infections result in catheter associated urinary tract infections (CA-UTIs) and ventilator-associated pneumonias (VAPs) (Martin *et al.*, 1989). These types of infections are now seen as increasingly common their occurrence has increased with the use of inter-body devices.

Wisplinghoff *et al.* (2004) concluded that 9% of CLABSI are caused by *Enterococcus* species, of which 2% of *E. faecalis* isolates; 60% of *E. faecium* isolates were found to be vancomycin resistant. VRE contamination has been found in up to 37% of samples obtained from the environment associated with gowns, health care workers, medical equipment, and microsphere beds.

Clostridium difficile spores are durable and resistant to unusual cleaning methods and contamination of the hospital environments by spores has been reported. Such spores are highly resistant and may survive for months in the environment.

Hospital floors remain contaminated with these spores for up to five months and contamination density is increased by presence of patients infected with diarrhea

(Hidron *et al.*, 2008). The gastrointestinal tract of young people is a reservoir of *C. difficile* which is transmitted via the faecal-oral route, directly by hand carriage by health care workers (HCWs), or by patient-to-patient contact or indirectly from a contaminated environmental source (Burke, 2010). Best *et al.* (2010) sampled the air and environmental surfaces adjacent to patients with symptomatic CDI and found *C. difficile* near a majority of the patients. Clinicians around the world have noted an increase in disease caused by this bacterium in patients with CDIs.

Hidron *et al.* (2008) demonstrated that *E. coli* and *P. aeruginosa* are the Gram-negative organisms which are most commonly isolated from health care-associated infections; less commonly isolated organisms include *Klebsiella pneumoniae*, *Enterobacter* species, *Acinetobacter baumannii*, and *Klebsiella oxytoca*.

1.9.2 Opportunistic mycoses

Opportunistic mycoses affect the skin, mucosa and the internal organs and are caused by both yeast and moulds. Such infections are associated with weakness in the host's immune defences, especially as the result of long-term and severe immunosuppressed. The most important pathogenic fungi are *Candida albicans*, *Aspergillus* spp., *Cryptococcus neoformans*, *Cryptosporidium*, *Malassezia* spp., and *Saccharomyces cerevisiae* (Badiee and Hashemizadeh, 2014). In addition to *Candida* and other yeasts, infections can also be caused by phaeohyphomycetes and hyalohyphomycetes. These organisms act as primary infection foci, usually affecting the upper or lower respiratory tract and can spread hematogenously and lymphogenously to infect additional organs (Badiee and Hashemizadeh, 2014). At

least 70% of all human *Candida* infections are caused by *C. albicans*, and the rest by *C. parapsilosis*, *C. tropicalis*, *C. guilliermondii*, *C. krusei*. Diba *et al.* (2012) identified pathogenic fungi from the environment including: *Candida albicans*, *C. krusei*, *C. glabrata*, *C. tropicalis* and *C. parapsilosis*. Of the isolates 35(31.5%) were shown to be *Candida* species, 48(43.2%), *Aspergillus* and 28 (20.3%) of other species.

1.10 Microorganisms used throughout the following

Staphylococci aureus is a non-motile small spherical cell (1µm) found in grapelike clusters. It is a Gram- positive coccus, catalase-producing bacteria, facultative anaerobe and can be cultured on normal nutrient media at 37° C. Extracellular enzymes and exotoxins produced by this bacterium, such as coagulase, alphatoxin, leukocidin, exfoliatins, enterotoxins, and toxic shock toxin are responsible for the resultant clinical symptoms and infections. *Staphylococcus aureus* is a frequent pathogen in nosocomial infections and it causes limited outbreaks in hospitals of problems including: furuncles, food poisoning, dermatitis exfoliativa, toxic shock syndrome, carbuncles, wound infections, sinusitis, otitis media, and mastitis puerperalis (Spaulding *et al.*, 2014).

Escherichia coli is a Gram-negative, flagellated rod whose natural habitat is the intestines of animals and humans. This bacterium is a major human pathogen that causes many infections including: lower urinary tract infection such as urethritis,

cystitis, urethrocystitis; infections of the renal pelvis and kidneys (cystopyelitis, pyelonephritis), wound infections, gallbladder and bile ducts infections, peritonitis, meningitis patients. *E. coli* also causes acute urinary tract infections in 70–80% of cases and in chronic, persistent infections in 40–50% of cases, as well as about 15% of all cases of nosocomial sepsis (*S. aureus* 20%).

Candida rugosa is a budding oval yeast Branched pseudohyphae are frequently seen and occasionally septate mycelia, with chains of elongated blastoconidia. *Candida rugosa*-pseudohyphae differentiates it from *C. lusitaniae*, *C. parapsilosis*. *Candida rugosa* has recently been cited as a possible “emerging” fungal pathogen capable of causing invasive infection in immunocompromised patients especially following the use of catheters but also via other modes of nosocomial acquisition (Pfaller, 2006). Behera *et al.* (2010) identified *Candida rugosa* as a causative pathogen of candidaemia, thirteen out of the 19 patients (68.4%) with *C. rugosa* candidaemia died.

1.11 Molecular biology techniques

16S rRNA and 18S rRNA analysis has been applied here to identify both bacteria and the yeasts isolates studied here. DNA can be isolated from a variety of specimens including from tissues, blood, bones, sperm, plant, hair and bacteria (Lahiri, 1992). DNA can also be used for laboratory diagnosis and in forensics (Hill *et al.*, 2000). In DNA analysis tissues or cells are first broken and the cells

are lysed using detergents or enzymes. This is followed by centrifugation to separate the DNA from other components and by DNA purification (Amann *et al.*, 1995).

1.11.1 Deoxyribonucleic acid (DNA)

Deoxyribonucleic acid (DNA) is the genetic material in living organisms. DNA plays an essential role for storing of the biological information due to its polymeric structure and because there are different nucleotides.

DNA is a polymer; a long, chainlike molecule consists of two long polynucleotide chains made up of four types of nucleotide sub-units called monomers. Each chain is called a “DNA chain” or “DNA strand”, and both chains are linked together by hydrogen bonds. The Nucleotides are composed of five carbon sugars which are linked with one or more phosphate groups and nitrogen containing base. The sugar is a deoxyribose attached with a single phosphate and one of the nitrogen bases Adenine (A), cytosine (C), guanine (G) or thymine (T). These nucleotides are attached together in a chain with the sugars and the phosphate groups form a backbone. The nucleotide subunits are linked together in a specific way that gives the DNA molecule chemical polarity. The two ends of the chain will be differing One is a hole 3' hydroxyl and the other is a knob 5' phosphate (Madigan *et al.*, 2012).

1.11.2 Polymerase Chain Reaction (PCR) technique

PCR is a powerful method of amplifying specific DNA sequences invented by Kary Mullis in 1983 (Mullis, 1990). The PCR technique used in clinical microbiology and research laboratory for identify the microbial pathogens. It used for detect pathogens which has a slow growth rate, clarify the taxonomic positions of known pathogens and help genotyping for microbial characterization which is become more specific and easily quantified between different organisms (Valones *et al.*, 2009). This technique depends on repeated synthesis of target DNA using DNA polymerase through three different steps under controlled temperatures in many thermal cycles. The steps are: **Denaturation** involves melting the double strand of the target DNA and separate them to single strands under high temperature 90-98°C for 1 minute. **Annealing** which allows the two added primers (oligodeoxyribonucleotides) to anneal with the separated DNA to be amplified. The primers anneal to the opposite side of the DNA strands in their 3' end under low temperature 37-65°C for 45 seconds. In the **extension**, the DNA polymerase synthesis a new DNA strand which is complementary to the target DNA by adding the deoxynucleotides (dNTPs) from the reaction mixture of the 5'-to-3' direction under optimum temperature for DNA polymerase 75–80°C for 2 minutes. These steps run in a single cycle so many cycles are needed to amplify the specific DNA to millions of copies. PCR reaction set of 30 cycles. Final elongation: this step is optional and requires a temperature of 70-74°C for 5-15 minutes after the final step of PCR to ensure that all single strand of DNA is elongated completely. Final

hold: in this step, the reaction mixture is cooled at 4-15°C for a short time to allow the products to be stored.

The aim of the work described here was to:

- 1) Studies the distribution of bacteria in the environment and their survival in the healthcare settings including: sinks, lift buttons, computer keyboards and computer mice, mobile telephones, mirrors in toilets, dust obtained from vacuum cleaners, under the surfaces of shoes, books and shelves in libraries and upper surfaces of water taps.
- 2) Determine of the number of bacteria on hands after washing and drying normally and with warm air dryer and quantify of bacteria transferred from hand warm air dryers
- 3) Determine the factors which influence the survival of bacteria and to a lesser extent *Candida* in the built environment on ceramic tiles, on copper and plastic plumbing surfaces, and on new toothbrushes.
- 4) Isolation pathogenic bacteria from used toothbrushes and determine the effect of toothpastes on growth of pathogenic bacteria.
- 5) Determine of effectiveness of antibacterial cloths in inhibiting the growth of bacteria and yeast.
- 6) Determine the survival and relevant metabolic diversity of bacterial isolates.

CHAPTER 2

THE DISTRIBUTION OF BACTERIA IN THE ENVIRONMENT AND THEIR SURVIVAL IN HEALTHCARE SETTINGS

2.1. Isolation of bacteria from sinks, toilets and other medical environments

Hospitals and other health care settings act as an obvious potential reservoir for pathogens (Dancer, 2009). (Bauer *et al.*, 1990, Kayabas *et al.*, 2008, Medina *et al.*, 1997, Sehulster *et al.*, 2003). Contaminated hand washing sinks are an obvious source of infection, especially where they are used for disposing of body fluids, where they become a focus for the survival of pathogens and especially biofilms; surprisingly perhaps, healthcare workers can become contaminated when washing their hands in a contaminated sink (Roux *et. al.*, 2013).

2.2. Isolation of bacteria from sinks

2.2.1. Materials and Methods

1) Isolation of samples and collection

Sterile cotton swabs wetted by dipping in normal saline were used to collect samples from sinks. All samples were labelled and streaked on Nutrient Agar (Oxoid) plates, followed by incubation at 25°C under aerobic conditions for 24hrs. After incubation the colonies were identified. The sinks examined were located as follows: Sheffield University: Firth Court Building, Disability and dyslexia support service, The Alfred Denny Building, Information Commons Building,

Students Union Building. Other areas in Sheffield: Sheffield Train Station, local supermarket, local hospital, various private dwellings.

2) Purification of isolates

All samples were isolated from sinks and then streaked on media in Petri dishes. The main medium used was Nutrient Agar, incubation was then at 37°C for 48 hours. The isolates were purified to single colonies and subjected to molecular identification.

3) Identification of bacterial isolates using 16s rRNA technique

A bacterial suspension in LB medium was prepared and incubated overnight at 37°C. After the incubation period, 1-3 ml of media was transferred to a sterile Eppendorf tube and centrifuged at 6000xg for 2 min at room temperature and the supernatant was decanted completely. A KeyPrep bacterial DNA extraction kit supplied by ANACHEM® was used and all steps were conducted as described in the instructions provided by the Company. Buffer (100µl) R1 was added to the pellet and the cells were re-suspended completely by pipetting up and down. After full cell homogenising, 20 µl of lysosyme was added, mixed and incubated at 37°C for 20 min. The mixture was then digested by centrifugation at 10,000xg for 3 min and the supernatant was completely decanted. The pellet was then re-suspended in 180 µl of buffer R2 and 20 µl of proteinase K was added and incubated at 65°C for 20 min in a water bath with occasional mixing every 5 min.

400 µl of buffer. BG was added and mixed thoroughly by inverting the tube several times until a homogeneous solution was obtained and then incubated for 10 min at 65⁰ C. After the incubation period, 200 µl of absolute ethanol was added and mixed thoroughly.

The sample was next transferred into a column assembled in a clean collection tube and centrifuged at 10,000 xg for 1 min while the flow was discarded. The column was washed by addition of 750 µl of wash buffer and centrifuged at 10,000 xg for 1 min while the flow was discarded. Finally, the column was placed in a clean micro-centrifuge tube and 70µl of elution buffer was added and centrifuged at 10,000xg for 2 min to elute DNA. DNA was stored at -20⁰C until the next step.

4) Gel electrophoresis

Gel electrophoresis was conducted to ensure that the bacterial DNA was well-extracted and purified. The following steps were used.

4.1 Agarose preparation

Powdered agarose was weighed carefully (0.5 g) into a conical flask and 50 ml of 1x TAE (Tris Acetate EDTA) buffer was added with 40ml of distilled water. The contents of the flask were then mixed and placed in a microwave plate until the contents just began to boil and all the powdered agarose is melted. After the contents cooled to 50-55 °C, ethidium bromide solution was added to give a final concentration of 5 pg/mL. The mixture was then be poured into a gel tray and left

to solidify. The gel was finally placed in an electrophoresis tank and submerged in 1x TAE buffer to ensure that the sample diffused into the wells.

4.2 Sample loading

DNA samples (10 µl) were prepared by the addition of 5X (2µl) loading dye (blue or orange) to the samples and loaded into the wells of the gel. All samples were loaded at the same time. 6 µl of hyper ladder was then added into an adjacent well as a reference. The voltage was set to the desired level at 80V for 40 minutes to initiate electrophoresis. The leads were then attached allowing DNA to migrate within the gel toward the anode. After electrophoresis the gel was removed from tray, and the DNA fragment was viewed on a UV transilluminator; images were then captured using a connected digital camera.

4.3. Sample amplification

The Polymerase Chain Reaction (PCR) technique was used. A mixture in a sterile Eppendorf tube was prepared as follows: 12 µl of master mix, 1 µl of forward primer, 1 µl of reverse primer, 1 µl of the DNA sample, and 35 µl of sterile distilled water. The mixture was inserted in a PCR machine and the programme was adjusted as follows:

Table 2.1 The Polymerase Chain Reaction (PCR) cycle.

Steps	Temperature	Time (Min)	Number of cycles
Initialization (Initial denature)	94°C	3	1
Denature	94°C	1	35
Annealing	60°C	1	35
Extension/Elongation	72°C	1	35
Final elongation	72°C	5	1
Hold	4°C		1

5) 16S rRNA sequencing and phylogenetic analysis

After PCR, aliquots of 10 µl of each sample were allowed with 1µl of forward primer and 1µl of reverse primer in a sterile small size tube and sent to the Medical School Core Genetics Unit (University of Sheffield) to be sequenced. 16S rRNA gene sequences were adapted using the Finch TV software and then exported into the Basic Local Alignment Search Tool (BLAST), available from the website of the National Centre for Biotechnology Information (NCBI), to identify matches with existing characterized reference sequences. Partial sequences, generated in this experiment, were assembled and the errors of

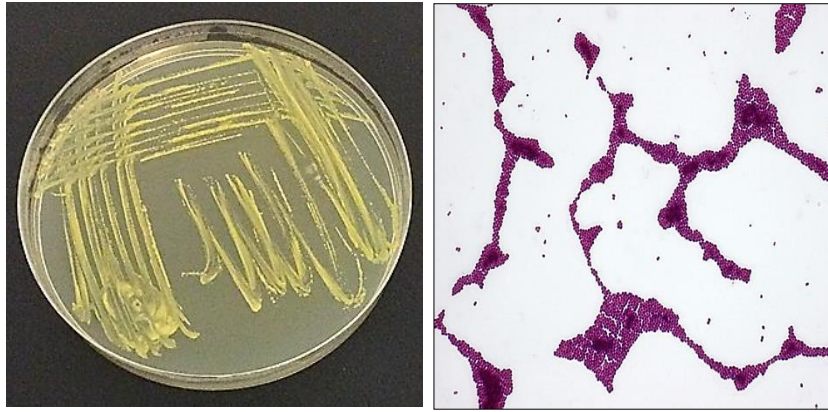
consensus sequences were corrected manually by using Finch TV software (version 1.4). In Finch TV software, the unknown nucleotide is represented as N, and it could be either A, or T, or G, or C, according to the different colours which appear (Mishra *et al.*, 2010).

2.2.2. Results

1) Isolation of bacteria from various sinks by cultivation on Nutrient Agar medium

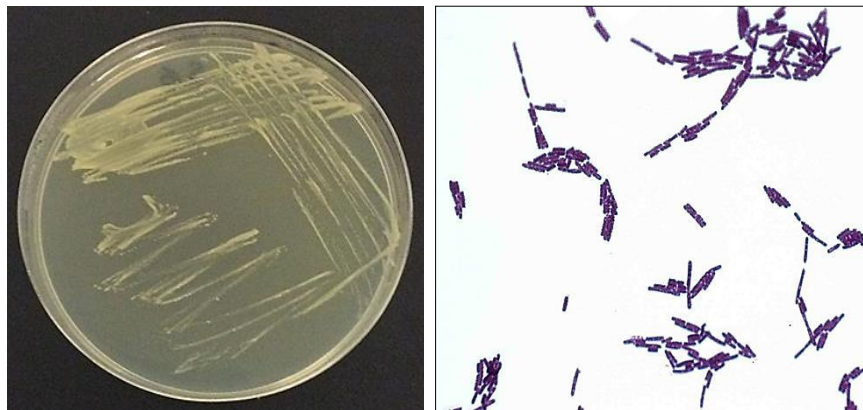
Bacteria were isolated using Nutrient Agar which facilitates the rapid isolation of bacteria from mixed cultures and used for biochemical or serological tests. This medium is a basic culture medium used for maintaining microorganisms, cultivating fastidious organisms by enriching with serum or blood and are also used for purity checking prior to biochemical or serological testing. The presence of peptic digest of animal tissue, beef extract and yeast extract provides the necessary nitrogen compounds, carbon, vitamins and also some trace ingredients necessary for the growth of bacteria; sodium chloride maintains the osmotic equilibrium of the medium. The following bacteria were isolated from the surface of ceramic sinks.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.1) in order to confirm that they were bacteria



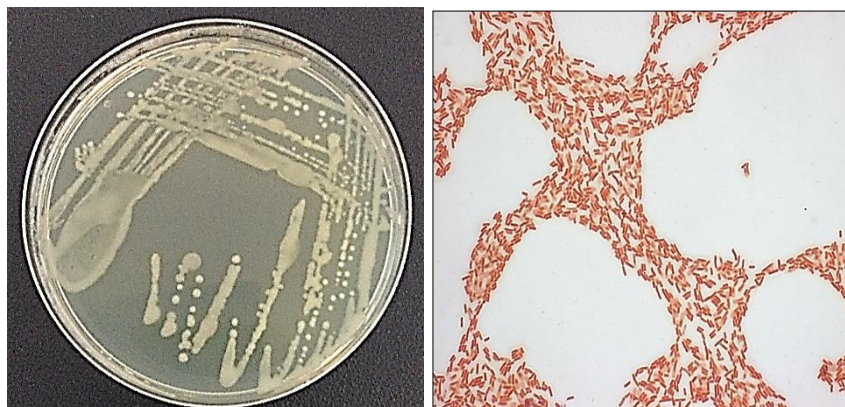
Kocuria rhizophila

Gram stained. Magnification: 100x.



Micrococcaceae Bacterium

Gram stained. Magnification: 100x.



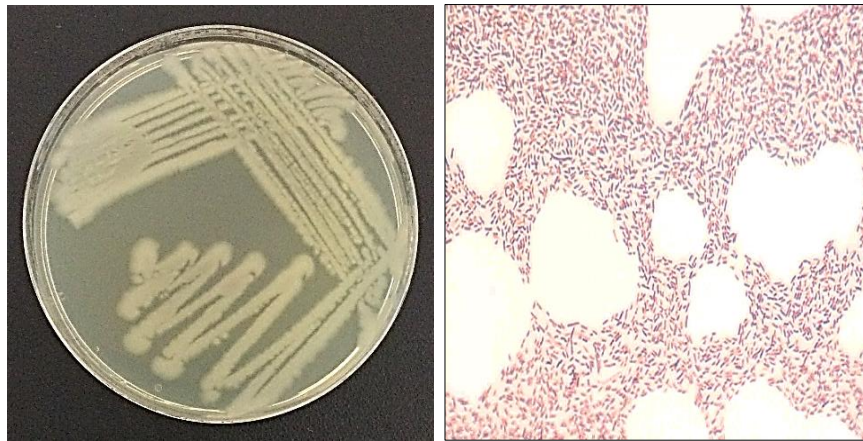
Klebsiella oxytoca

Gram stained. Magnification: 100x.



Bacillus subtilis

Gram stained. Magnification: 100x.



Bacillus cereus

Gram stained. Magnification: 100x.

Figure 2.1: Nutrient Agar plates showing cultured bacteria isolated from various sinks, also microscopy images shows the bacteria under the light microscope.

Table 2.2: Bacteria isolated from various sinks.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
S1	<i>Klebsiella oxytoca</i>	100%	CP011618.1
S2	<i>Bacillus subtilis</i>	100%	KP340123.1
S4	<i>Kocuria rhizophila</i>	99%	KM978822.1
S5	<i>Bacillus cereus</i>	99%	KC731425.1
S6	<i>Micrococcus luteus</i>	99%	KT339390.1

2.2.3. Discussion

The bacteria shown in Table 2.2 were isolated from chosen sinks which are in general use and so represent the environments typical of health care settings. Five bacteria were isolated and identified, two of which were species of *Bacillus*.

Klebsiella oxytoca

Klebsiella is commonly isolated in the clinical laboratory from specimens of blood, urine and respiratory secretions. *Klebsiella* are non-motile, Gram-negative bacilli, glucose fermenters, and form red colonies on MacConkey agar; negative results for cytochrome oxidase activity, hydrogen sulfide production in triple sugar iron agar, arginine and ornithine, decarboxylation, phenylalanine deamination, and citrate utilization. *Klebsiella oxytoca* is a diazotroph, which colonises plants and

fixes atmospheric nitrogen in, for example, the barley rhizosphere. This organism causes colitis and sepsis (Hogenauer *et al.*, 2006).

Bacillus cereus

B. cereus group are Gram-positive, rod-shaped, spore formers, motile. *Bacillus cereus* is the most frequently isolated of *Bacillus* spp involved in nosocomial infections. It has been described an outbreak of invasive *B. cereus* infections in neonatal intensive care units. Kalpoe *et al* (2008) reported an outbreak of vancomycin resistant *B. cereus* respiratory tract colonization in patients in pediatric intensive care units. *Bacillus cereus* is also common in bloodstream infections in patients with hematologic malignancies due to contamination of re-used towels. *Bacillus cereus* is a highly important eye pathogen because it can cause blindness rapidly; endophthalmitis associated with this bacterium often results in significant vision impairment. This bacterium is the most known food-borne pathogen associated with food-poisoning. It has also been found to be associated with fulminant pneumonia in immunocompetent patients and with oral diseases; meningoencephalitis and meningitis in infants, neonates and immunocompromised individuals; necrotizing fasciitis and myositis (Liu, 2011).

Bacillus subtilis

Bacillus species, including *B. subtilis*, can form endospores which allow them to survive heat treatment and disinfection procedures and as a result, present serious health problems as contaminants of food or in the general hospital settings as a cause of nosocomial infections, especially in immunocompromised patients. These bacteria produce toxins which have been linked as the causative agent of several human diseases including endophthalmitis, inflammation of intraocular tissues or fluids due to intraocular infection (Liu, 2011). They also contribute to foodborne infections by producing toxins with a cereulide-like mode of action (e.g. in oral disease, mainly gingivitis and periodontitis). Cryptic pyogenic infections of the central nervous system (CNS) following dental affections have also been implicated to *B. subtilis* and *B. circulans* (Liu, 2011).

Kocuria rhizophila

Kocuria rhizophila cells are Gram- positive, non-acid-fast, non-motile, non-endospore forming, aerobic, and occur in pairs, tetrads, and packets. *Kocuria rhizophila* is widely distributed in the natural environments such as soil, the rhizosphere, freshwater, marine sediments, and fermented foods. *Kocuria* spp are also found in the skin, mucosae, and oropharynx of humans as commensals, although several of them have been associated with human diseases (Liu, 2011). Becker *et al.*, (2008) reported a case of *Kocuria rhizophila* infection in a boy with methylmalonic aciduria. The patient developed sepsis and showed symptoms of

acute pancreatitis and fever. A *Kocuria rhizophila* strain was also isolated from blood samples drawn through a port system and from peripheral veins during septic episodes. A number of *Kocuria* species are opportunistic human pathogens associated with catheter-related bacteraemia, pneumonia, intracranial abscesses, septic arthritis, meningitis, peritonitis, and endocarditis in immunocompromised patients (Liu, 2011).

Micrococcus luteus

Members of the genus *Micrococcus* are Gram-positive cocci, aerobic, non-motile, and non-endospore forming. *Micrococcus* spp. are commonly found as members of the normal skin flora of humans and other mammals. *Micrococcus* strains such as *M. luteus* are opportunistic human pathogens associated with catheter-related bacteraemia in patients undergoing haemodialysis or leukaemia treatment, pneumonia, endocarditis patients particularly immunocompromised individuals, intracranial abscesses, continuous ambulatory dialysis peritonitis, septic arthritis, and meningitis (Liu, 2011).

The results show that sinks are clearly a potential source of pathogens, both in the domestic and healthcare settings.

Hospital sinks are considered to be one of the most frequently implicated reservoirs for MDR Gram-negative bacilli, including MDR coliforms (Roux *et al.*, 2013). *K. pneumoniae* strains which have the ability to survive for long periods

within plumbing components are also more likely to contain bacteria exhibiting extended-spectrum β -lactamases (Yang and Zhang, 2008). Removal and replacement of the sink and associated pipe work together with upgrading practices for sink use and cleaning is often required to end an outbreak. Outbreaks of MDR *Klebsiella* are also often associated with the bad practise of tipping patient fluids down the nearest available sink instead of removing clinical waste to the designated waste sluice located at a distance (Roux *et al.*, 2013). Not surprisingly, lower rates of sink contamination are significantly linked to daily bleach disinfection, in addition to restrictions being placed on sink use for hand washing only and not the routine disposal of fluid or clinical waste (Roux *et al.*, 2013, Yang and Zhang, 2008). It is noteworthy in the finding reported in this Thesis that the bacteria isolated from sinks were essentially environmental species and not those capable of causing major life-threatening infections. It needs to be emphasised, however, that even these bacteria can cause major problems in immunocompromised patients.

2.3 Isolation of bacteria from lift buttons surfaces

2.3.1 Methods

Sterile cotton swabs wetted by dipping in normal saline were used to collect samples from environmental surfaces such as lifts buttons from different floors in the MBB building and Sheffield University Students' Union. All samples were labelled and streaked on to Nutrient Agar Plates which were incubated at 25°C

under aerobic conditions for 24hrs. After incubation, the colonies were identified. Genomic DNA was extracted from each isolate and was identified using 16S rRNA. The extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit (supplied by ANACHEM). The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.3.2 Results and Discussion

1) Isolation of bacteria from various lift buttons by cultivation on Nutrient Agar medium

Bacteria were cultured using Nutrient Agar having been isolated from lift buttons. The bacteria shown in (Figure 2.2)

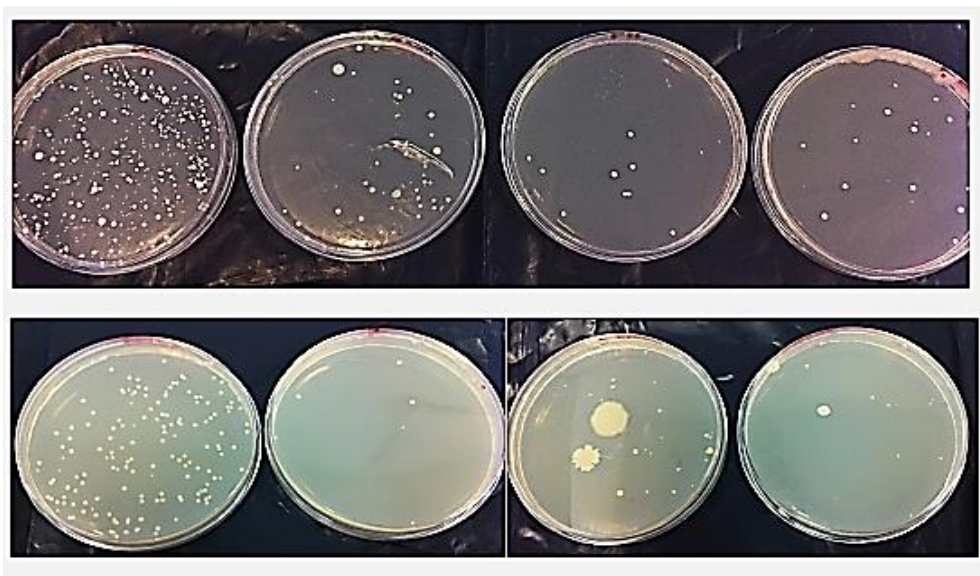


Figure 2.2: Bacteria contaminating lift buttons. From left to right, shows ground floor to upper floors from various locations.

Table 2.3: Bacteria isolated bacteria from lift buttons.

Representative sequence	Closest matches identification	Sequence identity	NCBI (Accession number)
6W3	<i>Staphylococcus warneri</i> BCL-34	99%	KT582294.1
7W1	<i>Staphylococcus epidermidis</i>	98%	FR797804.1
7Y3	<i>Micrococcus luteus</i> MBS022	99%	KM378607.1

Staphylococcus

The characteristics of these bacteria include:

Staphylococcus is a genus of catalase positive Gram-positive bacteria.

Staphylococcus bacteria appear as round (cocci) and come together as grape-like clusters. *Staphylococci* consist of >40 species and are noted for their ability to cause disease, with *Staphylococcus aureus* being a most common pathogen of man and animals. Most *Staphylococcus* species are however, harmless and normally live on the skin and mucous membranes of humans and other organisms (Liu, 2011).

Staphylococcus aureus is mainly a commensal organism residing on the human epithelia of the mucosae and skin. It has its primary ecological site in the vestibulum nasi, located in the forefront region of the nose. None-nasal sites typically harbouring the organism include the skin, perineum, and pharynx; other

sites include the gastrointestinal tract, vaginas, and axillae. This bacterium causes superficial skin lesions like impetigo, styes and furuncles, cellulitis folliculitis, carbuncles, staphylococcal scalded skin syndrome (SSSS), and abscesses (Liu, 2011). More serious infections include pneumonia, mastitis, phlebitis, meningitis, urinary tract infections (UTIs), and septicaemia. *Staphylococcus aureus* infections including osteomyelitis, hospital-acquired (nosocomial) infections in surgical wounds and infections associated with indwelling medical devices. *S. aureus* also causes food poisoning by producing enterotoxins and toxic shock syndrome (TSS) by release of superantigens into the blood stream (Liu, 2011).

Staphylococcus epidermidis is a skin commensal which can be isolated from mucous membranes, such as the groin or axilla, exposed skin surfaces and saliva. It is the main human pathogen in intravascular catheter-related infections, nosocomial bacteraemia, endocarditis, urinary tract and surgical wounds infections and infections of the central nervous system, ophthalmologic infections, peritoneal dialysis-related infections and infections of prosthetic joints (Liu, 2011). Some isolates produce slime or biofilm which is the major virulence factor of *S. epidermidis*, enabling colonization and persistence on prosthetic material, the resistance to the effects of antibiotics, and the ability to evade the immune system (Liu, 2011).

Micrococcus are Gram-positive, non-motile, non-endospore forming, aerobic cocci. *Micrococcus luteus* have been associated with catheter-related bacteraemia in patients undergoing haemodialysis or leukaemia treatment, pneumonia,

endocarditis, intracranial abscesses, continuous ambulatory dialysis peritonitis, septic arthritis, and meningitis (Liu, 2011).

The number observation of bacterial contamination of lift buttons decreased from ground floor towards upper floors as shown in Figure 2.2. These results agree with the findings of Al-Ghamdi *et al.* (2011) who showed that the average rate of bacterial contamination of different objects within the hospital environment was 95.5% with elevator buttons showing the highest percentage (97%). Scott and Bloomfield (1990) suggested that when contaminated surfaces are contacted with the fingers, a significant number of organisms can be transferred which can be subsequently recovered on an agar surface. Inanimate objects can play a role in the transmission of human pathogens. These surfaces have been shown to carry both non-pathogenic and pathogenic bacteria and even a single hand contact with a contaminated surface results in a variable degree of pathogen transfer.

2.4 Isolation of bacteria from computer keyboards and computer mice

Multidrug-resistant strains of *S. aureus*, particularly methicillin resistant *S. aureus* (MRSA), present a major clinical and epidemiological problem in healthcare settings as they are easily transferred among hospital staff and patients, especially in intensive care units (ICUs). Hails *et al.* (2003) found that at least 16% of patients were colonised with MRSA. A significant factor contributing to the transmission of microorganisms is their ability to survive on environmental

surfaces. Also the microbial contamination of environmental surfaces in the hospital may contribute to the spread of potential pathogens without direct patient contact. Coagulase-negative Staphylococci have been cultured from all keyboards investigated in different studies conducted in the USA (Anastasiades *et al.*, 2009). In another study which focused on the isolation of MRSA, 65% of nurses, who had contact with MRSA-infected patients, bacteria contaminated their uniforms as well as the keyboards and computer mice (CM) in the hospital wards. These results confirmed that inanimate objects can serve as reservoirs for bacteria. Hartmann *et al.* (2004) also found that keyboards and CM might serve as a source for the transmission of microorganisms.

In a study investigating the presence of both Gram-positive and Gram-negative bacteria on computer keyboards, coagulase-negative staphylococci and *S. aureus* were isolated from 100% and 4% of keyboards, respectively (Rutala *et al.*, 2006). It was suggested that the use of plastic keyboard covers could reduce contamination. It was then recommended that the same infection prevention measures employed during direct contact with patients (i.e. hand washing and use of gloves), should be enforced when handling computer hardware in healthcare settings.

As has been mentioned in Chapter 1, medical surfaces present a major source of outbreaks of community-acquired and nosocomial infections. Such surfaces include frequently used computer keyboards and accompanying mice which

harbour skin and dust bacteria, such as species of *Staphylococcus* and *Streptococcus epidermidis* and fungi (yeasts), notably *Candida albicans*.

Keyboards were also implicated in burns unit-associated nosocomial infections by *A. baumannii* (Neely *et al.*, 1999) MRSA and *Enterobacter* spp. (Bures *et al.*, 2000; Goldman, 2000). It goes without saying that hospital keyboards and computer mice should be routinely cleaned and covered with transparent plastic covers when not in use. Hand hygiene is also necessary to avoid cross contamination of keyboards and other computing devices. This is especially important since contaminated personal computers are often contaminated with *Staphylococci* and *Pseudomonas* spp. And can transmit MRSA (Isaacs *et al.*, 1998).

2.4.1 Materials and Methods

All samples are isolated from computer keyboards using swabs (70 samples) streaked on petri dishes containing Nutrient Agar and incubated at 25°C for 48 hours. Genomic DNA extracted from each isolate was identified using 16S rRNA. The extraction of genomic DNA by using (KeyPrep bacterial DNA extraction kit supplied by ANACHEM), The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.4.2 Results

1) Isolation of bacteria from computer keyboard

Bacteria were cultured using Nutrient Agar after isolation from computer keyboards and mice.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.3).



Bacillus amyloliquefaciens

Gram stained. Magnification: 100x.



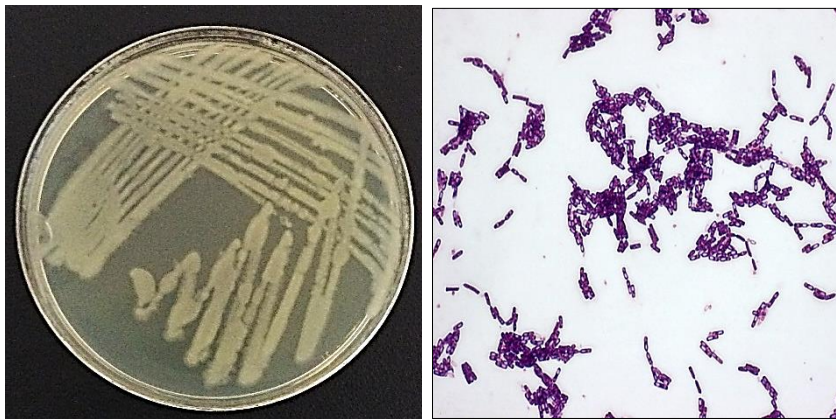
Bacillus subtilis

Gram stained. Magnification: 100x.



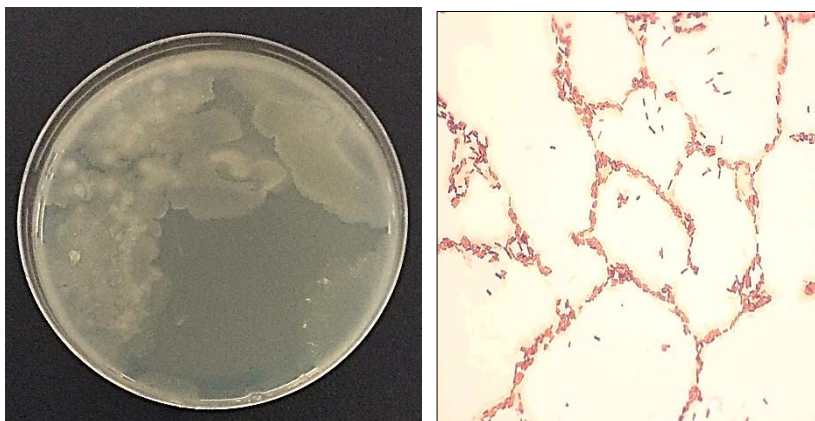
Brevibacillus brostelensis

Gram stained. Magnification: 100x.



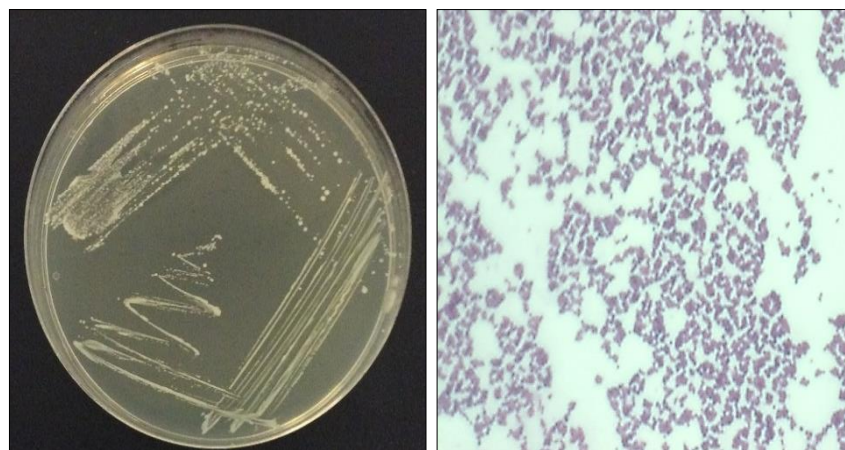
Bacillus cereus

Gram stained. Magnification: 100x.



Pantoea caldia

Gram stained. Magnification: 100x.



Staphylococcus epidermidis

Gram stained. Magnification: 100x.

Figure 2.3: Nutrient Agar plates showing cultured bacteria isolated from various computer keyboards, also microscopy images showing the bacteria under the light microscope.

Table 2.4 Bacteria isolated from various computer keyboards.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
KB1	<i>Bacillus subtilis</i>	99%	KJ746466.1
KB4	<i>Bacillus amyloliquefaciens</i>	99%	AB301004.1
KB6	<i>Brevibacillus brostelensis</i>	98%	EU816699.1
KB11	<i>Staphylococcus epidermidis</i>	99%	KF575163.1
KB13	<i>Pantoea caldia</i>	99%	AB907785.1
KB14	<i>Bacillus cereus</i>	99%	DQ923480.1
KB21	<i>Pseudomonas luteola</i>	99%	KC429633.1

Bacillus amyloliquefaciens

This bacterium typically presents as motile, Gram-positive rods, often forming chains, with peritrichous flagella. Optimal temperature for growth is 30 to 40°C, while no growth occurs below 15°C or above 50°C. *B. amyloliquefaciens* can be isolated from soil and brackish sediments (Priest *et al.*, 1987). *B.*

amyloliquefaciens is not known to produce any mammalian toxins, and is not associated with food-borne disease. No problems have been reported, and none are expected from exposure to *B. amyloliquefaciens* via drinking water (Priest *et al.*, 1987).

Brevibacillus brostelensis

Brevibacillus brostelensis cells present as Gram-positive rods, motile with peritrichous flagella. It is a thermophilic, spore-forming rod with a growth optimum at 50°C. Colonies are flat, smooth, circular, and entire. This bacterium produces a soluble brown-red pigment(s) on nutrient agar and is strictly aerobic. The habitat of *Brevibacillus* overlaps with that of *Bacillus* and is associated with soils and dairy environments (Sanders, 2003; Shida *et al.*, 1995).

Pseudomonas luteola

Pseudomonads appear as straight or slightly curved Gram-negative rods. They are usually easily differentiated from Enterobacteriaceae by a lack of a bipolar

staining. Isolates often seen as clusters of rods encapsulated in a thick pink staining alginate coat. *Pseudomonas* spp. grow on several different nonselective agars, including nutrient, 5% blood, chocolate, and MacConkey agars (Liu, 2011). Most species are aerobic and grow at 37°C, and within 24h, form visible colonies. This bacterium is oxidase negative and produces a non-diffusible yellow pigment. *Pseudomonas* spp. act as opportunistic pathogens and *P. luteola* can cause community- or hospital acquired infection in humans especially patients in an intensive care unit. *Pseudomonas luteola* causes dialysis associated peritonitis; cellulitis; sepsis; prosthetic valve endocarditis; and postoperative endophthalmitis (Liu, 2011).

Pantoea calida

Pantoea calida cells are Gram-negative rods that are facultatively anaerobic and motile. Colonies are non-pigmented and convex, oxidase-negative and catalase-positive (Fritz *et al.*, 2014). *Pantoea calida*, a recently described environmental Enterobacteriaceae organism, has yet to be associated with human infection, although Fritz *et al.*, (2014) isolated this organism from a patient with postoperative meningitis.

2.5. Isolation of bacteria from mobile telephones

2.5.1 Materials and Methods

Isolates were obtained from a variety of mobile telephones (30 samples) using nutrient agar medium. Incubation was at 37°C for 48 hours and the isolates were identified using 16S rRNA. Extraction of genomic DNA was by using Norgen's Fungi/Yeast Genomic DNA Isolation Kit. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.5.2 Results

1) Isolation of bacteria from mobile phones

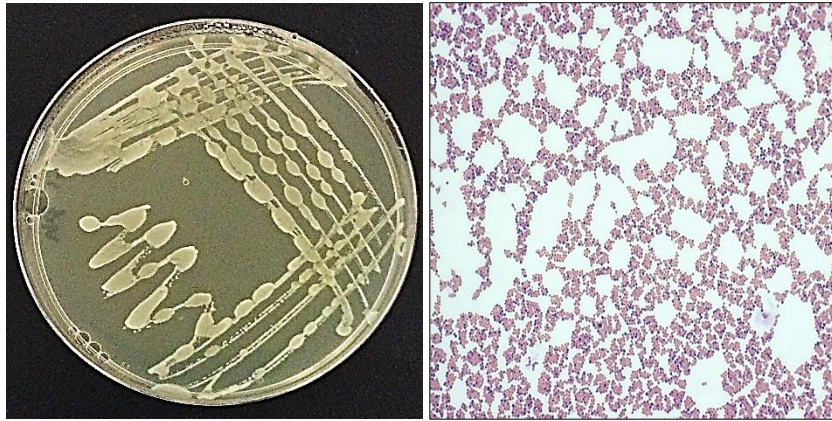
Bacteria were cultured using Nutrient Agar after isolation from mobile phones.

2) **Light microscope images.** The isolates were examined under a light microscope (Figure 2.4).



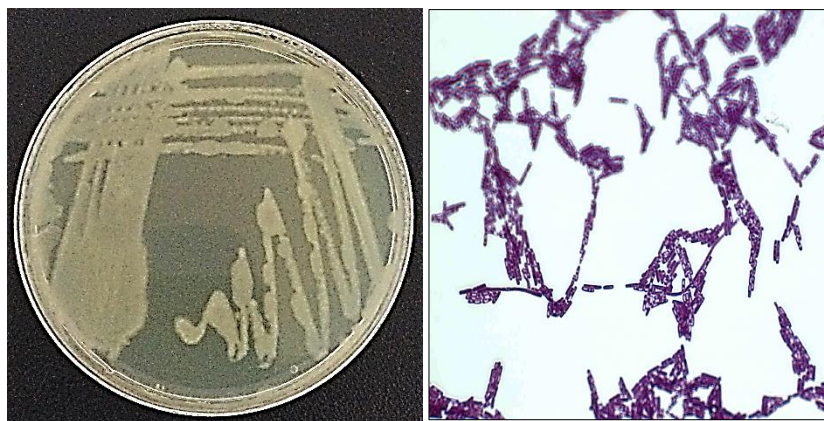
Staphylococcus epidermidis

Gram stained. Magnification: 100x.



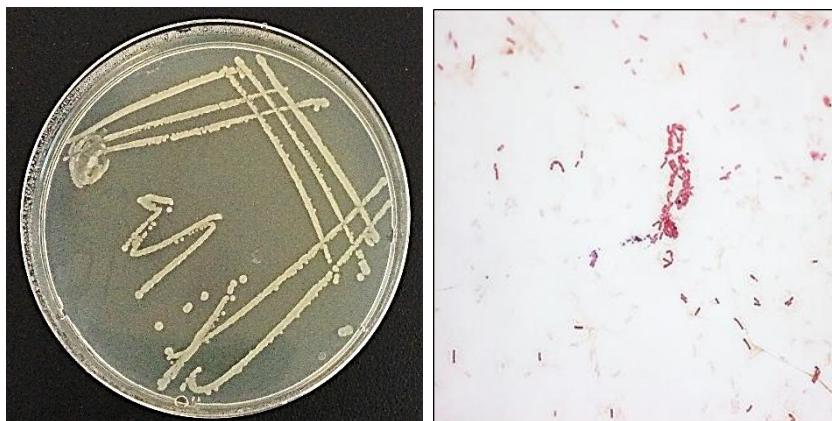
Staphylococcus warneri

Gram stained. Magnification: 100x.



Bacillus cereus

Gram stained. Magnification: 100x.



Bacillus subtilis

Gram stained. Magnification: 100x.

Figure 2.4: Nutrient Agar plates showing cultured bacteria isolated from various mobile phones, and also microscopy images showing the bacteria under the light microscope.

2) Table 2.5: Bacteria isolated from various mobile phones.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
3P	<i>Staphylococcus epidermidis</i>	99%	KJ398217.1
15P1	<i>Staphylococcus warneri</i>	99%	KP771665.1
15P2	<i>Bacillus subtilis</i>	99%	HG764646.1
18P	<i>Bacillus cereus</i>	99%	KJ612539.1

Staphylococcus warneri a Gram-positive member of the microbiota, normally found on the skin of humans and animals. *S. warneri* is a significant opportunistic nosocomial pathogen causing complications to the use of central venous catheters, prosthetic heart valves and joints, and neurosurgical ventricular shunts (Incani *et al.*, 2010). *Staphylococcus warneri* has been associated with bacteraemia in hospitalized patients; with intravascular catheter infections in immunocompromised patients (Incani *et al.*, 2010).

2.6 Isolation of bacteria from mirrors in toilets

2.6.1 Materials and Methods

Isolates were obtained from a variety of toilet mirror surfaces (30 samples) using Nutrient Agar medium. Incubation was at 25°C for 48 hours and the isolates were

identified using 16S rRNA. Extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.6.2 Results

1) Isolation of bacteria from mirrors

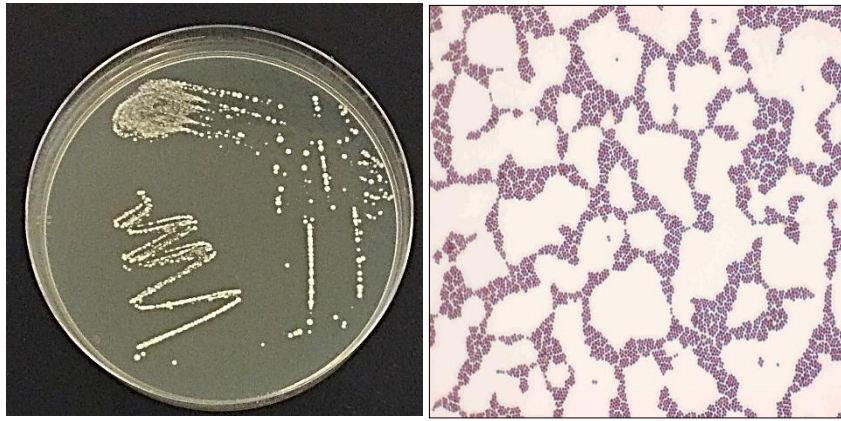
Bacteria were cultured using Nutrient Agar after isolation from mirror surfaces.

2) **Light Microscope images.** The isolates were examined under a light microscope (Figure 2.5).



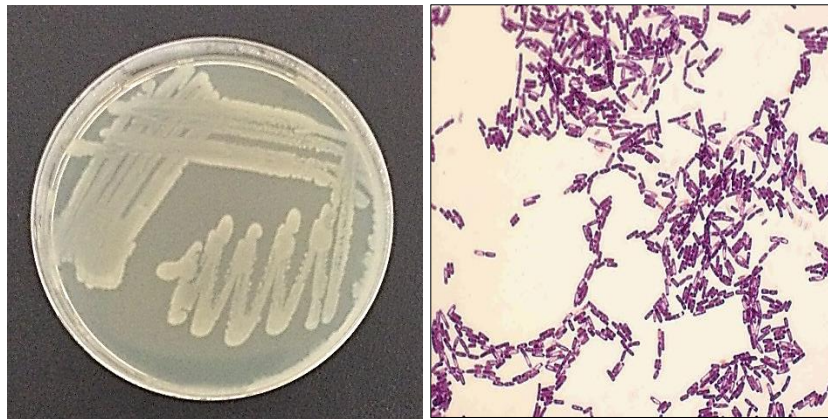
Bacillus cereus KP100400.1

Gram stained. Magnification:
100x.



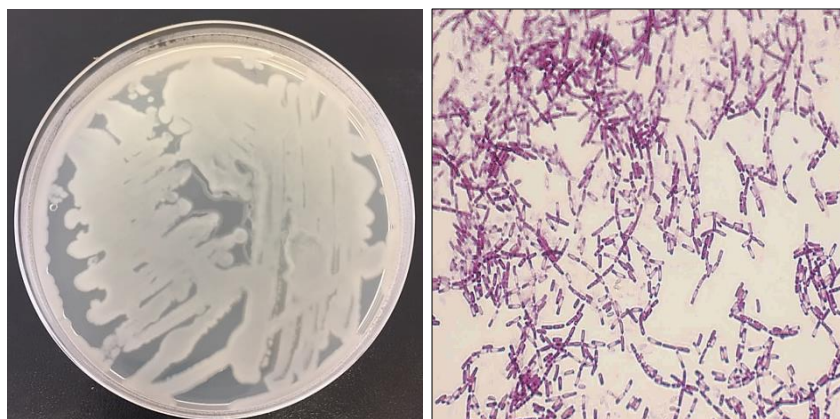
Staphylococcus haemolyticus

Gram stained. Magnification: 100x.



Bacillus cereus Q659737.1

Gram stained. Magnification: 100x.



Bacillus subtilis DQ683077.1

Gram stained. Magnification: 100x.

Figure 2.5: Nutrient Agar plates showing cultured bacteria isolated from various mirror surfaces, and also microscopy images showing the bacteria under the light microscope.

Table 2.6: Bacteria isolated from mirrors.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
M8-8	<i>Bacillus cereus</i>	99%	KP100400.1
M6-1	<i>Bacillus subtilis</i>	99%	DQ683077.1
M15-1	<i>Bacillus cereus</i>	99%	JQ659737.1
M14-1	<i>Staphylococcus haemolyticus</i>	99%	KC139455.1

Staphylococcus haemolyticus is a major cause of human disease in humans and has been implicated in native valve endocarditis (NVE), septicaemia, urinary tract infections, peritonitis, and wound, bone, and joint infections. It forms biofilms which allow it to colonize and persist on prosthetic material where it shows resistance to antibiotics and is able to evade the immune system (Liu, 2011).

2.7 Isolation of bacteria from vacuum cleaner dust samples obtained from carpets, textiles and upholstered furniture

2.7.1 Materials and methods

Dust samples were obtained from a variety of different vacuum cleaners (30 samples) for use in determining the bacterial content of the waste. The dust was placed on the surface of Nutrient Agar medium plates (NA) and then incubated at 25°C for 3 to 5 days. Extraction of genomic DNA was achieved using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

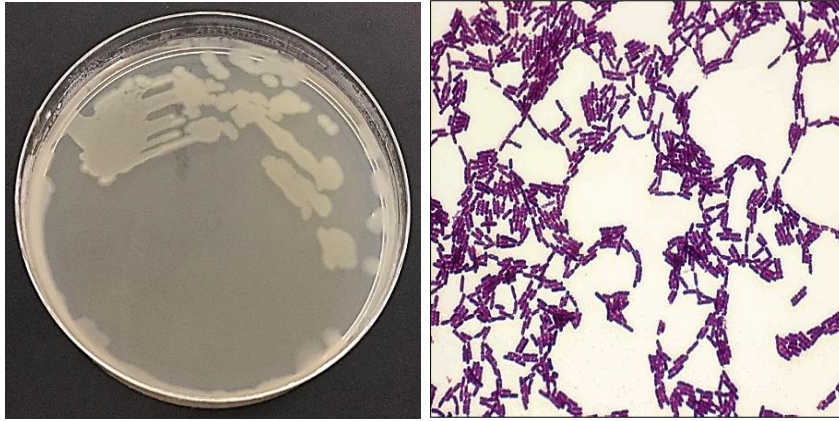
2.7.2 Results and Discussion

1) Isolation of bacteria from dust samples of various vacuum cleaners

Bacteria were cultured using Nutrient Agar which isolated from dust samples.

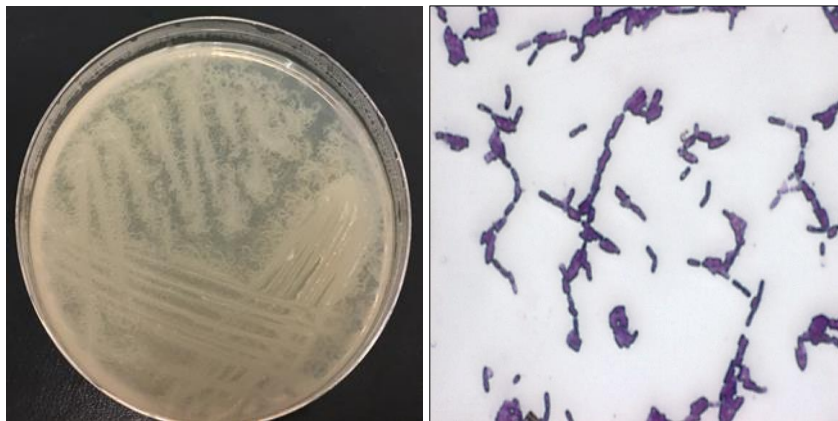
2) Light microscope images

In order to confirm that they were bacteria, the isolates were examined under a light microscope (Figure 2.6).



Bacillus thuringiensis FJ174596.1

Gram stained. Magnification: 100x.



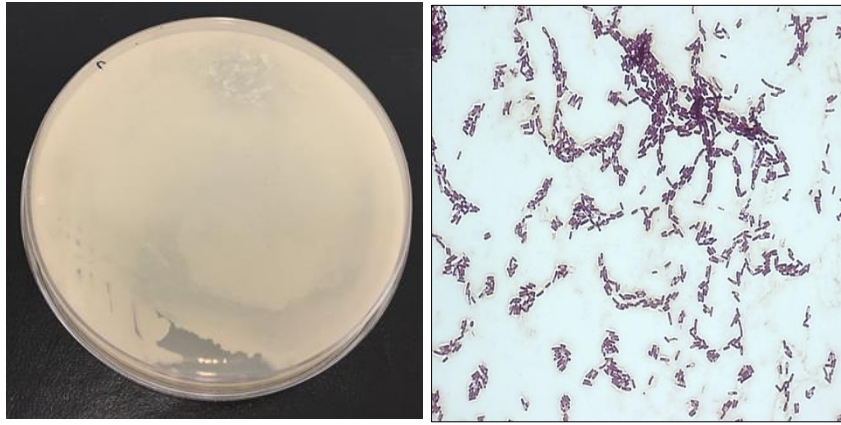
Bacillus mycoides KR088435.1

Gram stained. Magnification: 100x.



Bacillus licheniformis DQ071560.1

Gram stained. Magnification: 100x.



Bacillus subtilis KF220577.1

Gram stained. Magnification: 100x.

Figure 2.6: Nutrient Agar plates showing cultured bacteria isolated from dust samples of various vacuum cleaners, and also microscopy images showing the bacteria under the light microscope.

Table 2.7: Bacteria isolated from dust samples of various vacuum cleaners.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
VC1	<i>Bacillus thuringiensis</i>	98%	FJ174596.1
VC2	<i>Bacillus mycoides</i>	99%	KR088435.1
VC3	<i>Bacillus licheniformis</i>	99%	DQ071560.1
VC4	<i>Bacillus subtilis</i>	98%	KF220577.1

Bacillus licheniformis

B. licheniformis has been shown to be the cause of bacteremic episodes in patients having hemotological malignancies. Only one case has been reported of *B.*

licheniformis nosocomial infection in neonates. In immunocompetent individuals, *B. licheniformis* bacteraemia has been reported in association with central venous catheters and with peritonitis in patients undergoing peritoneal dialysis (Liu, 2011). *B. licheniformis* has also been shown to be involved in foodborne illnesses due to its ability to produce toxins with a cereulide-action mode. Infections caused by this bacterium include, ophthalmitis, septicaemia, bacteraemia, and peritonitis.

Bacillus licheniformis was isolated from patients suffering from eye trauma, lymphoma, leukaemia and metastatic lung cancer (Liu, 2011).

Bacillus thuringiensis

Bacillus thuringiensis is an insect pathogen widely used as a biopesticide. Kuroki *et al.* (2009) demonstrated *B. thuringiensis* species can form biofilms in nosocomial bacteraemia due to catheter infection. There is also a report of fatal *B. thuringiensis* bacteraemia in a neutropenic patient suffering from severe pulmonary disease showing that this entomopathogenic bacterium may be an opportunistic pathogen in immunocompromised patients; it is also the causative agents of infectious endophthalmitis and oral diseases, particularly gingivitis and periodontitis (Kuroki *et al.*, 2009).

Bacillus mycoides

Bacillus mycoides is typically isolated from soil and plant rhizosphere but has also been shown to be a causative agent of infectious endophthalmitis (Liu, 2011).

2.8 Isolation of bacteria from soles of shoes

An obvious potential infection hazard is presented by the transfer of microorganisms from the soles of shoes to floors and other surfaces within health care environments and pathogens can be carried in this way from environmental sources, notably soils and cat and dog faeces. In order to prevent the contamination, shoe-cover-alls should be provided for all persons entering

sensitive medical areas, such as where immunocompromised patients are being treated.

2.8 Isolation of bacteria from the under surface of shoes

2.8.1 Materials and Methods

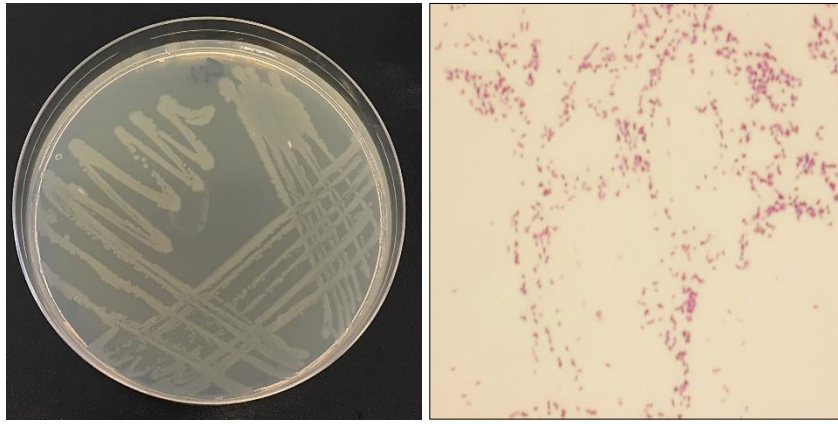
Isolates were obtained from a variety of shoe surfaces (30 samples) using Nutrient Agar medium; incubation was at 37°C for 48 hours and the isolates were identified using 16S rRNA. Extraction of genomic DNA by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.8.2 Results

1) Isolation of bacteria from the under surface of shoes

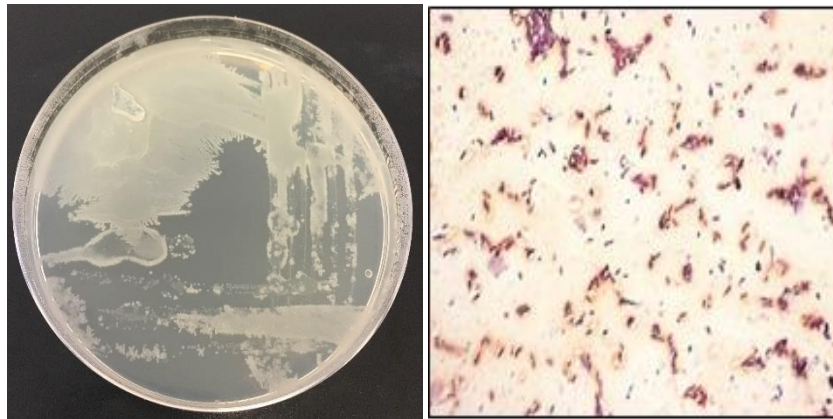
Bacteria were cultured using Nutrient Agar after isolation from the under surfaces of shoes.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.7) in order to establish that they were bacteria.



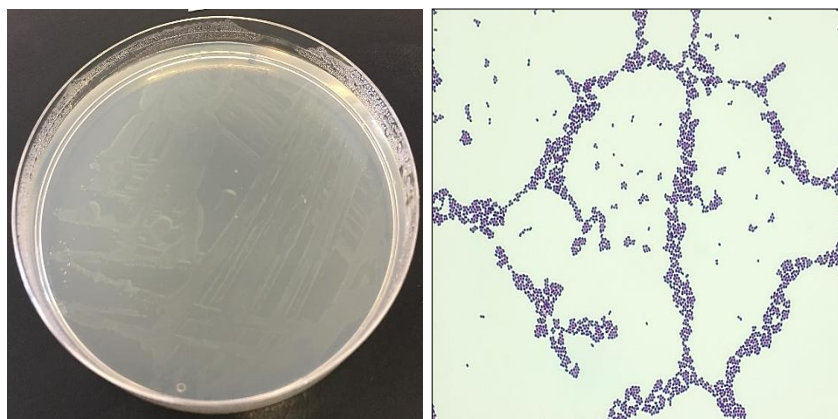
Escherichia coli LN558643.1

Gram stained. Magnification: 100x.



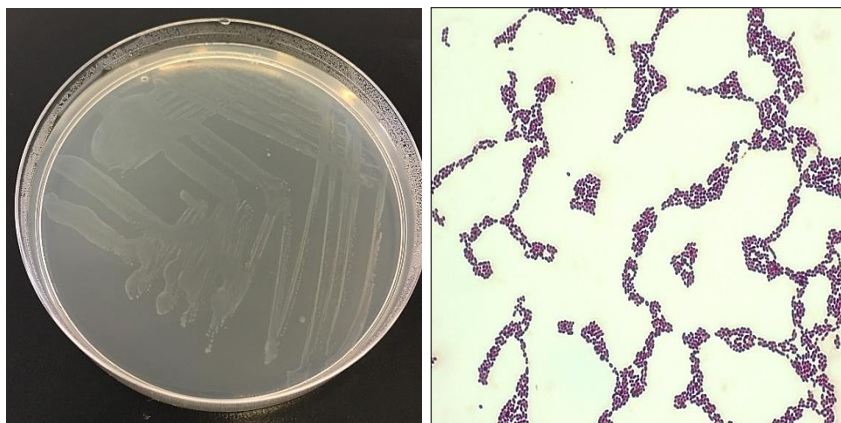
Bacillus licheniformis KP772335.1

Gram stained. Magnification: 100x.



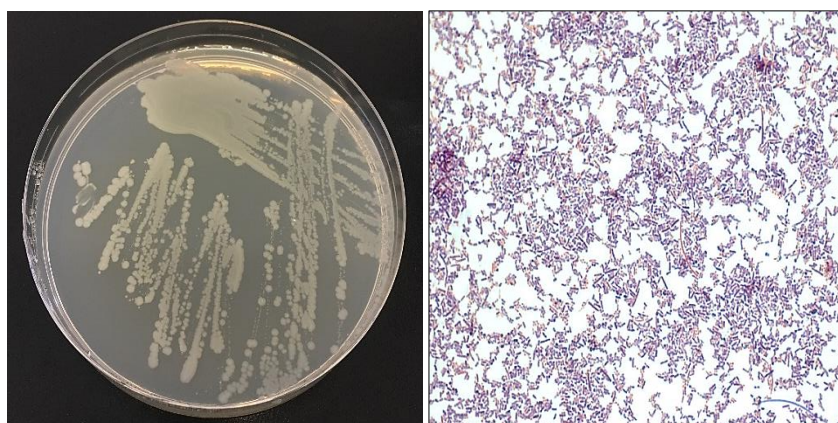
Enterococcus mundtii KR085796.1

Gram stained. Magnification: 100x.



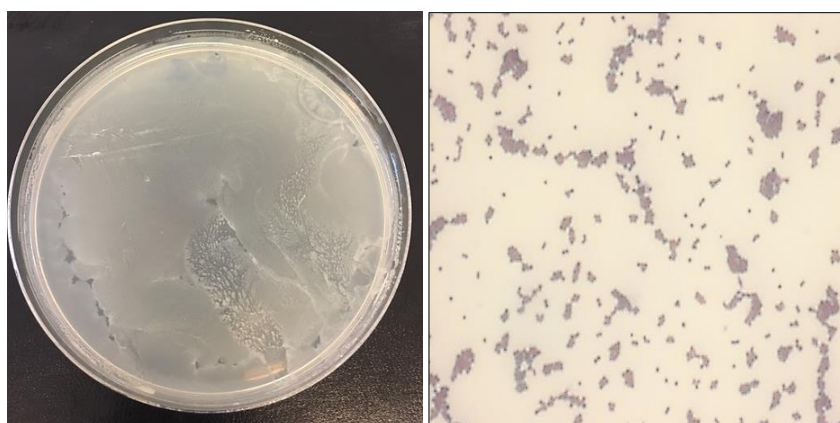
Enterococcus hirae KT261200.1

Gram stained. Magnification: 100x.



Bacillus licheniformis DQ071560.1

Gram stained. Magnification: 100x.



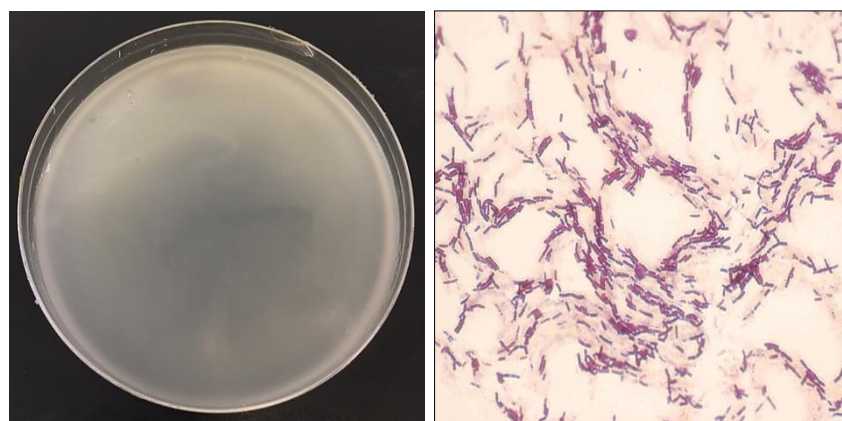
Brevibacillus borstelensis KT239000.1

Gram stained. Magnification: 100x.



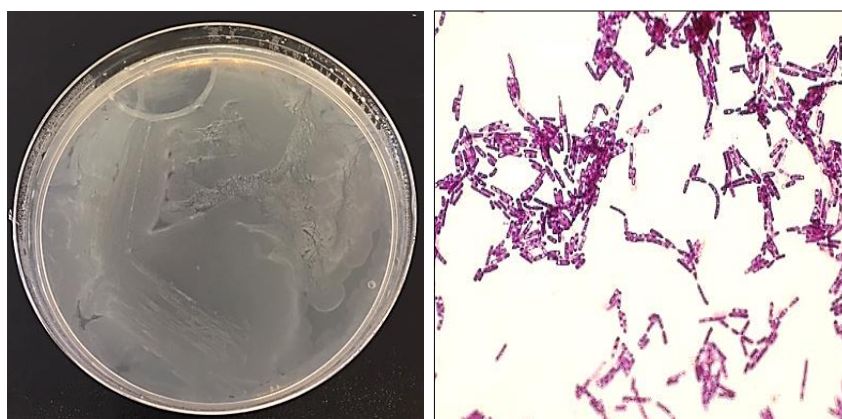
Lysinibacillus fusiformis KP872952.1

Gram stained. Magnification: 100x.



Aneurinibacillus migulanus NR_113764.1

Gram stained. Magnification: 100x.



Bacillus subtilis EF488088.1

Gram stained. Magnification: 100x.

Figure 2.7: Nutrient Agar plates showing cultured bacteria isolated from the under surface of shoes, and also microscopy images showing the bacteria under the light microscope.

Table 2.8: Bacteria isolated from the under surfaces of shoes.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
G1	<i>Escherichia coli</i>	99%	LN558643.1
G2	<i>Brevibacillus borstelensis</i>	98%	KT239000.1
G3	<i>Bacillus licheniformis</i>	99%	KP772335.1
G6	<i>Enterococcus mundtii</i>	99%	KR085796.1
G7	<i>Enterococcus hirae</i>	98%	KT261200.1
G9	<i>Bacillus licheniformis</i>	99%	DQ071560.1
G10	<i>Lysinibacillus fusiformis</i>	99%	KP872952.1
G13	<i>Aneurinibacillus migulanus</i> (<i>Bacillus brevis</i>)	99%	NR_113764.1
G14	<i>Bacillus subtilis</i>	99%	EF488088.1

***Aneurinibacillus migulanus* (*Bacillus brevis*)**

Aneurinibacillus migulanus (formerly *Bacillus brevis*) produces the antibiotic peptide gramicidin S. The cells are rod shaped, motile, and peritrichous, and ellipsoidal spores are formed in swollen sporangia (Berditsch *et al.*, 2007).

Colonies are flat and smooth on nutrient agar. This organism is positive for catalase activity, development of an alkaline pH in Voges-Proskauer broth, and has the ability to reduce of nitrate to nitrite (Berditsch *et al.*, 2007; Takagi *et al.*, 1993). *Bacillus brevis* is a common environmental organism which is not associated with human disease.

Escherichia coli

Escherichia coli is a Gram-negative, facultative anaerobic, rod-shaped, coliform bacterium that is commonly found in the lower intestine of warm blooded organisms. Most *E. coli* strains are harmless, but some serotypes can cause serious food poisoning (Liu, 2011). *E. coli* are part of the normal flora of the gut, and can bring benefit to their hosts by producing vitamin K₂ and by preventing colonization of the intestine with pathogens. This bacterium is expelled into the environment within faecal matter. Faecal transfer is the main route through which pathogenic strains cause disease. Virulent strains can cause gastroenteritis, urinary tract infections, and neonatal meningitis. *E. coli* are a major cause of urinary tract infections (Liu, 2011).

Lysinibacillus fusiformis

The is a Gram-positive rod with motile spore arranged in 2-3 chains. It is frequently isolated from groundwater-derived drinking. Wenzler *et al.*, (2015) report a case of severe sepsis caused by persistent *Lysinibacillus fusiformis* and *Paenibacillus* bacteraemia in a patient with a history of intravenous drug abuse and splenectomy; report a rare case of bacteraemia due to *Lysinibacillus* and *Paenibacillus spp.* in an immunocompetent patient without the involvement of implanted prosthesis or intravascular catheters (Rajesh *et al.*, 2013; Ahmed *et al.*, 2007; Wenzler *et al.*, 2015).

Enterococcus mundtii

Members of the genus *Enterococcus* are Gram-positive ovoid cells that appear singly, in pairs, or in short chains and are catalase- and oxidase-negative; facultative anaerobes; and homofermentative, with lactic acid being the end product of glucose fermentation. Surface colonies on blood agar or nutrient agar are circular, smooth, and entire. Enterococcal infections are a major health concern, and as nosocomial pathogens, they often prolong hospital stays. *Enterococcus mundtii* can cause postoperative endophthalmitis, although it is rarely identified in endogenous endophthalmitis (Higashide *et al.*, 2005).

Enterococcus hirae

Enterococcal disease comprises upper and lower airway, wound, hepatobiliary, intra-abdominal and urinary tract infections, meningitis, infective endocarditis and bacteraemia (including neonatal sepsis) (Poyart *et al.*, 2002). *E. hirae* has been reported to cause wound infections, gastritis, and occasional bacteraemia. It is a zoonotic pathogen rarely isolated from human infections. Poyart *et al.* (2002), however, found *E. hirae* colonized the native aortic-valve endocarditis in a 72-year-old man (Bourafa *et al.*, 2015; Poyart *et al.*, 2002; Savini *et al.*, 2013).

The results show that a wide range of bacteria could be isolated from the soles of shoes worn in normal use. This reality is reflected by the fact that in critical care facilities, visitors and staff are made to wear flexible plastic overshoes to prevent microbial contamination from outside (Falvey and Streifel 2007; Lai, 2001; Mehta, 1990).

2.9 Isolation of bacteria from books and shelves in libraries and archive storerooms

Libraries and archives act as collections for the storage of large amounts of books and documents which are affected by a variety of different environmental factors including: microclimate in store rooms (i.e. temperature and humidity of the air), type and the amount of light (i.e. type of light waves on the electromagnetic

spectrum and radiation intensity), the type and amount of chemicals used for hygiene purposes and the air quality present in storage rooms. Books or other archival materials are a rich reservoir of many nutritional substances, such as cellulose, which is the main constituent of paper, and proteins, which are present in leather books. These nutritional substances stimulate the growth of a specific group of microorganisms (Kalwasińska *et al.*, 2012). Fungi are particularly strong cellulose decomposers, with species being represented, including members of *Botrytis*, *Chaetomium*, *Trichoderma*, *Penicillium*, while those fungi which exhibit strong proteolytic activity include species of the genera *Mucor*, *Aureobasidium*, *Chaetomium*, *Trichoderma*, *Verticillium* and *Epicoccum*. Bacteria however, are only infrequently found on paper, except where it is very damp when species of *Cellulomonas*, *Cellfalciculata*, *Cellvibrio* and *Cytophaga* tend to predominate (Kalwasińska *et al.*, 2012).

2.9.1 Materials and Methods

Sterile cotton swabs were wetted by dipping them in saline and used to collect samples from books and shelves in libraries and archive storerooms (Western Bank Library and the Information Commons, University of Sheffield). All samples were labelled and streaked on to nutrient agar plates; the plates were then incubated at 25°C under aerobic conditions for 24 hrs. After incubation the colonies were identified using 16S rRNA. Extraction of genomic DNA was by

using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.9.2 Results

1) Isolation of bacteria from books and shelves in libraries and archive storerooms

Bacteria were cultured using Nutrient Agar after isolation from books and shelves.

2) **Light microscope images.** The isolates were examined under a light microscope (Figure 2.8).



Pseudomonas jessenii LN774645.1

Gram stained. Magnification: 100x.



Bacillus cereus KP192930.1

Gram stained. Magnification: 100x.



Bacillus altitudinis KT758615.1

Gram stained. Magnification: 100x.



Bacillus pumilus KP322017.1

Gram stained. Magnification: 100x.



Acinetobacter lofwii KT387352.1

Gram stained. Magnification: 100x.



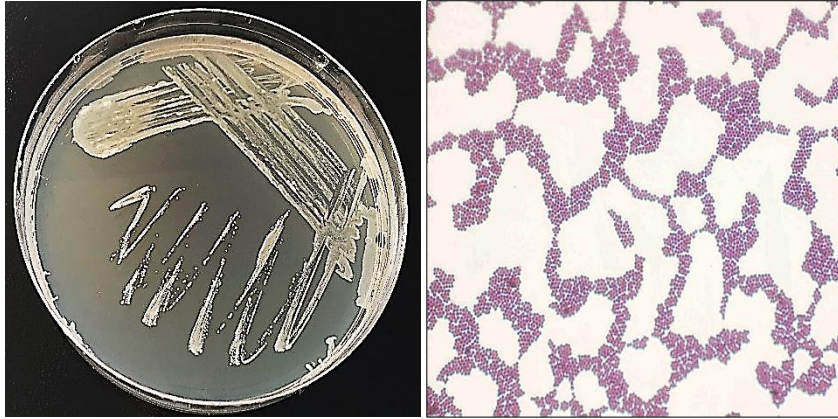
Bacillus licheniformis DQ071560.1

Gram stained. Magnification: 100x.



Bacillus megaterium KU550043.1

Gram stained. Magnification: 100x.



Staphylococcus succinus KJ888125.1

Gram stained. Magnification: 100x.



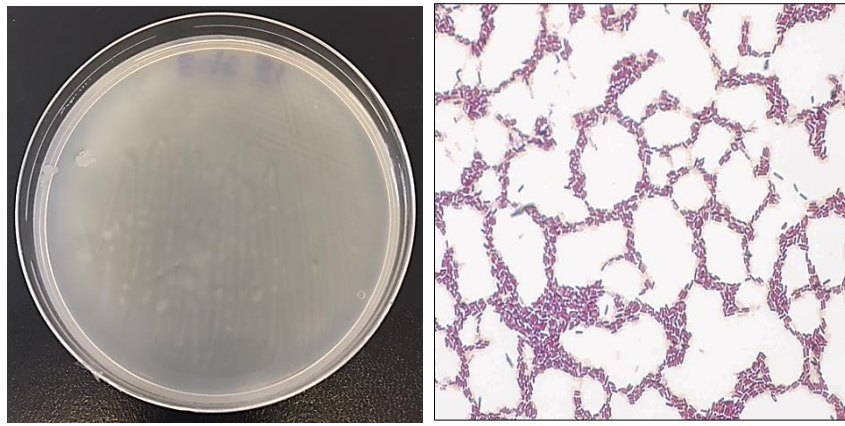
Bacillus stratosphericus KJ672335.1

Gram stained. Magnification: 100x.



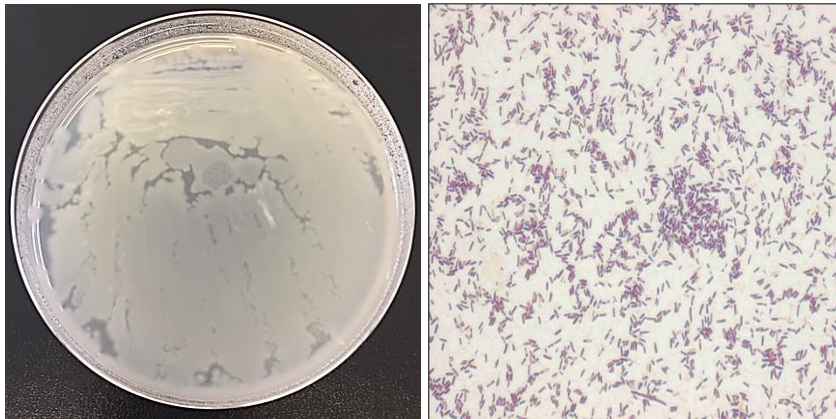
Bacillus licheniformis KT200463.1

Gram stained. Magnification: 100x.



Bacillus pumilus KU230023.1

Gram stained. Magnification: 100x.



Bacillus weihenstephanensis KC527665.1

Gram stained. Magnification: 100x.

Figure 2.8: Nutrient Agar plates showing cultured bacteria isolated from various from books and shelves in libraries and archive storerooms, and also microscopy images showing the bacteria under the light microscope.

Table 2.9: Bacteria present in dust settled on the surfaces of books and archive materials in the studied storerooms.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
9B3	<i>Pseudomonas jessenii</i>	99%	LN774645.1
3 SH11	<i>Bacillus cereus</i>	99%	KP192930.1
4 SH8	<i>Bacillus altitudinis</i>	98%	KT758615.1
7 SH11	<i>Bacillus pumilus</i>	98%	KP322017.1
1 SH6	<i>Bacillus stratosphericus</i>	98%	KJ672335.1
4 SH11	<i>Bacillus weithenstephanensis</i>	98%	KC527665.1
16 SH3	<i>Acinetobacter lofwii</i>	99%	KT387352.1
10B2	<i>Bacillus licheniformis</i>	99%	DQ071560.1
12SH4	<i>Bacillus megaterium</i>	99%	KU550043.1
14SH2	<i>Staphylococcus succinus</i>	99%	KJ888125.1
15SH3	<i>Bacillus pumilus</i>	99%	KU230023.1
2SH11	<i>Bacillus licheniformis</i>	99%	KT200463.1

Acinetobacter lwoffii

The genus *Acinetobacter* are Gram-negative, strictly aerobic, non-fermenting, non-fastidious, non-motile, catalase-positive, oxidase-negative, rod-shaped bacteria.

Acinetobacter spp. are widely distributed in soil, water, vegetables, and are frequently isolated from animals and humans *Acinetobacter lwoffii* is part of the communal microflora on human skin and mucous membranes and causes nosocomial infections like septicaemia, pneumonia, meningitis, urinary tract infections, skin and wound infections (Regalado *et al.*, 2009) and is associated with polluted water systems (Liu, 2011).

Staphylococcus succinus

This is a Gram-positive, spherical bacterium which forms characteristic rosettes with one central cell surrounded by two to five peripheral cells; strains can be isolated from clinical specimens such as blood culture, pus, cerebrospinal fluid, exudate, eye and wound swabs (Liu, 2011).

Bacillus weihenstephanensis

Bacillus weihenstephanensis is a psychrotolerant species capable of growing at temperatures as low as 4°C–6°C is implicated in food spoilage and in foodborne illnesses (Liu, 2011).

Bacillus altitudinis

Colonies are Gram-positive, rod-shaped, endospore-forming and catalase-positive bacteria. Colonies on nutrient agar are white, convex with a regular margin and 2–3 mm in diameter (Shivaji *et al.*, 2006). Growth occurs between 8°C and 45°C and at pH 5–8. *B. altitudinis* was isolated from cryogenic tubes used for collecting stratospheric air samples from high altitudes (Shivaji *et al.*, 2006). Its pathogenicity is unknown.

Bacillus pumilus

Bacillus pumilus is one of many species within the genus *Bacillus* which have emerged as novel foodborne human pathogens that cause severe fatal infections (Kimouli *et al.*, 2011). This bacterium has also been reported to cause infectious endophthalmitis and a case of central venous catheter infection with *Bacillus pumilus* in immunocompetent patients. *Bacillus pumilus* is a member of the intestinal flora of humans and can produce exotoxins that are cytotoxic for cultured mammalian cells (Kimouli *et al.*, 2011).

Bacillus stratosphericus

Colonies on nutrient agar are white, irregular, and raised. Growth occurs between 8°C and 37°C, but not at 40°C (Shivaji *et al.*, 2006). *Bacillus stratosphericus* were isolated from cryogenic tubes used to collect air samples at altitudes of 24, 28 and

41km. Also it was isolated from cryogenic tubes used for collecting air samples from high altitudes (Shivaji *et al.*, 2006). Its pathogenicity is unknown.

Bacillus megaterium

Bacillus megaterium is a Gram-positive, mainly aerobic spore-forming bacterium found in soil, seawater, sediments, rice paddies, honey, fish, and dried food. It is not regarded as an important human pathogen (Scholle *et al.*, 2003)

Pseudomonas jessenii

Cells are Gram-negative, asporogeneous and rod shaped motile by means of single polar flagellum. Colonies are smooth on nutrient agar circular non-pigmented and non-haemolytic when they are grown on blood agar. It is not regarded as human pathogen (Verhille *et al.*, 1999).

2.9.3 Discussion

The most frequently isolated bacteria belong to the genus *Bacillus* which is a Gram-positive rod (Table 2.9), while Gram-positive cocci were represented by *Staphylococcus* spp. Gram-negative rods, like *Pseudomonas jessenii* were also isolated, as was *Acinetobacter lofwii*, a non-fermentative Gram-negative a member of the genus *Acinetobacter*. Most of the identified microorganisms have been

previously isolated from a wide variety of different materials such as leather, paintings and books (Karbowska-Berent *et al.*, 2011). *Bacillus* may cause the decomposition of adhesives in paper and book bindings. *Bacillus* has the ability to transform into spores allows them to preserve viability in unfavourable, dry conditions for a long period of time. Owing to that, these bacteria were the most numerous species in settled dust (Karbowska-Berent *et al.*, 2011).

2.10 Isolation of bacteria from the upper surface of water taps

Hospital water systems often act as a source of nosocomial infection, particularly among immunocompromised and high-dependency patients (notably in intensive care units) (Boyles *et al.*, 2012). Patients are exposed to waterborne microorganisms in hospitals when bathing, showering or washing hands following direct contact with contaminated fixtures (e.g. wash-basins and taps); by ingestion of water; via indirect contact (e.g. medical equipment rinsed with water); and by staff-transfer via medical equipment rinsed with water; and finally, by inhalation of aerosols produced by a water source, and the aspiration of contaminated water. Such microbes often originate from biofilms and sediments in supply water, water storage tanks, and water distribution network pipes as well as associated equipment (Boyles *et al.*, 2012). Water quality can become corrupted rapidly due to the formation of biofilms by bacteria in the supply water. Taps are frequently contaminated with biofilm-containing opportunistic pathogens, notably *P.*

aeruginosa, and numerous cases of cross-infection from hospital taps have been reported (Boyles *et al.*, 2012). Immunocompromised patients are particularly susceptible to infection by such microorganisms, which can cause bacteraemia, pneumopathy, meningitis, and other conditions. *Pseudomonas aeruginosa* is particularly often isolated from these sources and has been reported to be lethal in 50%, 70%, and 20% of bacteraemia, pneumopathy, and meningitis cases, respectively (Boyles *et al.*, 2012).

2.10.1 Materials and Methods

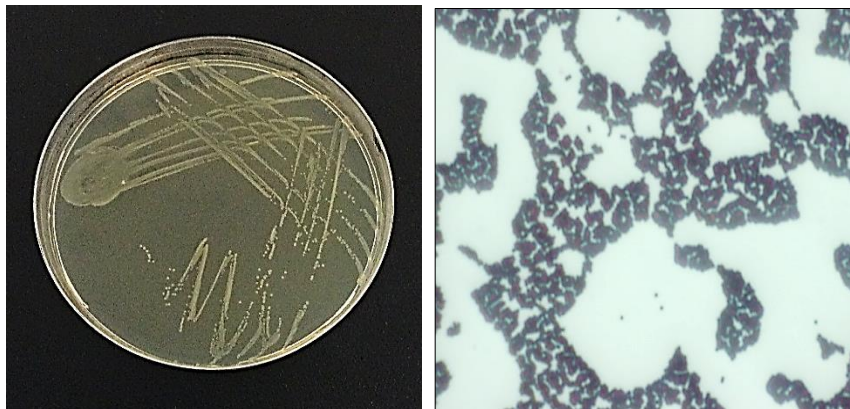
Sterile cotton swabs were wetted by dipping them in saline and used to collect samples from tops of water taps in toilets in Firth Court and the Student Union Building (University of Sheffield), and from volunteer house owners. All samples were labelled and streaked on to Nutrient Agar plates; the plates were then incubated at 25°C under aerobic conditions for 24 hrs. After incubation, the colonies were identified. Genomic DNA extracted from each isolate was identified using 16S rRNA. The extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. The methods used for preparation of PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis are detailed above.

2.10.2 Results

1) Isolation of bacteria from the upper surface of water taps

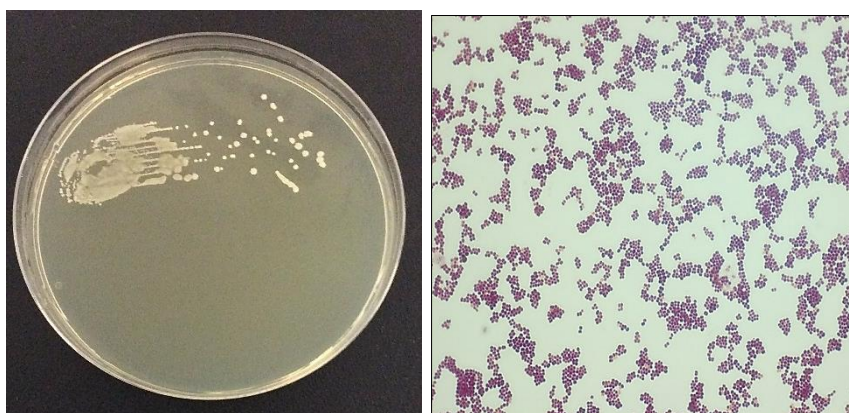
Bacteria were cultured using Nutrient Agar after isolation from the upper surface of water taps.

2) Light microscope images. The isolates were examined under a light microscope (Figure 2.9).



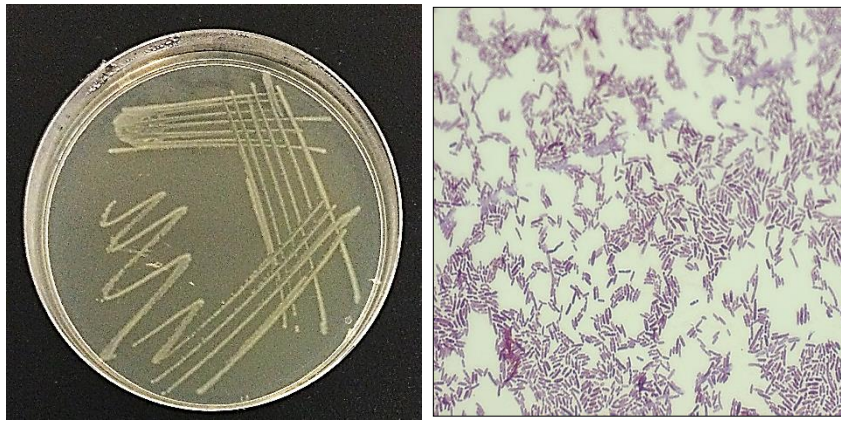
Delftia lacustris KT958881.1

Gram stained. Magnification: 100x.

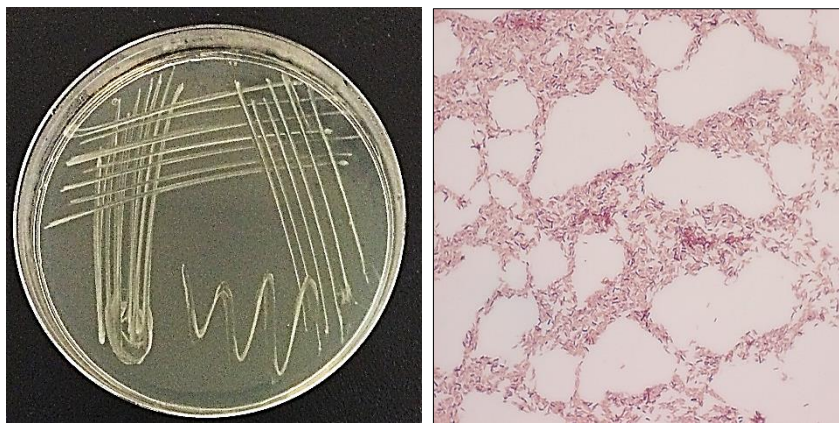


Rothia amarae NR_029045.1

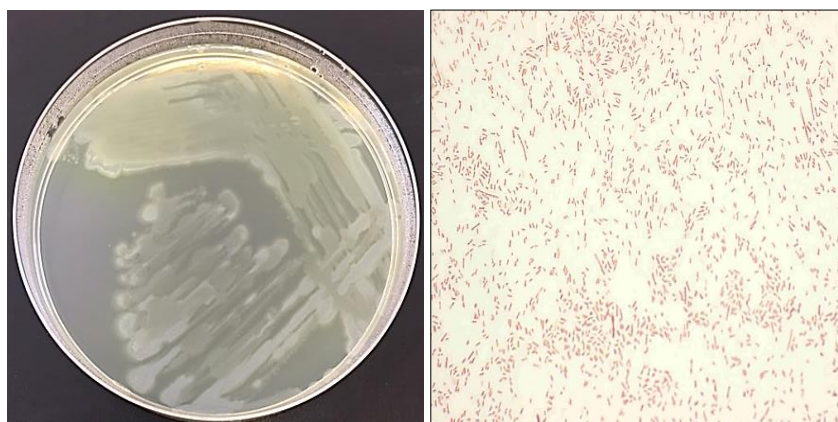
Gram stained. Magnification: 100x.



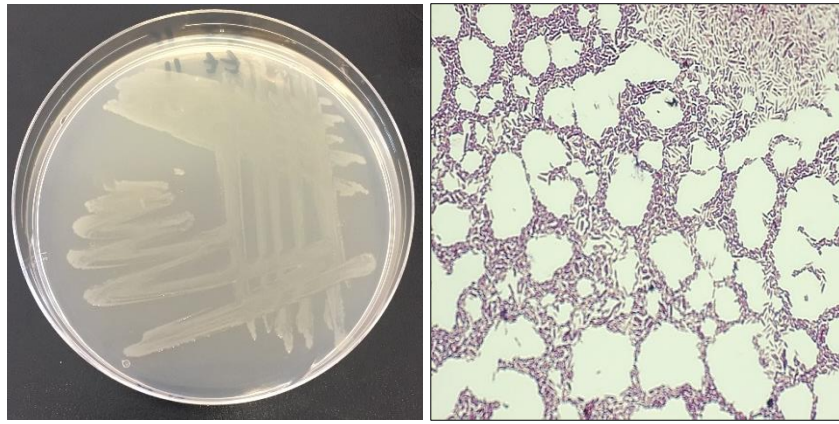
Delftia acidovorans (comamonas
acid) JX090199.1. Gram stained. Magnification: 100x.



Arthrobacter sanguinis NR_044399.1
Gram stained. Magnification: 100x.



Pseudomonas aeruginosa KF680991.1
Gram stained. Magnification: 100x.



Bacillus cereus GQ344804.1

Gram stained. Magnification: 100x.

Figure 2.9: Nutrient Agar plates showing cultured bacteria isolated from the top-surface of water taps, and also microscopy images showing the bacteria under the light microscope.

Table 2.10: Bacteria isolated from the upper surface of water taps.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
2tt	<i>Staphylococcus epidermidis</i>	98%	KT887972.1
3tt	<i>Rothia amarae</i>	99%	NR_029045.1
4tt	<i>Acinetobacter johnsonii</i>	99%	KP763485.1
6tt	<i>Delftia lacustris</i>	99%	KT958881.1
7tt	<i>Pseudomonas aeruginosa</i>	98%	KF680991.1
8tt	<i>Delftia acidovorans</i> (<i>comamonas acid</i>)	99%	JX090199.1
9tt	<i>Arthrobacter sanguinis</i>	99%	NR_044399.1
10tt	<i>Delftia acidovorans</i> (<i>comamonas acid</i>)	98%	KJ781879.1
11tt	<i>Bacillus cereus</i>	94%	GQ344804.1

Pseudomonas aeruginosa

Pseudomonas spp. act as opportunistic pathogens with *P. aeruginosa* being the most clinically significant as a major cause of lower respiratory tract, bloodstream, and urinary tract infections, particularly among patients in intensive care units or with compromised immunity (Liu, 2011). This bacterium is often isolated from

moist environments within health care settings, such as taps, toilets, showers, respiratory equipment, and cleaning solutions. *Pseudomonas aeruginosa* is a major pathogen in chronic lung diseases, most notably in cystic fibrosis. It causes infections in critically ill patients in intensive care units, burn patients, and particularly when immunocompromised (Liu, 2011). Such infections include ventilator-associated pneumonia, catheter-related bloodstream or urinary tract infections; other infections are derived from medical devices or penetrating trauma, sepsis in neutropenic cancer and transplant patients, skin and soft-tissue infections in burn and diabetic patients, postoperative endophthalmitis and necrotizing otitis externa in diabetics (Liu, 2011).

***Delftia acidovorans* (*Comamonas acidovorans*)**

Cells of this bacterium are Gram-negative straight to slightly curved rods, which occur singly or in pairs. They are motile and are oxidase and catalase positive. This organism has been isolated from serious infections such as central venous catheter-associated bacteraemia, corneal ulcers, otitis media exist (Khan *et al.*, 2012). *Delftia acidovorans* infections have also been reported from cases of endocarditis, ocular infections, acute suppurative otitis media, bacteraemia, urinary tract infections, intravascular catheter related infections and empyema; serious infections have been found amongst immunocompromised patients. Patients with indwelling devices are at particular high risk of acquiring *Delftia acidovorans* infections (Khan *et al.*, 2012).

Acinetobacter johnsonii

Acinetobacter spp. have been isolated from a variety of opportunistic infections, including septicaemia, pneumonia, endocarditis, meningitis, skin and wound infection, and urinary tract infection. *Acinetobacter* spp. (e.g., *A. johnsonii*) have been shown to form an important component of the communal microflora on human skin and mucous membranes (Bergogne-Bérézin *et al.*, 1996)

Rothia amarae

All *Rothia* species are Gram-positive and are isolated from humans or animals as well as from soil and water. They are particularly associated with dental caries and periodontal disease notably in immunocompromised hosts, but rarely in healthy hosts. Clinical syndromes linked to *Rothia* infection include bacteraemia, endocarditis, meningitis, peritonitis, bone and joint infections, pneumonia, skin and soft tissue infection, endophthalmitis, and prosthetic device infection (Fan *et al.*, 2002; Xion *et al.*, 2013; Ramanan *et al.*, 2014).

Arthrobacter sanguinis

Arthrobacter are Gram-positive aerobic bacilli commonly found in the environment, although they can also cause endophthalmitis after intra-ocular implantation, infective endocarditis in intravenous drug- abuser, featal demise and disseminated intravascular coagulation in pregnant women and catheter-related bacteraemia in leukemic patients (Mages *et al.*, 2008; Yap *et al.*, 2015).

Arthrobacter infections are often difficult to diagnose using conventional

biochemical assays. *Arthrobacter* related peritonitis can be successfully treated with appropriate parenteral antibiotics without the need for catheter removal (Mages *et al.*, 2008, Yap *et al.*, 2015).

2.11 Determination of the number of bacteria on hands after washing and drying normally and with warm air hand dryer

2.11.1 Introduction

Hand drying represents the final part of the hygiene procedure in a public toilets and if the toilet is designed well, the number of surfaces which the user subsequently touches will be limited or reduced to near zero. This is essential because wet hands can spread up to one thousand times more bacteria than can dry hands (Smith, 2009). This is essentially because water transfers easily between surfaces and because bacteria do best in moist environments (Redway and Fawdar, 2008). As a result, it is critical that hands are not contaminated with bacteria due to the drying process (Harrison *et al.*, 2003). The most frequently used means of hand drying are paper towels, hot air dryers, jet air dryers and cloth towels. It has been suggested that air dryers should be avoided as they accumulate aerosols from toilets which then contaminate hands (Snyder, 1998). This last mentioned author concludes that the use of paper towels decreases the number of bacteria on hands, while hot-air dryers conversely increased contamination by some bacteria; a finding suggestion that has been widely debated (Holah, 2011); many contend

however, that hot air dryers are often slow and inefficient, leaving the hands of users moist and possibly still contaminated. Cloth roller towels are similarly not recommended essentially because they are generally of low capacity, and when a roll comes to an end it becomes available for common use and is therefore likely to increase the spread of pathogens (Snyder, 1998).

This work was undertaken with the aim of evaluating the performance of warm air hand driers, in toilets relative to bacterial contamination. First, the ability of warm air driers to dry hands hygienically was evaluated by measuring the number of microorganisms on different working days. Secondly, it was determined if warm air driers influence the level of air-borne microorganisms in the washroom environment, as was suggested by Knights *et al.* (1993). Finally, the surfaces of warm air driers and other washroom areas were examined for total viable counts in order to determine if the use of air driers affects bacterial distribution.

2.11.2 Materials and Methods

General purpose, non-antimicrobial bar soap was used for hand washing. Hands were wetted with tap water, and the soap applied and lathered together vigorously for 5 minutes, covering all surfaces of hands followed by rinsing with tap water. Finally, the hands were shaken five times in order to remove of excess water before drying. Areas of the washed hands were applied to the surface of Nutrient Agar plate; the plates were then incubated at 37°C for 48h. Warm air hand-dryers were next used for hand drying; the air temperature applied was 0°C -40°C at a

distance of 15 cm. When hands were placed in the 15 cm drying space, warm air flowing automatically was turned on. During drying, hands were either rubbed or had their palms turned upward and were held stationary. Then dried hands and fingers top were touched onto the surface of nutrient agar plate. The plates were incubated at 37°C for 48 h and scored for the presence or absence of growing bacteria.

2.11.3 Results

The results show that no bacteria were isolated from hands dried in the normal way, but were isolated from hands when dried using an air dryer.



Figure 2.10: Agar plates used to for the assessment of bacterial contamination on hands. (Left) hand washed and dried with a warm air hand dryer, (Right) hand washed and dried without warm air dryer.

2.12. Quantification of bacteria transferred from hand warm air dryers

2.12.1 Materials and methods

Nutrient Agar plates were used in this experiment to isolate the transferred bacteria from hand warm air dryers. Any bacteria transferred to the plates from the hot air driers were counted and the effect of drying times was determined.

2.12.2 Results

Table 2.11: Showing the mean of the bacteria count using three different types warm air dryer (WAD,1-3) for different drying times.

	5 Sec	10 Sec	15 Sec	20 Sec	25 Sec	30 Sec	35 Sec	40 Sec	45 Sec	50 Sec	55 Sec	60 Sec
WAD1	0	2	5	6	3	9	12	10	23	41	31	30
WAD2	12	17	18	22	18	16	50	37	15	49	44	63
WAD3	19	20	15	24	10	8	11	20	18	19	21	30

Table shows that there was a correlation between the drying time and the resultant bacterial counts, with the counts generally increasing with the drying time exceeding 30 seconds. Redway and Fawdar (2008) investigated the spread of

contamination from different drying methods and concluded that microorganisms were spread significantly further when an air blade dryer was used instead of paper towels. While not significant if only non-pathogenic microorganisms are spread, the risk of pathogen contamination of the environment will result when pathogens are present on hands and are liberated into the air (Blackmore, 1989).

2.12.3 Discussion

From the results presented in this Thesis, it is clear that hand dryers distribute bacteria into the surrounding air. As has been mentioned elsewhere in this Thesis, many of these bacteria, even when not direct pathogens, pose a potential problem for immunocompromised patients here. Hand dryers have recently become increasingly popular and for reasons of cost, these machines are often supplied in place of traditional cloth and paper towels (Huang *et al.*, 2012). This does not present a health problem for most hospital patients and visitors (except where immunocompromised). The use of hand dryers in medical settings should therefore be reviewed and, if necessary, they should be replacing by traditional approaches, the consensus being that paper towels are the preferred method of hand drying in such settings (Best *et al.*, 2014; Huang *et al.*, 2012).

2.13. General Discussion regarding potential pathogens in the environment

Boyce *et al.*, (1997) have shown that surfaces in hospitals are often contaminated with MRSA, as are staff. This points out the necessity of using detergents and antibacterial wipes to reduce bacterial loads, especially postoperative MRSA.

Vancomycin-resistant enterococci (VRE) can achieve long-term survival in medical environment, even following extensive cleaning and disinfection.

Pathogens can, in fact, often be spread when cleaning cloths are a) re-used on surfaces, b) when little contact time between a surface and the applied disinfectant, and c) when surface are casually sprayed and wiped instead of being aggressively scrubbed. VRE appears to be particularly efficient at surviving successive disinfection, regimes, even so-called “double bleach-based cleaning”.

Disinfectants can be effective if hospital surfaces are scrubbed daily and when this is associated with a hand cleaning programme. The surface screening of cultures has also proved effective, as has the use of 5% sodium hypochlorite to clean all surfaces at least three times daily. Both MRSA and VRE can survive for long periods which presents an obvious risk in relation to the movement of patients into rooms which have been previously occupied by infected patients.

Hospital sinks are an obvious source of pathogenic bacteria, with antibiotic resistant *Klebsiella pneumoniae* strains showing particularly extended survival within plumbing thereby, in extreme cases, necessitating the replacement of sinks and related pipe-work. Pathogen survival is also linked to biofilms adhering to

surfaces on sinks and associated plumbing and unfortunately the use of chlorine-containing products is often ineffective and even prolonged use fails to destroy all biofilms thereby necessitating the physical disruption of any biofilm which is found to be growing in contaminated plumbing systems.

2.13.1 Antimicrobial surfaces

Some high-tech solutions are currently available to control surface pathogen contamination, by so-called, “self-sanitizing surfaces” which include hard metals (e.g. copper and silver) and novel materials such as light-activated titanium dioxide-containing surfaces which prevent the accumulation of surface microbes (Dancer, 2014). Several types of antimicrobial surfaces including antiadhesive coatings, and antimicrobial coatings exist including **Triclosan** (Microban triclosan): which is used on surfaces and in many antibacterial liquid and bar soaps and 30% of bar soaps, although evidence for resistance is developing. **Silver:** Silver ions (Ag^+) bind to thiol ($-\text{SH}$) groups in microbial enzymes and proteins and lead to microbial-cell inactivation; such coatings are not, however, permanently active and resistance can develop (Dancer, 2014). **Copper:** Copper (as will be discussed below) is also toxic to pathogens and coatings can be used to reduce microbial contamination. **Polycationic antimicrobial surfaces:** these are surfaces which are treated with hydrophobic, negatively charged poly-cations which kill bacteria by bringing about physical damage to the cell-envelope. **Light-activated antimicrobial surfaces:** These produce unselective, reactive radicals

which kill a range of microorganism and reduce the chances of resistance developing. Two types are available, the first is based on a photosensitizer immobilized within a coating, while the second is a coating which contains a titanium dioxide (TiO₂)-based catalyst (Dancer, 2014).

CHAPTER 3

STUDIES ON SURVIVAL OF BACTERIA ON VARIOUS SURFACES

3.1 Studies on the survival of bacteria on smooth and rough unglazed ceramic tiles under ambient conditions

3.1.1 Introduction

Ceramic tiles are frequently used in the built environment, notably in hospitals and provide an obvious survival environment for potentially pathogenic microorganisms. Ceramic tiles are widely used in domestic and health care settings. They are obviously advantageous in being durable, water repellent, easy to keep clean, and are readily exposed to bacteria-killing sunlight. The grouting between tiles may however, act as a reservoir for microbes, including pathogens. Tile surfaces differ, between smooth and rough. Logically, one would assume that rough, unglazed tiles would provide a more suitable environment for microbes and as a result, these are generally preferred for use in health-care settings. The aim of this work was to determine the survival (in the ambient environment) of a variety of bacteria on smooth (glazed) and rough (unglazed) tiles.

3.1.2 Materials and Methods

Ceramic tiles having either smooth or rough surfaces (9 cm²) were firstly sterilized by autoclaving (Figure 3.1). The test bacteria were then grown on Nutrient Agar medium as a pure culture, while yeasts (*Candida albicans* and *C. rugosa* were

grown on Sabouraud Dextrose agar). Bacterial and yeast suspensions from colonies were prepared in sterile saline equal to McFarland 0.5 turbidity (the density of a bacterial suspension equal 1.5×10^8 colony forming units CFU/ml). The tiles were inoculated by placing on their uppermost chosen surface approximately 10 microliters of either a bacterial or yeast suspension, which was allowed to outspread and dry under sterile conditions and left at room temperature (18-23°C). The tiles were transferred to sterile Falcon tubes containing sterile water and serial dilution counts were made.

In the second series of experiments, the tiles were inoculated with *Candida rugosa* and at the end of the same exposure period were washed and serial dilution plated onto Sabouraud Dextrose agar.



Figure 3.1: The two types of ceramic tiles used: A) smooth ceramic tile, B) rough ceramic tile.

3.1.3. Results

3.1.3.1 Bacterial survival on ceramic tiles

Figure (3.2) shows the number of *S. aureus* and *E. coli* bacteria isolated from the two tile types. Numbers of *S. aureus* on the rough tiles increased over the incubation period, while number on the smooth tiles decreased. In both cases however, bacteria were still present after 65h. In the case of *E. coli*, the number decreased over the incubation period, while there was a slight increase in the numbers of bacteria on smooth tiles (Figure 3.3). The results show that both *S. aureus* and *E. coli* can survive on both tile types over a 65 h period, a finding which is obviously relevant to pathogen survival and transfer in healthcare settings. Staphylococci are among the most resistant non-spore forming bacteria, and they can survive at various environmental conditions. They can be cultured from dried clinical material after several months. Yazgi *et al.* (2009) found that *S. aureus* survived on laminate for 75.4 days and inox sheet for 68.2 days, whereas these survival days were 73.8 days on ceramic tiles and 63.4 days on vinyl flooring. The length of survival of these organisms on the various materials may have significance for infection control in hospitals. For example, the ceramic tiles tested in this study are the material used for floors and walls of hospitals that are handled by both patients and staff when they are touched or cleaned. *Staphylococci* and *E. coli* survived for days on this covering materials, suggesting that they could act as reservoirs for such pathogenic bacteria.

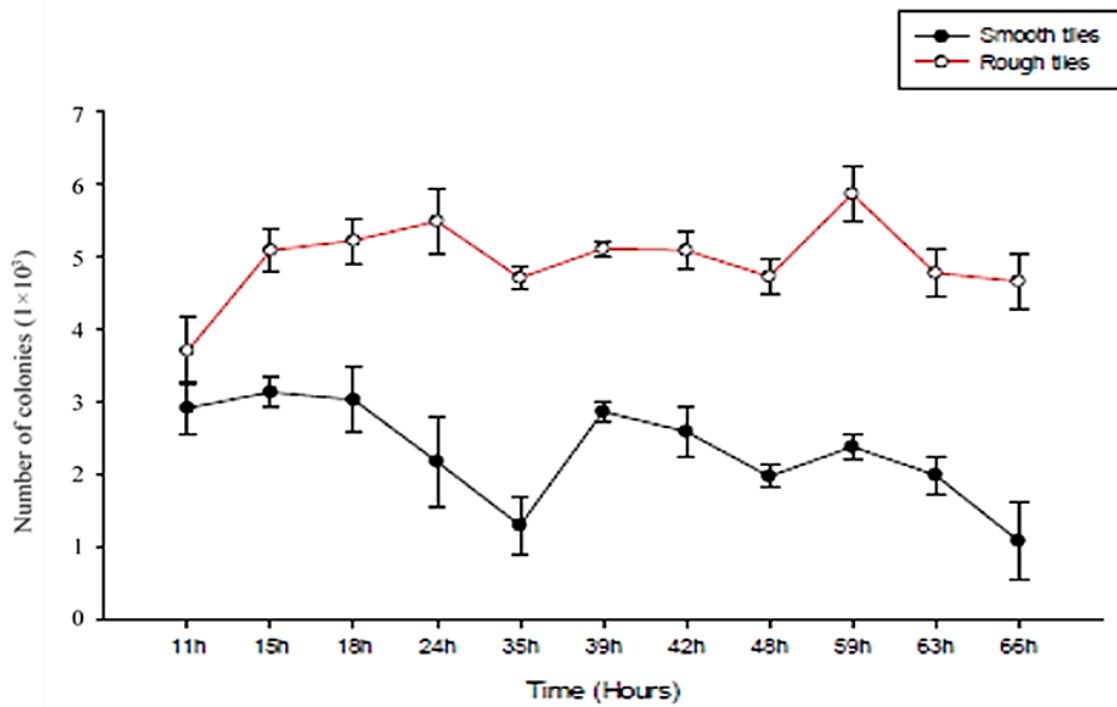


Figure 3.2: Survival of *S. aureus* on ceramic tiles during 65h.

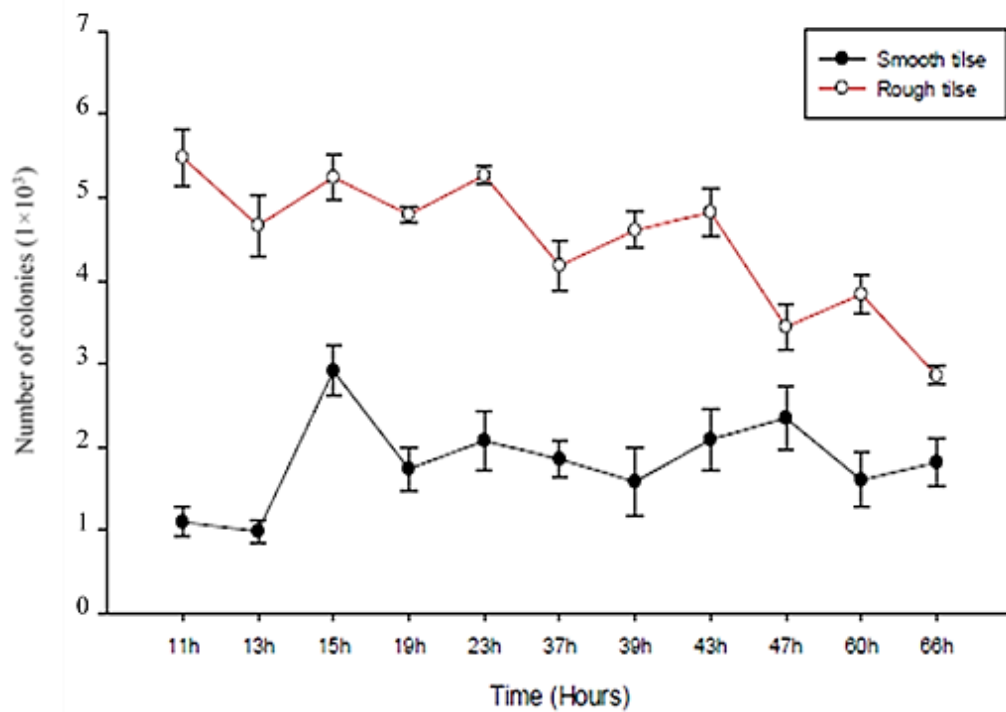


Figure 3.3: Survival of *E. coli* on ceramic tiles during 65h.

3.1.3.2 Yeast survival on ceramic tiles

The results (Figure 3.4) show that *Candida rugosa* survived for a period of 72h on both smooth and rough tiles with numbers generally increasing for 53-56h when they declined rapidly but the yeast was still present in a viable state after 72h. The yeast is associated with nosocomial infections in patients. Where comparisons are possible for the yeast data, this result agrees with Traoré *et al.* (2002) who found *C. albicans* and *C. parapsilosis* remained viable for at least 3 and 14 days when dried on surfaces materials. The results of the ability of *C. albicans* and *C. parapsilosis* to survive on glass and metal carriers over a 14-day period are summarized in a study by Traoré *et al.* (2002). *Candida parapsilosis* was found to survive much better than *C. albicans* on both types of non-porous surface. Also, *C. albicans* was undetectable at the end of the third day, while *C. parapsilosis* remained detectable even after 14 days under ambient conditions. There was a significant difference between the survival of *C. parapsilosis* and *C. albicans* after 14 days on glass carriers and stainless steel carriers. These results indicate that ceramic tiles play a role as reservoirs or vectors for fungi because those tested generally remained viable on these surfaces for many days. In this age of increasing antifungal resistance, these survival data indicate that the appropriate disinfection of the environment and control procedures should be used to control infections in hospitals.

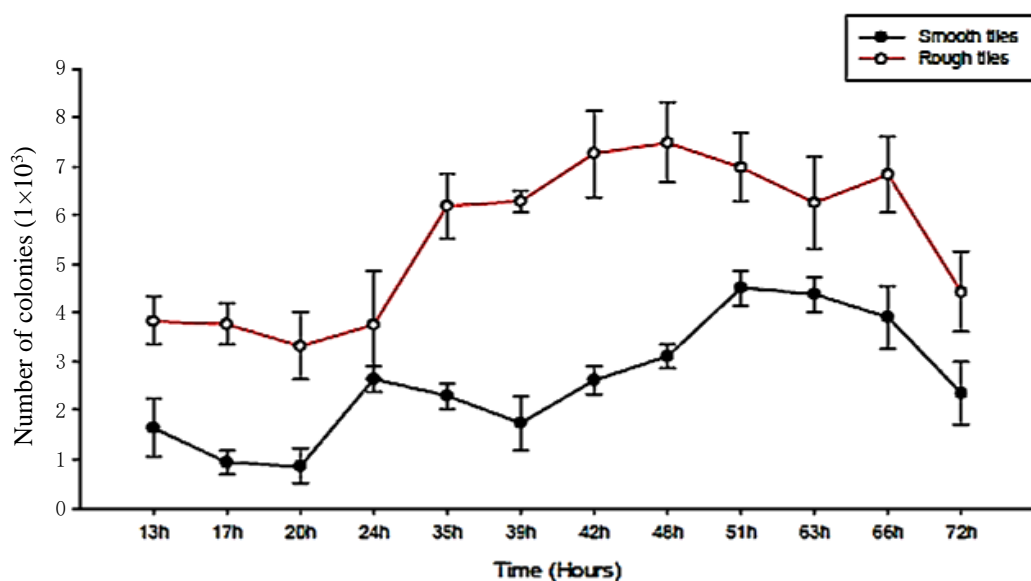


Figure 3.4: Survival of *Candida rugosa* on ceramic tiles.

3.1.3.4 Discussion

The most important factor in transmission of microorganisms in the environment is the ability of microorganisms to survive on environmental surfaces. The contamination and colonization of surfaces is important in relation to the transmission of nosocomial infections pathogens, such as methicillin-resistant *S. aureus* and vancomycin resistant Enterococci (VRE). For example, a range of floor covering materials are used in intensive care units, operating rooms, clinics, and laboratories of hospitals due to which influence the rate of microbial survival and as a result, need to be carefully chosen in order to minimize microbial colonization and survival. The preferred materials are smooth-surfaced and include ceramic tiles, laminates, inox sheets, and vinyl which are easy to disinfect

during cleaning. Neely and Maley (2000) studied the survival of several clinical and environmental *staphylococci* and *enterococci* on fabrics and plastic that are used in hospitals. They found that the survival period of *S. aureus* was more than 90 days on various surfaces, whereas it was 18 to more than 80 days for *enterococci*. It is clear from the results for both tiles and the study discussed in this Thesis that covering materials have a significant influence on the survival periods of bacteria. The Thesis results show that ceramic tiles play a role as reservoirs or vectors for the yeast, *C. rugose*, because this organism generally remained viable on these surfaces for many days. Strategies to reduce the rates of nosocomial infection with these pathogens should conform to established guidelines, with an emphasis on thorough environmental cleaning and use of Environmental Protection Agency–approved detergent-disinfectants.

3.1.4 Discussion

Bacterial and yeast survival was tested on both smooth and rough ceramic tiles. The results show that ceramic tiles provide reservoirs for microbes because the tested species generally remained viable on these surfaces for a number of days; the length of survival on both smooth and rough ceramic tiles being related to both the genus and the species. Clearly cleaning strategies aimed at maintaining low pathogen populations on hospital tiles, whether rough or smooth need to take such apparently anomalous findings into consideration.

3.2. Studies on the survival of bacteria on toothbrushes under ambient conditions

3.2.1 Introduction

The environment of the toothbrush is affected by many conditions whether it is the architecture of the toothbrush itself regarding bristles or by adjusting the pH level. These conditions alter the population of bacteria on the toothbrush. While the toothbrush is not the ideal niche for a microbe, the toothbrush is capable of supporting microbial life (Downes *et al.*, 2008).

3.2.2 Materials and methods

3.2.2.1 Bacterial survival on toothbrushes during a month

The new toothbrushes were prepared and were sterilized by 10% ethanol spray. The strains of the test bacteria were grown on the Nutrient Agar medium as a pure culture. Bacterial suspensions from the colonies were prepared in sterile saline equal to McFarland 0.5 turbidity. The toothbrushes were contaminated by placing on them on approximately 10 microliter volume of the bacterial suspension, and the inoculum allowed to outspread and dry itself under sterile conditions. Then the toothbrushes were kept at room temperature (18-23°C) (Figure 3.5). Beginning from the third day, toothbrushes were taken and placed in a sterile Falcon tubes contained sterile water. The numbers of viable organisms in the resultant suspension were determined by serial dilution and plating on to Nutrient Agar. The

step was repeated after 5, 15, and 20 days and then the plates were incubated at 37°C for 48 h.



Figure 3.5: New toothbrushes inoculated with the bacterial suspension of *S. aureus* and *E. coli*.

3.2.3. Results and discussion

Numbers of inoculated *S. aureus* and *E. coli* on toothbrushes decreased after 5 days of storage (Figure 3.6 A, B) and then numbers remained constant (except for small increase in numbers of *S. aureus* at 37 days. *Escherichia coli* also survived for 5 days on tested toothbrushes.

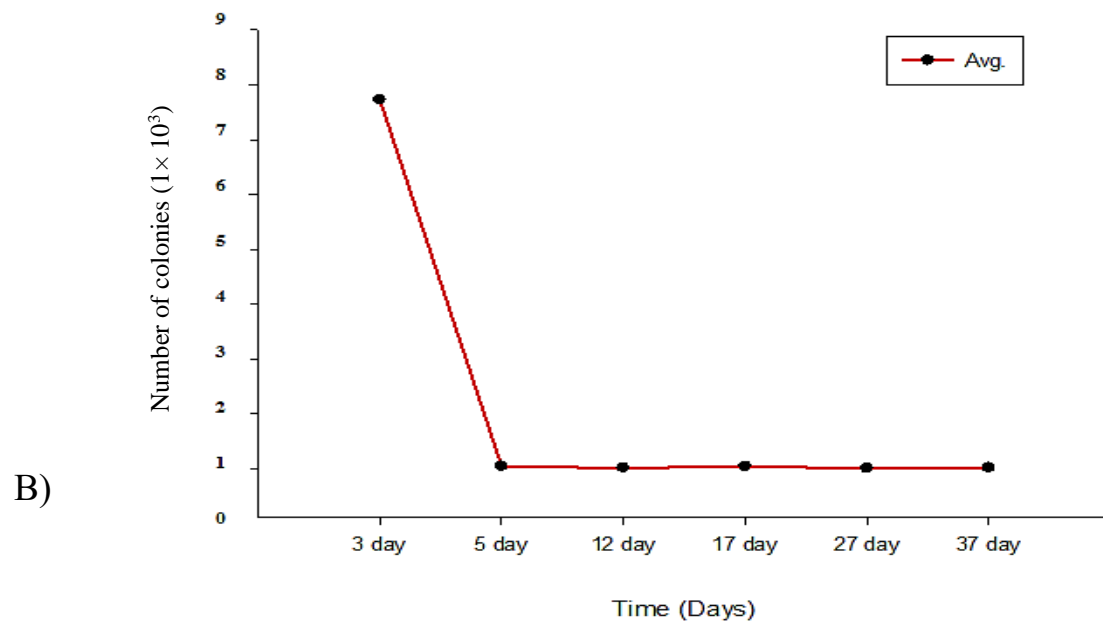
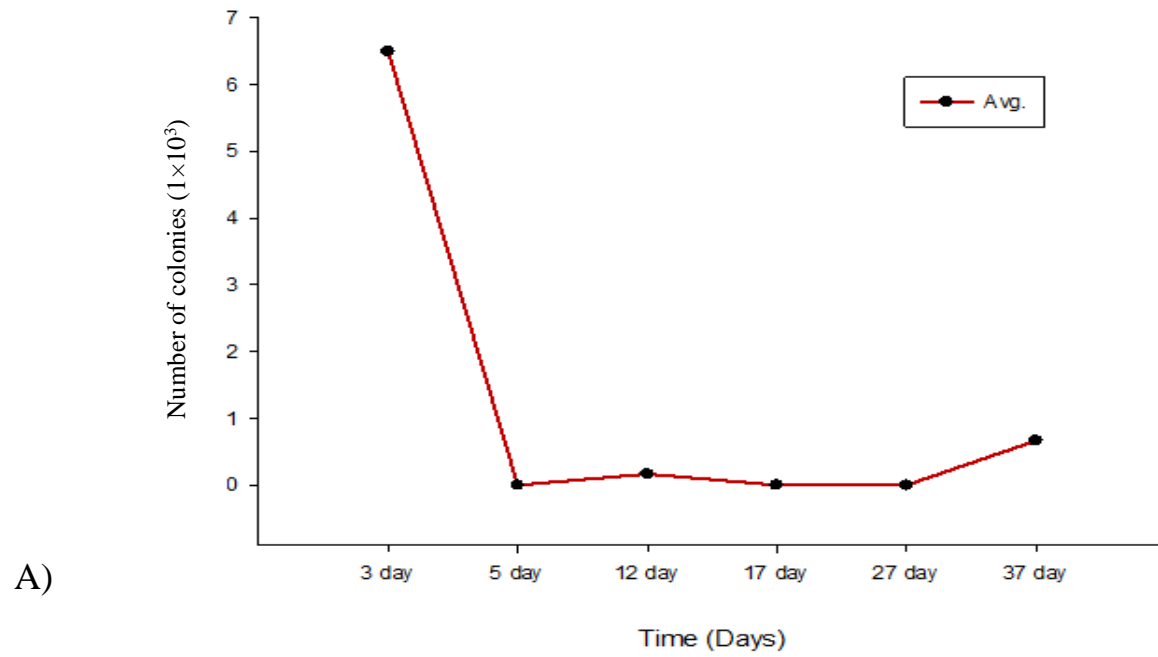


Figure 3.6: Population density of *Staphylococcus aureus* (A) and *E. coli* (B) when inoculated onto toothbrushes (analysis of variance).

* All values after 5 days were significantly different from control ($p=0.05$)

3.2.4 Discussion

The aim of this study was to compare bacterial survival on new conventional toothbrushes in the laboratory. The survival times of *Staphylococcus aureus* and *E. coli* were compared on toothbrushes stored in the laboratory. Toothbrushes can become contaminated through contact with the environment, and bacterial survival is affected by toothbrush storage containers. Dayoub *et al.* (1977) found that toothbrushes placed in closed containers and exposure to contaminated surfaces give higher bacterial counts than those left open to air. Mehta *et al.* (1990) found that the use of a cap for toothbrush storage increased bacteria survival. Increased humidity in the environment increases bacterial survival on toothbrushes and bacteria can survive for more than 24 hours in the presence of moisture. The persistence of viable *Staphylococci* on a drying toothbrush in the humid atmosphere of a toothbrush holder is not surprising; interventions such as chlorhexidine, toothpaste, mouthwash, and ultraviolet sanitizers can however, reduce bacterial survival (Downes *et al.*, 2008).

Although sterile at birth, a great variety of microbes develops during the first day of life, including: *Streptococcus*, *Staphylococcus*, *Neisseria*, *Candida*, *Lactobacillus*, *Veillonella* and coliforms (McCarthy *et al.*, 1965; Socransky and Manganiello, 1971). Mutans *Streptococci* (MS), which is the primary etiological agent of human dental decay, is however, only found following dental eruption because it needs a hard surface on which to develop. Catalanotto (1975), Fujiwara (1991), Glass, (1992) and Glass and Lare (1996) showed that toothbrushes are

particularly relevant to the transmission of pathogens amongst immunocompromised patients, including coliforms picked up from the bathroom environment (Verran, 1996). While it is of course unlikely that toothbrushes will be intentionally shared amongst families and individual patients in hospitals, the potential still remains for accidental contamination-transfer. More importantly perhaps, is the fact that pathogens can re-infect patients from toothbrushes at the initial point when they are first immunocompromised and recycle pathogens, including MRSA. Toothbrushes are often exposed to sunlight and nearly always to drying –wetting cycles, all of which can reduce survivability. As a result, toothbrushes rapidly become contaminated with oral microbes, including caries causing pathogens like *Streptococcus mutans*, and opportunistic yeast pathogens like *Candida albicans* (Sammons *et al.*, 2004). Additionally, organisms not usually considered to be components of the oral microflora can frequently be isolated from toothbrushes, including Enterobacteria and pseudomonads. Toothbrushes can therefore be seen as a potential source of both oral and systemic infection and re-infection (Sammons *et al.*, 2004), particularly in immunocompromised patients and such organisms may also cause worrying medical problems in pregnant women (Bunete *et al.*, 2000). The aim of the work described in this section was to therefore to determine if household toothbrushes are contaminated with bacteria.

3.3 Influence of copper and plastic surfaces on the survival of bacteria in relation to the health care environment

3.3.1 Introduction

Metallic copper (Cu) surfaces have antimicrobial properties against a variety of different microorganisms and copper touch surfaces are likely to be increasingly used in public places including hospitals (Espírito Santo *et al.*, 2011). In recent hospital trials, non-Cu surfaces in frequent contact with both patients and staff were replaced with their Cu equivalents. The use of metallic Cu resulted in diminishing bacterial surface-loads of up to 90% as compared to no-copper controls (Espírito Santo *et al.*, 2011). Molecular mechanisms that result in rapid killing of Cu surface-exposed bacteria and yeasts were studied and shown to result from a sharp shock of extreme and immediate Cu-ion overload combined with severe membrane and cell envelope damage, although similar low mutation rates were observed in cells obtained from both Cu and control surfaces (Espírito Santo *et al.*, 2008; Quaranta *et al.*, 2011).

Mehtar *et al.*, (2008) conducted studies in which cells in buffer were applied to copper surfaces, incubated under ambient conditions, and were seen to be killed within hours, thereby mimicking contact of microbes to dry copper touch surfaces. Under such conditions, most microbes were found to be killed within minutes. Quaranta *et al.* (2011) found that copper ions are released from metallic copper upon contact with cells, a process which undoubtedly contributes to contact-

mediated killing. Extracellular supplementation with substances which are protective against oxidative stress (e.g. Catalase, superoxide dismutase, or mannitol, a hydroxyl radical quencher) increased the time needed to kill copper surface-exposed *E. coli* cells, indicating a role for this process in Cu-mediated killing.

3.3.2 Materials and Methods

Survival of bacteria on the surfaces of copper and plastic plumbing surfaces

The antibacterial activity of copper surfaces was determined by overlying suspensions of *Staphylococcus aureus* and *Escherichia coli* on copper surfaces. Two copper items were used; a copper pipe push-fit elbow 15mm, a copper pipe compression stop end, 15 mm and (as controls) a plastic pipe connector 22mm (all obtained locally from Wickes Building Supplies Ltd) (Figure 3.7). All copper pipes were sterilized by autoclaving (120°C for 20 min), while the plastic pipes were sterilized using a 10% ethanol spray. Bacterial suspensions from colonies were prepared in sterile saline equal to McFarland 0.5 turbidity. The pipes were contaminated by transferring a 10 microliter volume of the bacterial suspension, and the inoculum was allowed to out-spread and dry itself under sterile conditions. The experiments were performed at 18-23°C and each assay was carried out in duplicate. Results were calculated after 20 days. The numbers of viable organisms in the suspension were determined by serial dilution and plating on to Nutrient Agar plates; the plates being incubated at 37°C for 48h.



Figure 3.7: Copper and plastic pipes used to evaluate the antibacterial activity of copper and plastic surfaces.

3.3.3 Results

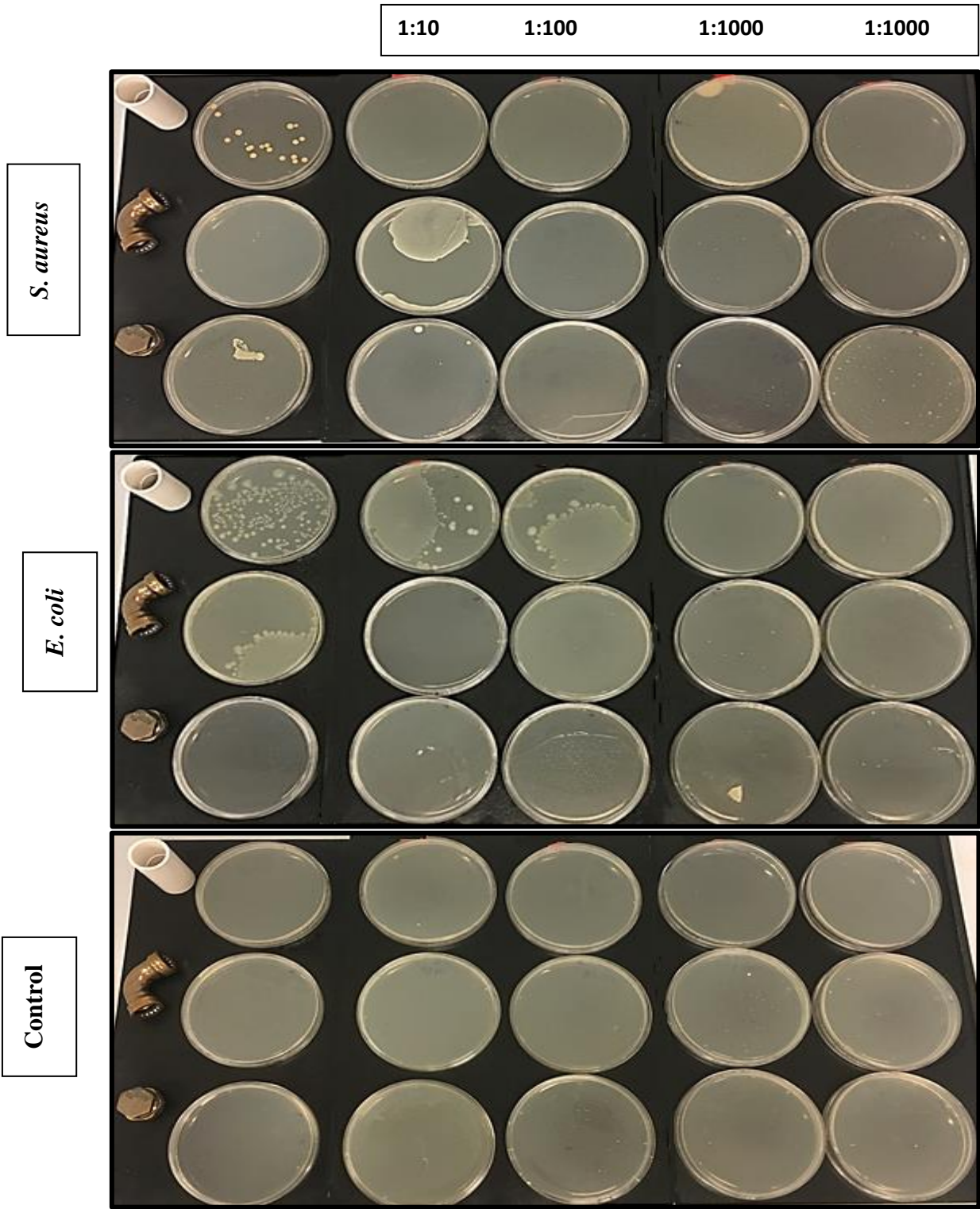


Figure 3.8: *S. aureus* and *E. coli* inoculated onto copper and plastic pipes.

Although copper and brass are traditionally used by UK plumbers for water pipes. in both hospitals and homes, they are gradually being replaced by plastic piping. It was of interest therefore to determine the survival of bacteria on both types of these surfaces. Copper is well known to be an antibacterial, and its use in medical environments is likely to lead to the continuous reduction of environmental microbial contamination, including MRSA. In viability assays, low counts of *Staphylococcus aureus* were seen on copper surfaces, as compared with those obtained on the plastic, control surfaces (Figure 3.8). The number of bacteria isolated from the plastic surfaces was consistently higher than the number isolated from copper surfaces. The survival rate of bacteria on the copper surfaces was low and none of the inoculated bacteria survived after 20 days of exposure. These results are similar to those found by Noyce *et al.* (2006) for copper, where survival was significantly lowered, with complete kill of MRSA being achieved after 90 min of exposure. Complete kills were produced on copper for all three strains of MRSA after 45, 60 and 90 min. The results presented here showed that *E. coli* failed to survive on copper pipes. These results are in agreement with those published by Wilks *et al.* (2005) who found that no viable *Escherichia coli* O157H7 were found on any of the copper pieces which they inoculated, even after only seventy-five to ninety minutes of exposure. Such results show that copper surfaces can remove cross contamination of pathogens in both households and health care settings and the continued replacement of other plumbing materials

with copper should minimize the risk of the survival and transmission of hospital-acquired infections.

3.3.4 Discussion

Copper has inadvertently been used as an antimicrobial agent for millennia, but its mode of action against microbes is still largely unknown. Today, nosocomial infections are a major health care threat and are responsible for large numbers of deaths and the resultant additional hospital costs, notably in the United States (Mehtar *et al.*, 2008). The effectiveness of copper as an antimicrobial material is not restricted to bacteria as it also kills *Candida albicans* (and other pathogenic yeasts) which have emerged as important agents of hospital-acquired infections (HAI) (Mehtar *et al.*, 2008). Numerous studies have focused on the antimicrobial properties of copper (and its alloys) surfaces mainly in relation to the killing kinetics of a variety of microbial species exposed to different copper alloys. However, although it is becoming increasingly clear that metallic copper has excellent antimicrobial properties the molecular mode of action exerted by copper surfaces and the sensitivity of various cellular targets to its action are still largely unknown (Quaranta *et al.*, 2011). Studies have suggested a role for copper in ion homeostasis as a factor in the survival of bacteria on copper surfaces. Cells of *Pseudomonas aeruginosa*, *Enterococcus hirae*, or *E. coli* which had their copper-ion defence systems deleted have been shown to die more rapidly than wild-type cells when exposed on metallic copper, although the survival times of mutants were not dramatically different from those of wild-type cells (Quaranta *et al.*,

2011). Mehtar *et al.*, (2008) reported that the minimum concentration of Cu to be an effective antimicrobial agent is > 55% for bacteria excluding *Mycobacterium tuberculosis* (MTB). Copper and its alloys showed a marked inhibitory effect on MTB, despite the strains being drug resistant. Growth of both strains showed inhibition by Cu (88–98% inhibition). The above mentioned work shows that the incorporation of Cu in healthcare facilities may dramatically help reduce the environmental microbial burden and act as a useful adjunct to current infection prevention and control systems.

3.3.4.1 Copper pipes

Although copper is a nutrient, when present in large amounts it becomes highly toxic to organisms including hospital pathogens like MRSA. This effect is due to its ability to bring about the rapid fragmentation of a cell's DNA which leads to death; as a result, it is an effective biocide (Warnes *et al.*, 2010). As a result, it is obvious that, wherever possible, plumbing materials in health care settings should be made of copper. As we have seen, hand to hand transfer is the most important means by which microbes, including pathogens, are spread in hospitals. More generalized microbial contamination of the hospital environment also takes place (Noyce *et al.*, 2006) with the result that inanimate surfaces can act as a source of nosocomial infections largely because most Gram-positive and Gram-negative bacteria are able to survive for months on dry surfaces (Hota, 2004), and the main factors influencing the survival of microbes on dry surfaces include exposure to sunlight, humidity, and temperature (Kramer *et al.*, 2006).

Recently, considerable attention has been focused on reevaluating metallic copper as a biocide for use in killing bacteria on so-called “touch surfaces”, which include door handles, bathroom materials, and metallic bed rails (Grass *et al.*, 2011). Weaver *et al.* (2010), for example, evaluated copper, as an alternative to aluminium, as an antimicrobial surface for use in air-conditioning systems and this showed increased pathogen-killing by the former.

The copper content in copper alloys varies from 60% to 99.9%, and initial studies have shown that the survival rates of *E. coli* O157 vary considerably depending on the type of alloy used relative to its copper content (Noyce *et al.*, 2006). These authors noted a gene-mediated response in *P. aeruginosa* to survival on copper alloys and in *E. coli* and *E. faecium* strains increased resistance to copper ions was due to the presence of additional plasmid-borne copper resistance genes. It was noted that the greatest differences in survival rates were related to moisture content in the sample, the type of medium the bacteria were suspended in, and whether it was a Gram-negative or a Gram-positive bacterium. Since copper ion toxicity had been reported previously as a possible driving force in the contact-killing of *E. coli* on copper alloys (Espírito Santo *et al.*, 2008), an experiment was designed to block copper ions released from the copper surface by applying a corrosion inhibitor which would consequently prevent copper ions from entering the bacteria. This was correlated to electrochemical measurements and calculated concentrations of copper released from the surface, and an inverse relationship

was shown between the copper ion concentration released from the surface and survival rates of copper ion-resistant *E. coli*. Subsequently it has been shown that bacterial cells very quickly take up copper ions when exposed to copper surfaces which results in rapid cell death presumably related to increased oxidative stress (Grass *et al.*, 2010; Espirito Santo *et al.*, 2011). Copper surfaces are now being tested in hospitals where their efficacy is compared to stainless steel touch surfaces for long term observation, isolation, and characterization of surviving microbes. However, bacteria will continue to acquire resistance and will continue to proliferate in nature due to pollution -exposure. Here copper resistance can be quite useful, such as in the bioremediation of copper mining wastes (Grass *et al.*, 2011).

CHAPTER 4

ISOLATION OF BACTERIA FROM USED TOOTHBRUSHES AND DETERMINATION OF THE ANTI BACTERIAL POTENTIAL OF TOOTHPASTES

4.1.1 Introduction

The oral cavity can be colonized by more than 700 microbial species, including fungi, viruses and a variety of unclassified microorganisms, most being commensal species that are beneficial for oral health. However, some are pathogens and can overcome the host responses and cause serious oral diseases.

The oral microflora can be divided into two groups:

- **Saprophytes:** These are the permanent microflora of the oral cavity whose presence is needed for the normal functioning of the dental system, as well as the body in general. The saprophytic microflora affects the conditions of the local immune system, prevents the development of pathological conditions and supports the overall bacterial balance.
- **Pathogenic Microflora:** These affect the organs and tissues of the mouth and entire body and bring about the emergence and development of a variety of diseases. This microflora ideally should not be present, or present in very low numbers that fail to substantially affect the oral cavity and the body in general (Otoikhian and Okoror, 2012).

An overload of pathogens in the oral microflora can result in caries, periodontitis and stomatitis, which are the most common oral microbial infections in humans. *Streptococcus spp.* and *Actinomyces spp.* are particularly common colonizers and cause supragingival plaque during the first stage of biofilm formation (Nascimento *et al.*, 2015). They also increase the acidity of the oral biofilm and thereby help to develop their cariogenic potential. After biofilm maturation, anaerobic and proteolytic bacteria, notably *Porphyromonas gingivalis*, *Tannerella forsythia*, *Treponema denticola* and *Fusobacterium spp.* are found to dominate the subgingival biofilm (Nascimento *et al.*, 2015).

Bacterial proteases and metabolic products often induce host responses like inflammation and immunoreactions which result in periodontitis/peri-implantitis. *Candida spp.* are the most common fungi in the oral cavity and often lead to denture stomatitis commonly isolated from infected root canals; additionally, they can act as opportunists in the formation of periodontal and peri-implantar lesions (Nascimento *et al.*, 2015).

Toothbrushes generally become highly contaminated with microorganisms which may arise from the oral cavity and from the environment in which toothbrushes are stored. As a result, toothbrushes may play a significant role in disease transmission and lead to an increase in infection risk (Frazelle and Munro, 2012). They act as a reservoir for microorganisms in both healthy and ill adults and contamination and survival of infectious organisms may occur on both animate and inanimate objects. Contamination of toothbrushes occurs after first use and increases with repeated

use and they can become contaminated from a variety of routes including: the oral cavity, environment, hands, aerosol contamination, and storage containers.

Bacteria may become attached to and accumulate and survive on toothbrushes which may transmit individual disease causing organisms (Frazelle and Munro, 2012).

There is a clear correlation between contaminated toothbrushes and the presence of diseases (Glass and Lare, 1986), and changing a toothbrush regularly has been linked to the elimination of symptoms and disease regardless of its nature.

Moisture is an obvious factor in promoting the survival of bacteria on toothbrushes and immunocompromised patients are at particular risk from their use.

4.1.2. Materials and Methods

Fifty toothbrushes were collected from volunteers aged 5 to 45 years. The toothbrushes were transported to the laboratory in a sterile polythene bag sealed with a rubber band. Brushes were processed within 12h by a method modified from that described by Sammons *et al.* (2004). The handle was cut off using a rotary saw and the head of the brush was retained in the bag to avoid contamination. Each brush head was then subjected to soaking in 10 ml of sterile water, for 20 min, followed by vigorous vortex mixing for 1 min and manual swabbing to dislodge persistently adherent bacteria. The resulting bacterial suspension was serially diluted and 0.1 ml aliquots plated onto nutrient agar to

select for bacteria. Plates were incubated aerobically at 37°C for 24–48 h. Total viable counts were estimated from the numbers of colonies on nutrient agar plates. Colony colour, morphology and haemolysis were recorded and Gram's stain was performed on a representative of each colony morphotype. Extraction of genomic DNA was by using KeyPrep bacterial DNA extraction kit supplied by ANACHEM. Test sample preparation for PCR, DNA quantification, polymerase chain reaction (PCR), agarose gel electrophoresis and phylogenetic analysis, has been explained above.

4.1.3. Results

1) Isolation of bacteria from various used manual toothbrushes by cultivation on Nutrient Agar medium

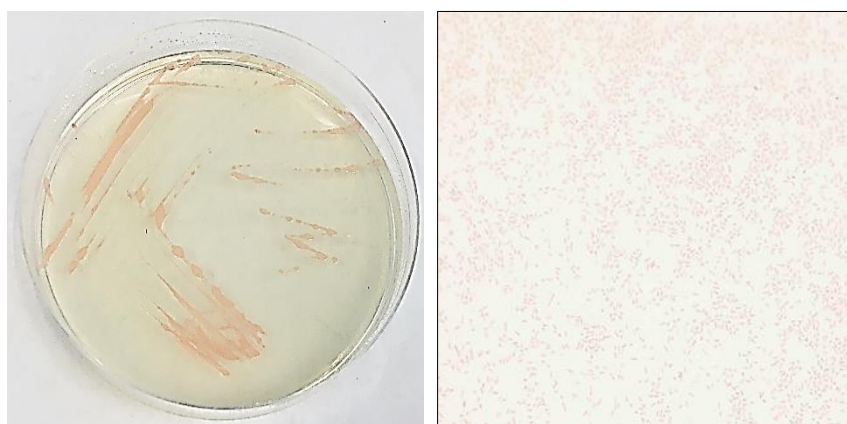
Bacteria were cultured using Nutrient Agar after isolation from used toothbrushes.

Table 4.1: Bacteria isolated from various used toothbrushes.

Representative sequence	Closest matches identification	Sequence identity	NBCI (Accession number)
1 TB	<i>Roseomonas mucosa</i>	99%	KF247232.1
2 TB	<i>Stenotrophomonas maltophilia</i>	99%	LN890169.1
3 TB	<i>Pseudomonas aeruginosa</i>	99%	KR815846.1
5 TB	<i>Leclercia adecarboxylata</i>	99%	KT899848.1
6 TB	<i>Enterobacter asburiae</i>	99%	EU239468.1
7 TB	<i>Candidatus Roseomonas massiliae</i>	99%	KT321690.1
8 TB	<i>Pseudomonas parafulva</i>	99%	KT758848.1
9 TB	<i>Bacillus licheniformis</i>	99%	KU314515.1
11 TB	<i>Pseudomonas aeruginosa</i>	99%	KF680991.1
13 TB	<i>Agrobacterium larrymoorei</i>	99%	EF178437.1
15 TB	<i>Pantoea septica</i>	99%	KF475883.1
16 TB	<i>Stenotrophomonas rhizophila</i>	99%	KP050794.1
18 TB	<i>Citrobacter freundii</i>	87%	CP007557
20 TB	<i>Pseudomonas frederiksbergensis</i>	99%	EU373369.1

2) Light microscope images.

The isolates were examined under a light microscope after the bacteria were identified. (Figure 4.1)



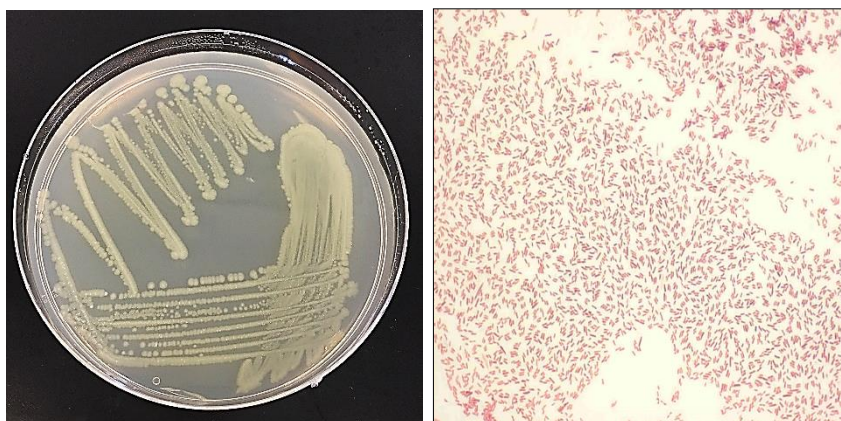
Roseomonas mucosa KF247232.1
Gram stained. Magnification: 100x.



Stenotrophomonas maltophilia LN890169.1
Gram stained. Magnification: 100x.



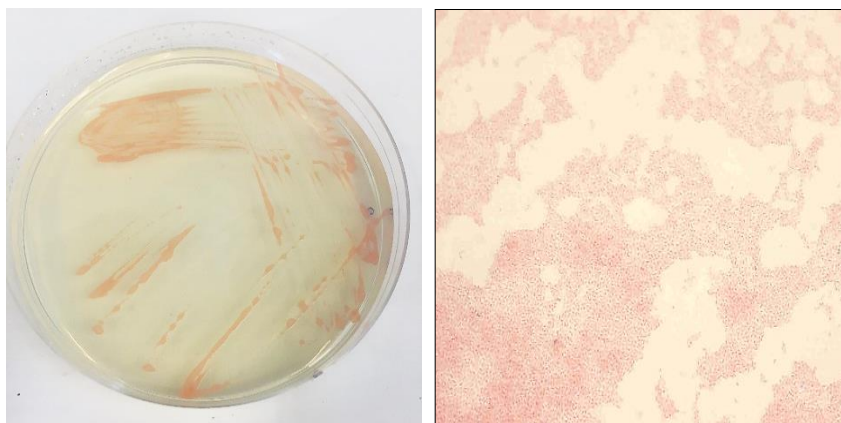
Pseudomonas aeruginosa KR815846.1
Gram stained. Magnification: 100x.



Leclercia adecarboxylata KT899848.1
Gram stained. Magnification: 100x.



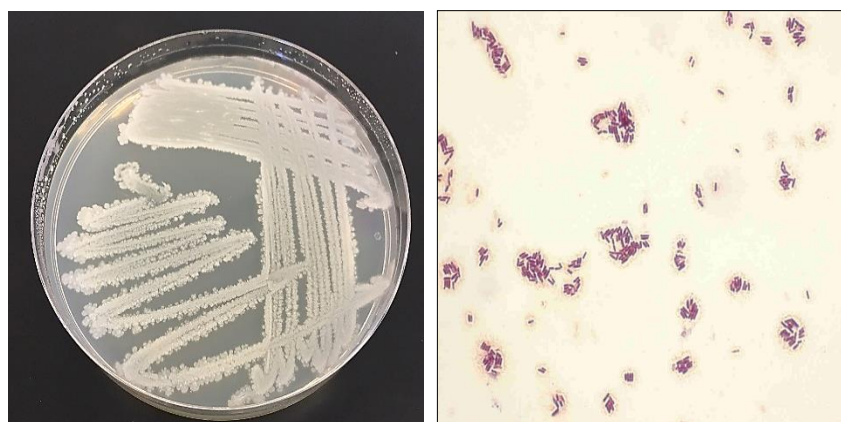
Enterobacter asburiae EU239468.1
Gram stained. Magnification: 100x.



Candidatus roseomonas massiliae KT321690.1
Gram stained. Magnification: 100x.



Pseudomonas parafulva KT758848.1
Gram stained. Magnification: 100x.



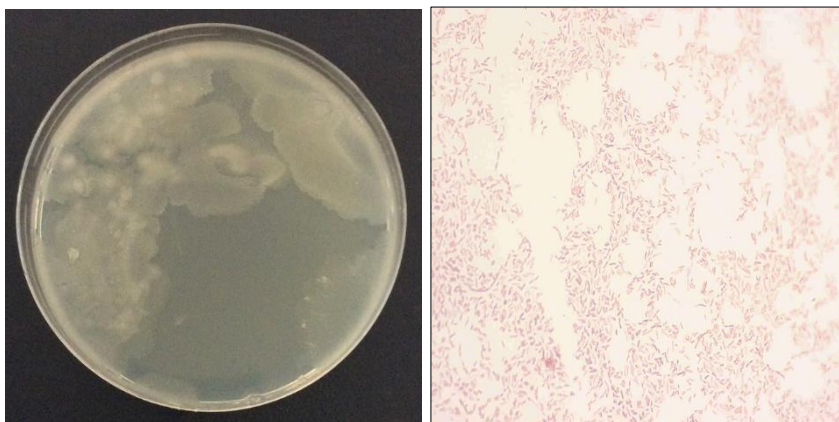
Bacillus licheniformis KU314515.1
Gram stained. Magnification: 100x.



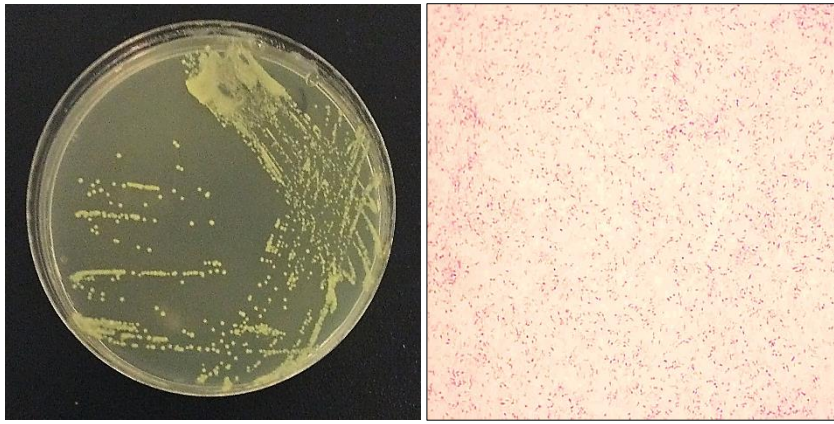
Pseudomonas aeruginosa KF680991.1
Gram stained. Magnification: 100x.



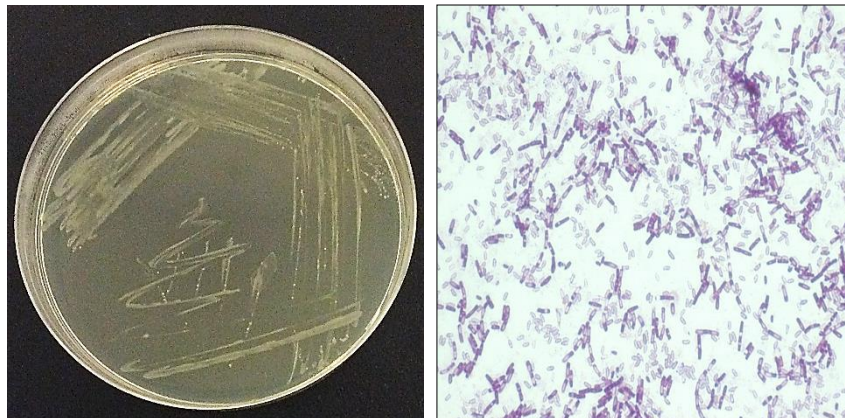
Agrobacterium larrymoorei EF178437.1
Gram stained. Magnification: 100x.



Pantoea septica KF475883.1
Gram stained. Magnification: 100x.



Stenotrophomonas rhizophila KP050794.1
Gram stained. Magnification: 100x.



Citrobacter freundii KP050794.1
Gram stained. Magnification: 100x.



Pseudomonas frederiksbergensis EU373369.1
Gram stained. Magnification: 100x.

Figure 4.1: Nutrient Agar plates showing cultured bacteria isolated from various used toothbrushes, and also microscopy images showing the bacteria under the light microscope.

Enterobacter asburiae

This bacterium is Gram-negative, non-motile and known to cause necrotizing fasciitis and pneumonia. Some *E. asburiae* isolates are identified as human pathogens which are an opportunistic pathogen and causes different human diseases such as community-acquired pneumonia, soft tissue infections, wound infection and other infections (Brenner *et al.*, 1986).

Pseudomonas aeruginosa

Pseudomonas spp., notably *P. aeruginosa*, act as opportunistic pathogens particularly of the lower respiratory tract, bloodstream, and urinary tract amongst patients in intensive care units or which are immunocompromised. It is often isolated from moist areas within health care environments, such as taps, toilets, showers, respiratory equipment, and cleaning solutions. Skin, throat, and faecal carriage amongst healthy individuals have also been reported (Liu, 2011).

Pseudomonas parafulva

It is a Gram-negative environmental bacterium, frequently isolated from rice paddies. Colonies on nutrient agar were smooth, entire, and flat to convex and had a water-insoluble yellow pigment after an incubation of 24h at 35°C. This bacterium rarely causes acute meningitis (Ramirez *et al.*, 2010).

Pseudomonas frederiksbergensis

This bacterium is an environmental organism of no known pathogenicity.

Roseomonas mucosa

Roseomonas species are a recently typified group of pink, slimy, waterborne, Gram-negative coccobacilli. They typically cause fever associated with persistent catheter colonization, and *R. mucosa* has been isolated from various clinical samples, including blood samples, samples obtained from wounds, and samples obtained from the aquatic environment (Bard *et al.*, 2010).

Stenotrophomonas maltophilia

This bacterium frequently colonizes damp surfaces such as tubes used in mechanical ventilation, endoscopes, indwelling urinary and suction catheters. In immunocompetent patients, *S. maltophilia* is a relatively rare cause of pneumonia, urinary tract infection, or bloodstream infection; however, is a growing source of latent pulmonary infections. *S. maltophilia* colonization of with cystic fibrosis patients has been seen to be increasing (Brooke, 2012).

Stenotrophomonas rhizophilia

This is a soil and rhizosphere organism of no known pathogenicity. Cells are straight or slightly curved yellowish rods. Strains are plant associated and isolated

from the rhizosphere of oilseed rape and from the rhizosphere and tubers of potato (Wolf *et al.*, 2002).

Leclercia adecarboxylata

L. adecarboxylata is a motile, Gram-negative rod. It is considered as the sole pathogen causing infection in an immunocompromised woman (Hess *et al.*, 2008). It is considered as normal flora in the gut of animals and it has been isolated from human stools, from the skin of an asymptomatic blood donor and from a variety of environmental sources and drinking water. *L. adecarboxylata* have the ability to infect a variety of bodily fluids; it has been cultured from blood, sputum, peritoneal fluid, urine, synovial fluid, gallbladder tissue, cardiac valve tissue and wounds often as a part of mixed microbial growth in immunocompetent hosts (Tam and Nayak, 2012).

Agrobacterium larrymoorei

This is a soil organism of no known pathogenicity.

Citrobacter freundii

Citrobacter freundii is a Gram-negative, motile, facultative anaerobic bacterium that appear as rods or coccobacilli. *C. freundii* is often the cause of significant opportunistic infections. Also, it has been associated with neonatal meningitis and brain abscesses and with nosocomial infections in the respiratory tract and causes

pancreatic pseudocyst after an acute necrotizing pancreatitis (Badger *et al.*, 1999). It has been distributed in water, soil, food and the intestinal tract of humans. Urinary tract infections (UTIs) caused by *Citrobacter* species have been described in bacterial urine isolates in adults (Metri *et al.*, 2013).

Candidatus roseomonas massiliae

This is an organism of no known pathogenicity.

Pantoea septica

Cells are Gram-negative, motile by peritrichous flagella. *Pantoea* species are recovered from humans and are opportunist pathogens associated with contaminated catheters and penetrating trauma. *Pantoea septica* referring to the septicaemia outbreak associated with these strains (Brady *et al.*, 2010).

4.1.4 Discussion

Tooth brushes can be heavily infected with microorganisms, especially Streptococci, within a day's use and a lack of toothbrush disinfection and care promotes the spread of such pathogens, thereby causing inflammation of the oral tissues (Badger *et al.*, 1999).

Out of the fifty toothbrushes used for this study, none was found to be uncontaminated with bacteria. *Pseudomonas* species were found on four of the fifty toothbrushes, *Stenotrophomonas sp.* and *Bacillus sp.* were found also. A

wide variety of bacteria belonging to different species were isolated from the investigated toothbrushes (Table 4.1).

Contaminated toothbrushes are recognized as a mode for microbial transport, and growth, and can result in the reinfection of a person with pathogenic bacteria or environmental microorganisms. The head area of tooth brush is prone to heavy contamination as fluids and food particles can be drawn by capillary action into the spaces between tufts, a process which may lead to bacterial growth (Frazelle and Munro, 2012).

4.2. Scanning electron microscope studies of the surface of a used toothbrush

4.2.1 Materials and Methods

Scanning electron microscope studies were conducted on the surface of bristles of tooth brushes undergoing normal use. A toothbrush used by the same individual for 6 weeks (alternating night and morning) was processed for scanning electron microscopy (SEM). Bacteria were fixed by immersing the brush head in 2.5% glutaraldehyde in 0.1 M sodium cacodylate buffer, pH7.3, for 60min. Brushes were rinsed in the same buffer, dehydrated in ethanol and critical point dried using liquid CO₂. A segment of brush head with one row of bristles was cut using a rotary saw, mounted on aluminium stubs, gold sputter coated and examined using a JEOL JSM-5300lv scanning electron microscope at an accelerating voltage of 10–30 kV (Sammons *et al.*, 2004).

4.2.2 Results and Discussion

Scanning electron microscopy of biofilms on toothbrushes used for approximately 3 months showed collections of cocci and the development of biofilms on the heads and bristles of both (Figure 4.2). The bristles were rough, providing ample sites for trapping organisms. Examination of a brush that had been used for more than 3 months revealed a biofilm on the brush head. The biofilm showed on the surface of the head to be made up of a compacted community of microorganisms, including cocci, bacilli and filamentous organisms, together with cellular and other debris.

SEM is showed biofilms on the heads and bristles of both conventional brushes, showing that bacteria colonized and grew on them. The rough surfaces of the bristles provide ideal sites for the entrapment of microorganisms and later development of biofilms, whilst nutrients will tend to accumulate at the base of the bristles thereby favouring the development of biofilms. Most dental practitioners recommend that brushes be changed after 2–3 months and following any bout of illness. The example shown here, of a brush that had been used in excess of five months, showed that after such a long period of use, extensive biofilms consisting of mixed communities of organisms developed at the base of the bristles and extended into the head of the brush. This process which could lead to the formation of a reservoir of organisms which can act as a potential source of infection for any vulnerable patient (notably where immunocompromised) or act as a source of cross-infection.

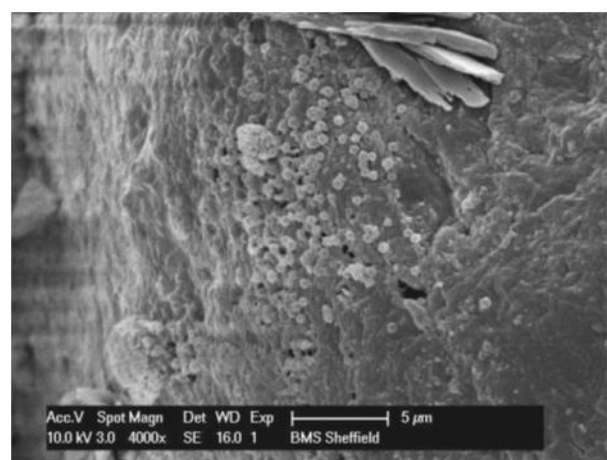
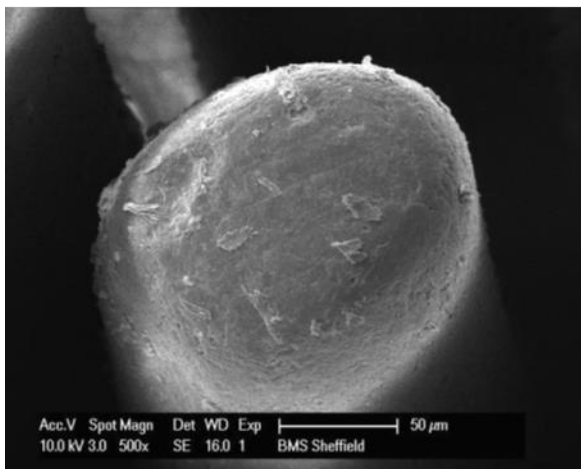
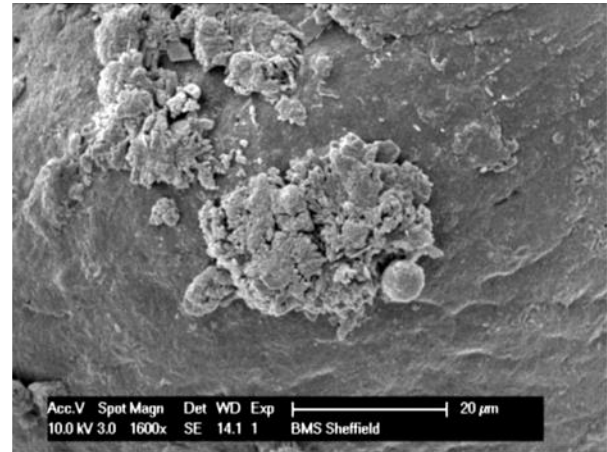


Figure (4.2): Scanning electron microscopy of toothbrush biofilms on antibacterial brushes used for approximately 3 months. Top left) showing debris and cocci bacteria 8000 \times , bar represents 2 μm ; top right) showing debris and biofilm; bottom left) image of tip of bristle, showing indentations and crevasses 500 \times , bar represents 50 μm ; bottom right) cocci embedded in the biofilm, 4000 \times bar represents 5 μm .

4.3. Effect of toothpastes on the bacterial growth

4.3.1 Introduction

Standard toothpastes are usually made up of a mix of fluoride and detergents both of which are generally thought to improve biofilm-control. Surprisingly, however, many toothpaste-based antimicrobials have yet to be effectively tested or proven to work in the oral environment.

Fluoride, in marketed dentifrice is usually comprised of sodium fluoride (NaF) which is a widely recognized anti-caries agent and NaF-containing oral hygiene products which significantly reduce dental caries (Barboza-Silva *et al.*, 2005). Fluoride inhibits a variety of bacterial processes which are mediated by enzyme binding (Barboza-Silva *et al.*, 2005). The main urease producers are *A. naeslundii* and *Staphylococcus epidermidis* while *S. epidermidis* is often the most ureolytically active organism present in plaque. Fluoride directly inhibits bacterial ureases (Barboza-Silva *et al.*, 2005), while chlorhexidine is bacteriostatic and bactericidal against both Gram-positive and Gram-negative microbes (based on its ability to damage cytoplasmic membranes and disrupt cell membrane integrity). Microbial contamination can be effectively removed by immersing them on chlorhexidine overnight.

4.3.2. Materials and Methods

In order to determine the antibacterial effect of toothpastes (fluoride and chlorhexidine toothpastes) a toothpaste agar was used containing 100 ml bacteriological agar and 4 g of a proprietary toothpaste in 9 laboratory bottles. The

basal medium used for isolation and growth for bacteria was bacteriological agar which is composed of (g/l): 70% agarose and 30% agaropectin, final pH 5.7-7.0 at 33-36°C. Suspend 23 g of bacteriological agar in 1000 ml of distilled water. The bacteriological agar was boiled to dissolve the medium completely and sterilized by autoclaving at 15 lbs. pressure (121°C) for 15 minutes. The melted agar was poured in half of Petri plates and left them to be cool and set. For preparation the toothpaste agar: bacteriological agar 50 ml was dissolved in 9 Duran laboratory bottles and sterilized by autoclaving, then added 4 gram of Commercial toothpastes in the bottle (Figure 4.3). The bottles were shaken and vortexed at low speed until complete mixing was achieved. The mixed toothpaste agar was poured in the other half of plates and left them to be cool and set. The plates were streaked by model strains of *Staphylococcus aureus* and *Escherichia coli* in both sides of plates over the medium. All plates were incubated in the 37°C for 18-24 hours. After incubation, an ellipse of inhibition was used to determine the effect of toothpaste on growth of bacteria.



Figure 4.3: (left) The commercial toothpastes and (right) chlorhexidine toothpaste all used to prepare toothpaste agar plates.

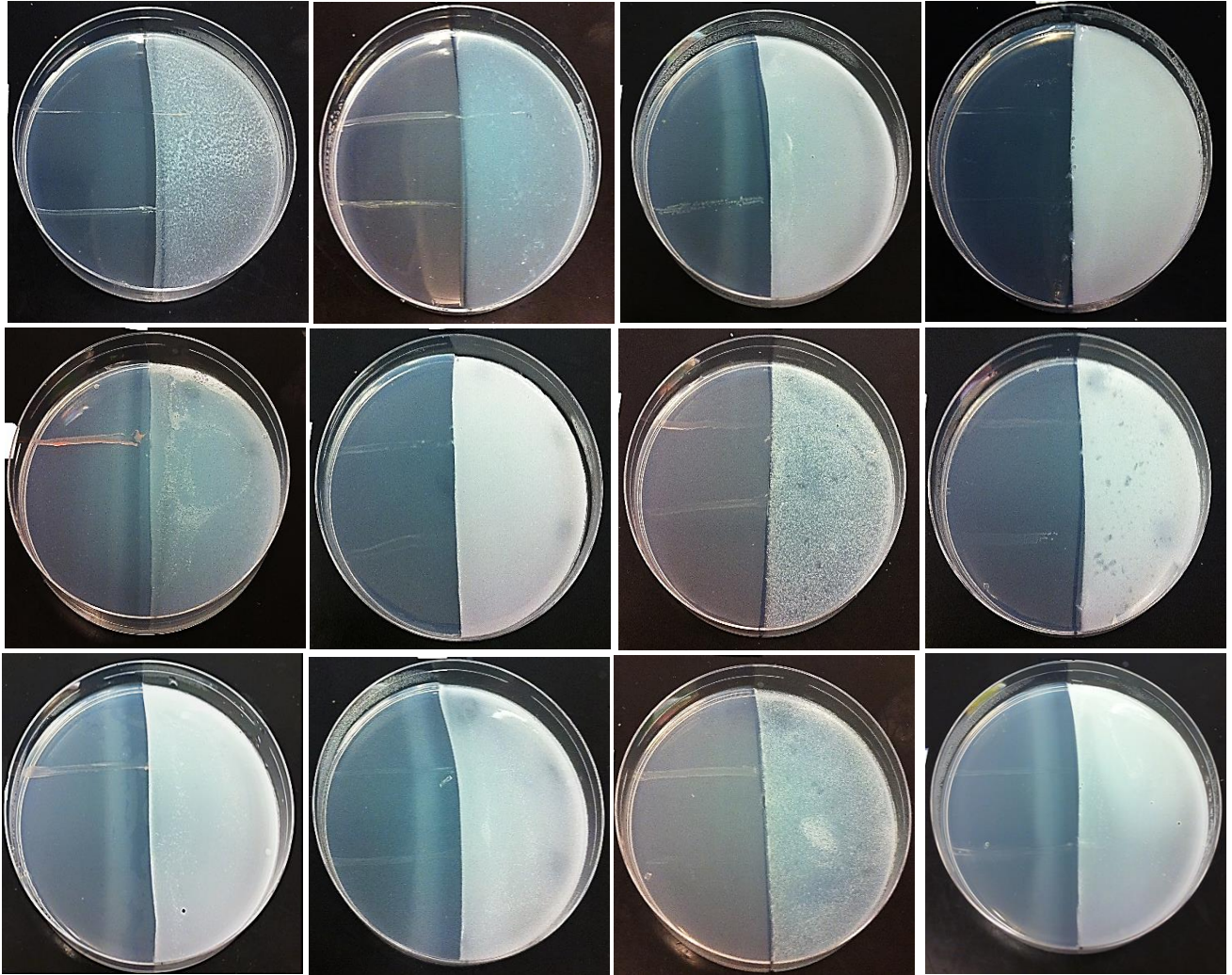


Figure (4.4 A) Half agar plates containing fluoride-containing commercial toothpastes inoculated with bacteria.

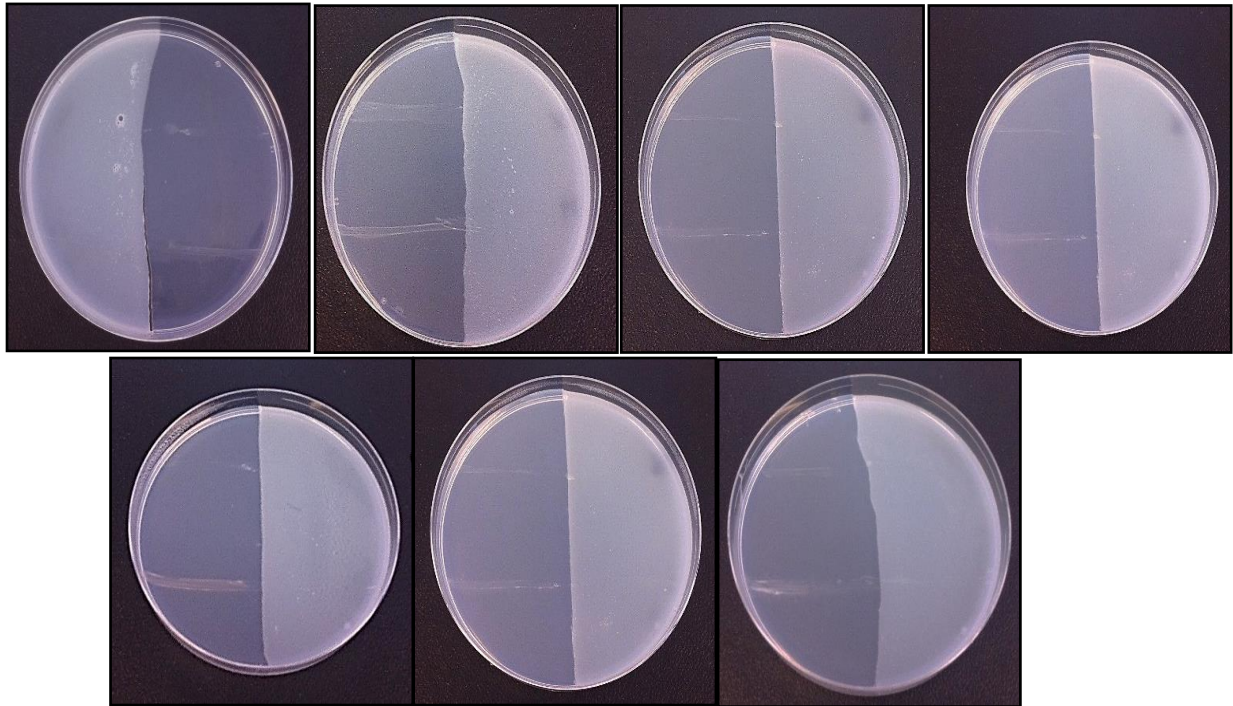


Figure (4.4 B) Half agar plates containing chlorohexidine commercial toothpastes inoculated with bacteria.

4.3.3. Results and Discussion

The results showed that the bacteria isolated from toothbrushes can grow on the agar only side, but no bacteria grew on the toothpaste-containing agar. This result proves that toothpaste acts as an antibacterial agent and can inhibit bacterial growth. The main reason for inhibition is the presence in toothpaste of fluoride which is widely used as an effective anti-caries agent (i.e. antibacterial) agent. Marquis (1995) showed that fluoride can affect bacterial carbohydrate metabolism and that it inhibits the functioning of essential bacterial enzymes; it also reduces the acid tolerance of bacteria and thereby prevents bacteria growth. Fluoridated

toothpastes showed antibacterial activity against *S. mutans* and both aerobic and anaerobic oral flora and also inhibited Streptococci selectively, and inhibited caries formation by interfering with biofilm development by Streptococci (Randall *et al.*, 2015).

Chlorhexidine toothpastes also inhibited bacterial growth when added to toothpaste agar plates. De Rossi *et al.* (2014), showed that chlorhexidine toothpastes presented antimicrobial activity against Gram-positive bacteria and yeasts and that the chlorhexidine molecule has both bactericidal and bacteriostatic antimicrobial effects on the tooth surface. In the present study, chlorhexidine was found to be effective in disinfecting contaminated toothbrushes, although in other studies Listerine was shown to be more effective. The higher efficacy of chlorhexidine could be due to its extended action-spectrum. It is also relatively non-toxic, odourless and is a commonly used mouthwash, properties which make chlorhexidine a good choice for the disinfection of contaminated toothbrushes.

In conclusion, the results presented in this Chapter show that all toothpastes exhibited effective antimicrobial activity against the tested bacteria and that these products were able to inhibit the growth of bacteria. In addition, adding chlorhexidine to dentifrices can result in effective antimicrobial activity against all the evaluated Gram-positive bacteria and yeasts. However, further studies are required to evaluate their clinical advantages in the treatment or prevention of biofilm-mediated diseases.

4.4 Discussion

Again, the importance of these isolates is not that they are major pathogens, but that they can infect immunocompromised patients (Ankola *et al.*, 2009). Toothbrushes get re-contaminated after each use (Frazelle and Muro, 2012) and such re-contamination of the oral cavity can result from the retention of microorganisms on toothbrushes (Filho *et al.*, 2000, Karibasappa *et al.*, 2011). Nascimento *et al.* (2012) showed that new toothbrushes may often carry bacteria even before their use, since no regulatory requirement for pre-use sterilization is generally required. This observation was confirmed in our study, where all (100%) of new tested brushes were contaminated positive for bacterial growth, a finding which explains the high incidence of *Bacillus cereus* and *Candida albicans* on toothbrushes and oral swabs.

The effectiveness of toothbrushes at cleaning, and bacterial plaque removal largely depends on their abrasive qualities, although some toothpastes also contain antibacterial agents (which are often general sterilants) such as hexachlorophene. Toothpastes also contain a combination of fluoride and detergents, compounds which increase their efficiency (Davies, 2008; Marsh, 2010; Prasanth, 2011); triclosan for example, is a low-toxicity, non-ionic, chlorinated bisphenol that is compatible with toothpaste additives like fluoride and surfactants, and it enhances the inhibition of cyclooxygenase/lipoxygenase pathways and also exhibits anti-inflammatory properties (Davies, 2008; Davies *et al.*, 2010). Chlorhexidine is usually regarded as the most effective antimicrobial agent for use in dentistry (Jones, 1997; Twetman, 2004). Its effectiveness is due to the dicationic nature of the

chlorhexidine molecule, which leaves a lasting antimicrobial effect on the tooth surface (Twetman, 2004). Compounds derived from plants also have useful antimicrobial properties which are relevant to toothpaste use (Verkaik, 2011; Pannuti, 2003). A toothpaste called Parodontax for example, is an herb-based product which contains sodium bicarbonate and a number of herbal extracts such as chamomile *Echinaceas* and *Mentha piperita* (Panuti, 2003). For obvious reasons, the use of easily utilizable sugars is avoided as additives to toothpastes, although carbon substances which can support microbial growth are always present in moist and dry sputum.

The results presented show that tooth brushes are contaminated with potentially pathogenic bacteria and that toothpastes act as a microbial nutrient source. As a result, it is highly desirable that only sterile, one time-use tooth brushes should be used by immunocompromised patients. Although modern toothpastes are generally free of readily utilizable sugars, they provide microbial substances in the form of dried sputum and food particles. The antibacterial effect of toothpastes was determined using a readily available family and thus the results showed that pathogenic bacteria failed to grow on bacteriological agar containing toothpastes. The most likely explanation for the inhibitory effect is that the bacteria were inhibited by fluorides which are added to toothpastes as anti-carries agents. Some toothpaste also contains more specific biocides which play a role, alongside

fluorides in inhibiting potentially pathogenic bacteria by inhibiting carbohydrate metabolism and inhibiting essential enzymes (Marquis, 1995).

CHAPTER 5

EFFECTIVENESS OF ANTIBACTERIAL CLOTHS IN INHIBITING THE GROWTH OF BACTERIA

5.1 Introduction

The silver ion has long been known to be effective in inhibiting or killing a broad range of microorganisms and silver is increasingly being used to control bacterial growth in a variety of medical applications, including dental work, catheters, and in the healing of burn wounds (Jung *et al.*, 2008). Slow-release “nanosilver” linings are also used in laundry machines, dishwashers, refrigerators, and toilet seats. The mechanism of the antimicrobial action of the silver ion relates to its interaction with thiol (sulfhydryl) groups. Amino acids, such as cysteine, and other compounds containing thiol groups, such as sodium thioglycolate, neutralize the activity of silver against bacteria (Jung *et al.*, 2008). Silver affects bacterial enzymes and brings about a marked inhibition of bacterial growth with elemental silver often being deposited in the vacuole and cell wall as granules. This element also inhibits cell division, damages the cell envelope, and can denature the contents of the bacterial cell (Jung *et al.*, 2008). Under the influence of silver ions, bacterial cells increase in size, and the cytoplasmic membrane, cytoplasmic contents, and outer cell layers all exhibit structural abnormalities. Finally, silver ions interact with nucleic acids and interact preferentially with the bases in DNA rather than with the phosphate groups (Jung *et al.*, 2008).

5.2 Materials and Methods

The aim of the following experiments was to determine the effect of antibacterial e-cloth (EnivroProducts Kent, TN3 8LE) and a control microfiber cleaning cloth on bacterial contamination (Figure 5.1).

Two types of silver impregnated cloths (antibacterial e-cloth “red colour” and a non-antibacterial microfiber cleaning cloth “yellow colour”) were applied to environmental surfaces (desks, air conditioner, windows, medical equipment, laboratories). Pieces of cloth were moistened with sterile water and wiped firmly over the entire surfaces. Two approaches were used. In the first method, silver containing cloths were used for isolation of various bacterial contaminants from environmental surfaces. Sterile water moistened pieces of cloths were wiped firmly over the entire surfaces. Half the number of cloth was placed in 50 ml of Nutrient broth in sterile tubes, and vortexed for one minute and was left in the shaker for overnight. After 18-24 hours of inoculation, the Nutrient Broth medium became turbid, the turbidity ensuring the presence of certain bacteria in the samples. The other half was placed on the surface of nutrient agar plates and the plates left in the incubator for 18-24 hour. After incubation, the presence or absence of bacterial growth was noted. For morphological identification of bacterial colonies, different types of bacterial colonies appeared on the Nutrient agar plates. Pure colonies of isolates were identified and characterized using standard microbiological techniques. The second method involved using silver containing cloths to determine the effect of silver as an antimicrobial on growth of

bacteria. Sterile water moistened some cloths were placed on the centre of the medium and streaked with cultures of *Staphylococcus aureus* and *Escherichia coli* (i.e. on both sides adjacent to the cloth). These plates were incubated at 37°C for 18-24 hours. Sterile water moistened cloths were placed on the centre of the Sabouraud Dextrose agar plates and streaked with cultures of *Candida inconspicua* and *Candida rugosa* then incubated at 25°C for 5 days. After incubation, the size of any resultant inhibition zone was measured.



Figure 5.1: A) The antibacterial e-cloths and B) non-antibacterial microfiber cleaning cloths.

5.3. Results

5.3.1: Isolation of various bacterial contaminants from environmental surfaces using silver containing cloths on nutrient agar plates and nutrient broth.

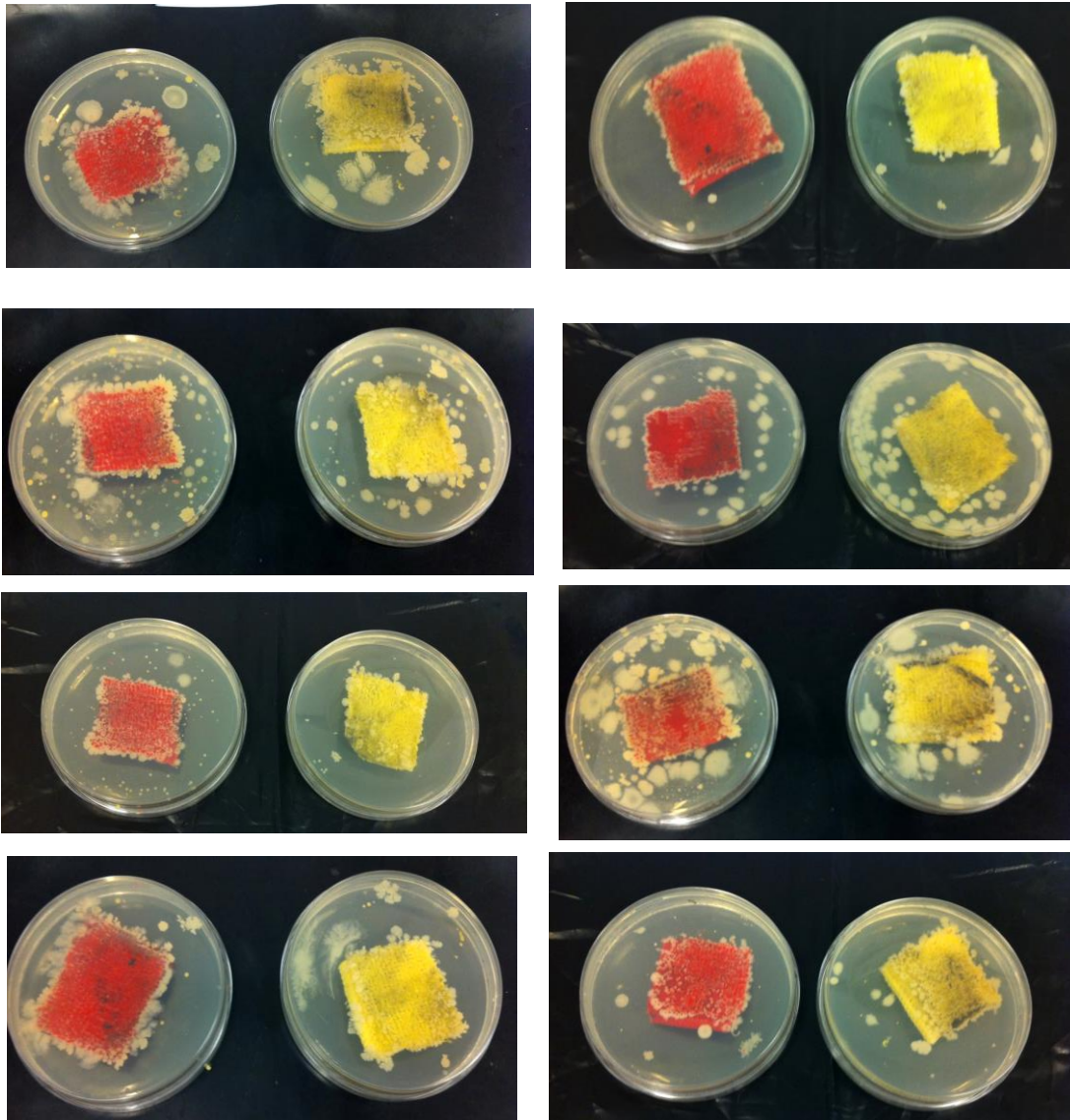


Figure 5.2: Bacterial growth around the antibacterial and control cloth pieces after isolation from different surfaces.

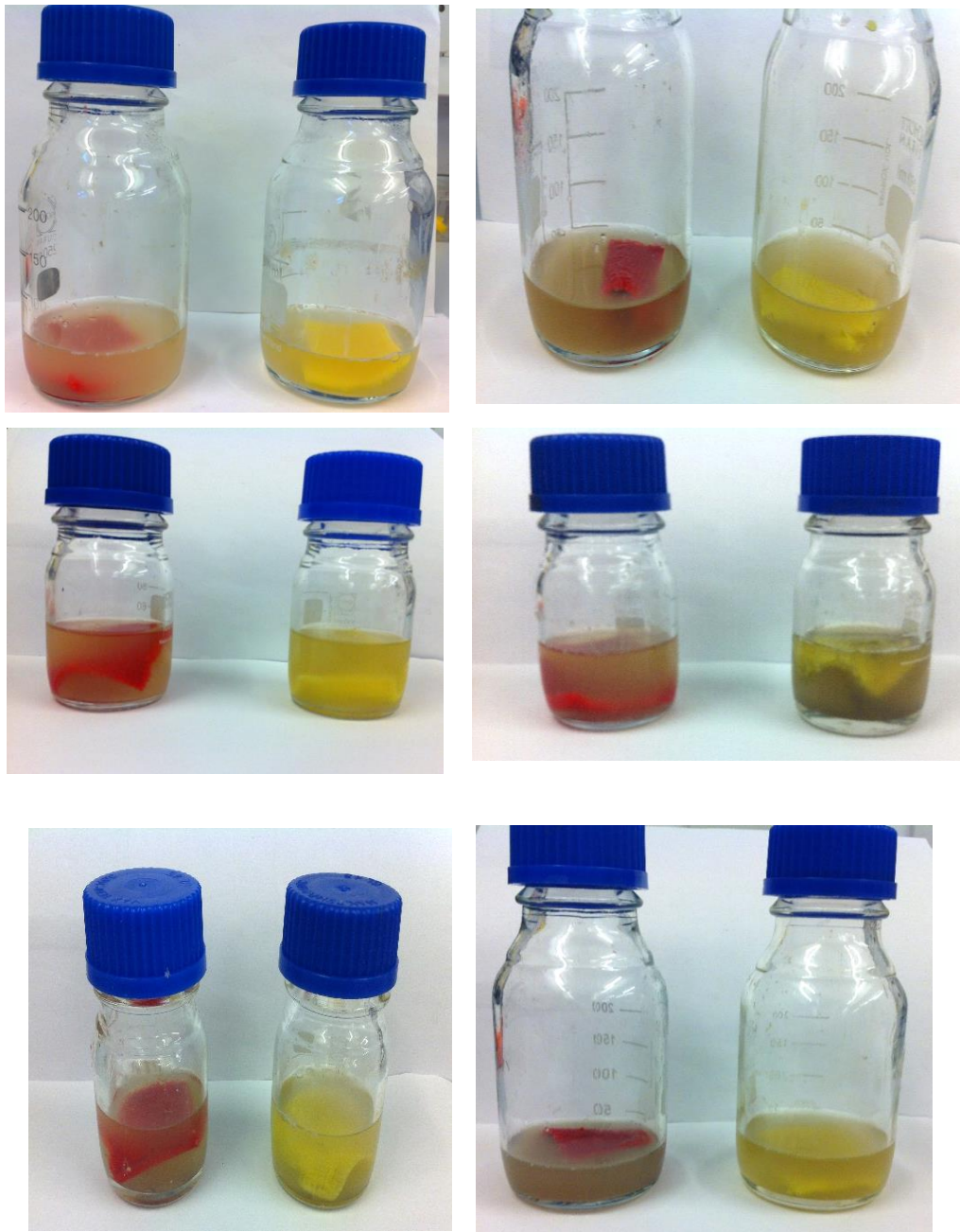


Figure 5.3: Growth of bacteria in the nutrient broth inoculated with the antibacterial and control cloths pieces after isolation from different surfaces.

5.3.2 Examination of the effect of silver as an antimicrobial on growth of bacteria

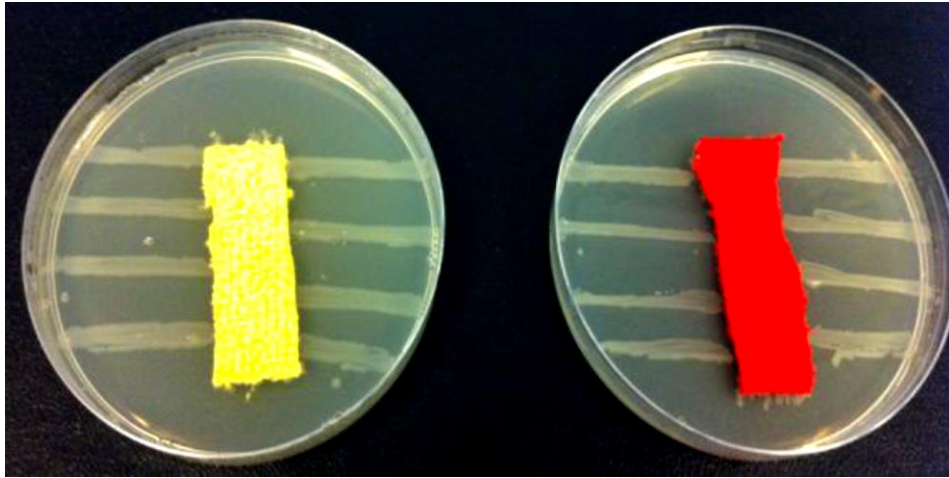


Figure 5.4: No inhibition zones appeared around the antibacterial cloths and the control. The bacteria strain used is *E. coli*; yellow is the control cloth and red is the tested antibacterial cloth.



Figure 5.5: No inhibition zones appeared around the antibacterial cloths and the control. The bacteria strain used is *S. aureus*; yellow is the control cloth and red is the tested antibacterial cloth.

5.3.3 Examination of the effect of silver as an antimicrobial on growth of *Candida* sp.

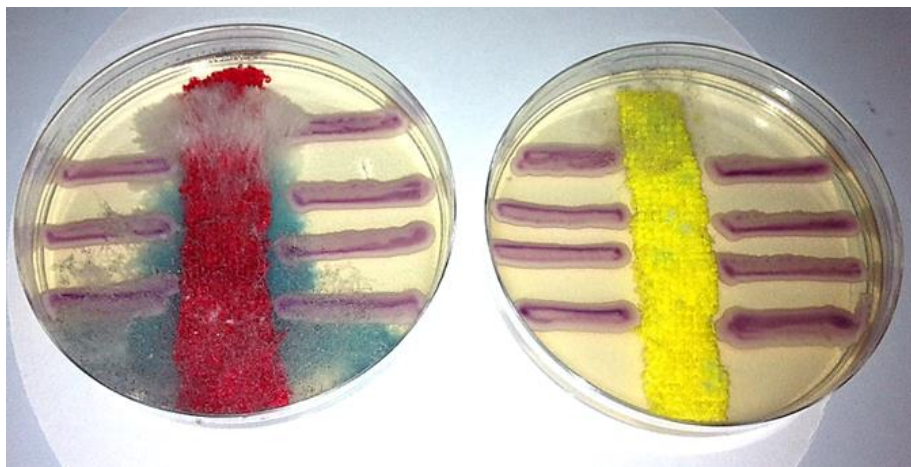


Figure 5.6: No inhibition zones appeared around the antibacterial cloths and the control. The fungal strain used is *Candida inconspicua*; yellow is the control cloth and red is the tested antibacterial cloth.



Figure 5.7: No inhibition zones appeared around the antibacterial cloths and the control. The fungal strain used is *Candida rugose*; yellow is the control cloth and red is the tested antibacterial cloth.

5.4. Discussion

The antibacterial effects of silver containing cloths against bacteria was determined using two techniques with two types of cloths, one an antibacterial cloth containing silver and a non-antibacterial cloth as control. The first method indicated that the growth of bacteria was heavy and there was no effect of silver on bacteria growth. There was no significant difference between the growth resulting from cloths containing silver and control cloths. Both showed the same results (Figure 5.2, 5.3). The bacterial growth was not affected by bactericidal activity resulting from silver.

The silver impregnated cloth used here was wiped on various surfaces and, not surprisingly, became contaminated with dust and dirt and, when this was transferred to growth medium, it supported bacterial growth. The antibacterial cloth did not prevent the growth of bacteria when incubated adjacent to the organisms on a solid growth medium. This result is entirely unexpected, since the cloth used here is marketed as an antibacterial material based on its silver content and its claimed ability to act as a broad spectrum micro-biocide capable of inhibiting bacteria and fungi including MRSA, and other antibiotic resistant species (Gupta and Chauhan, 2016). It is likely that the concentration of silver present in the cloth is too low to be capable of inhibiting bacteria and it appears that the antibacterial cloth, used here, would be of little use for controlling pathogens on surfaces in household and hospital settings.

The second method showed that no inhibition zones appeared around the antibacterial cloths, so the growth of bacteria was not affected by the sliver on cloths as shown in (Fig 5.4 ,5.5). Also, the antibacterial cloth did not prevent the growth of two yeasts when they were incubated adjacent to the material on a solid medium (Figure 5.6, 5.7). This result is both surprising and worrying, since it shows that cloths which are advertised and sold to kill bacteria do not do so. The result may be due, however, to the fact that heavy bacterial loads were used in this study. Further work will continue using lower loads.

It is known that the enhanced antibacterial effect of nanoparticles is due to their large surface to volume ratio, and therefore the smaller the particle, the greater this effect would be. Yu *et al.* (2013) reported that nanocomposites of larger sized AgNPs were much less cytotoxic than the smaller ones, without sacrificing the antibacterial potency of smaller particles.

The mechanism behind the antimicrobial potency of Ag nanoparticles is still not clear. It is believed that their antibacterial efficacy could stem from their absorption by bacterial cells, resulting in the shrinkage of the cytoplasmic membrane (Pattabi *et al.*, 2013). An alternate mechanism proposes that due to the interaction of Ag ions with the S-H bonds in proteins, the DNA loses its capacity to replicate and proteins become deactivated (Pattabi *et al.*, 2013).

CHAPTER 6

SURVIVAL AND RELEVANT METABOLIC DIVERSITY OF ISOLATES

6.1 Introduction

Bacteria play a major role in the cycling of elements in the outdoor and built environments, being involved in all of the major cycles, including participating in transformations of C, N, P, S as well as modifying elements such as copper, manganese and iron (Killham, 1994). The aim of the work reported below was to determine the ability of some of the bacterial isolates to hydrolyse urea, nitrify, oxidize reduced form of sulphur, solubilize insoluble phosphate and oxidise copper and iron sulphides. Some of these transformations are particularly relevant to healthcare environments, such as the hydrolysis of urea (present in urine) or the oxidation of copper and iron sulphides which are found in corroded hospital, metal pipes. Other transformations, such as the formation of thiosulphate may also enable bacteria to better survive in the medical environment by protecting them from the toxic effects of, for example, heavy metals, or even possibly biocides. The following is a brief survey of the transformations which occur in mineral cycling which are relevant to the work described here. Further details on these transformations can be found in the following references: Alexander (1977), Maier *et al.* (2009), Killham (1994) and Paul and Clark (1989).

Urea hydrolysis: Urea is water soluble and has a high nitrogen content exceeding that of ammonium nitrate and ammonium sulphate. Microbes secrete

ureases which converted urea to carbon dioxide and ammonia. A wide range of bacteria can mediate this process including species of *Pseudomonas*, *Achromobacter*, *Bacillus* and *Micrococcus*.

Nitrification: Nitrification is of major importance for the N-cycle in aquatic and terrestrial environments. It involves the oxidation of ammonium (NH_4^+) to nitrite (NO_2^-) and then nitrite to nitrate (NO_3^-) by chemoautotrophic bacteria and by some heterotrophic fungi and bacteria, which can also perform these oxidations. Two kinds of nitrification exist (Killham, 1994): The first involves the activity of chemoautotrophic nitrifying bacteria (*Nitrosomonas*) by which ammonia (NH_3) or ammonium (NH_4^+) ions are oxidised to nitrite (NO_2^-). The second part of the process involves chemoautotrophic Gram-negative bacteria which oxidize nitrite (NO_2^-) to nitrate (NO_3^-). *Nitrobacter* and some fungi such as species of *Penicillium* and most other *Deuteromycetes* can also perform these oxidations (Maier *et al.*, 2009).

Sulphur oxidation: Sulphur is an essential element for growth of all organisms, being a required element for the synthesis of the amino acids, cysteine and methionine, and vitamins such as vitamin B1 (thiamine), hormones such as biotin, coenzymes and lipoic acid. Filamentous fungi play a role in the S- cycle; for example, *Fusarium solani* (a soil fungus) oxidizes elemental sulphur to polythionates, thiosulphate and sulphate. Fungi oxidize sulphur to sulphate with the formation of tetrathionate and thiosulphate. These products, it has been

suggested, may protect fungi from the toxic effects of heavy metals (Alexander, 1977).

Phosphorus solubilisation: Bacteria and fungi are able to solubilise phosphate and release P when growing *in vitro* with calcium phosphate, apatite or other sources of insoluble phosphate; phosphate solubilizing fungi include species of *Aspergillus*, *Fusarium*, and *Penicillium* (Paul and Clark, 1989). A wide variety of heterotrophs solubilize insoluble phosphate including tricalcium phosphate, dicalcium phosphate, hydroxyapatite, and rock phosphate (Altomare, 1999). Bacteria which participate in the reactions include: *Micrococcus*, *Aerobacter*, *Flavobacterium*, *Erwinia*, *Pseudomonas*, *Bacillus*, *Rhizobium*, *Burkholderia*, *Achromobacter* and *Agrobacterium* (Kim *et al.*, 1997,1998). The mechanisms of solubilisation of insoluble phosphate are based on acidification of the medium following organic acid production.

Microbial oxidation of metal sulphides: A wide range of bacteria and fungi have the ability to oxidize metal sulphides, some being chemoautotrophs such as members of the genus *Thiobacillus*, while others are common heterotrophs. The final product of the oxidation reactions involved is sulphate which occurs as sulphuric acid (Killham, 1994). This sulphuric acid can be corrosive to metals, but on the positive side, can be used in ore leaching to obtain metals.

The biogeochemical processes mentioned above and studied here have generally been associated with chemolithotrophic bacteria and the potential role of

heterotrophs in their mediation has generally been underestimated, but are increasingly being shown to play a crucial role in environmental cycling (Killham, 1994). Heterotrophic nitrification and S-oxidation may not benefit organisms in relation to direct end product formation, but through the indirect benefits gained by a heterotroph from participating in these processes. The production of polythionates during S-oxidation by fungi, for example, may help protect these organisms from the toxic effects of heavy metals and other toxicants.

6.2 Materials and Methods

Growth of bacteria and analysis of ions

The bacteria were grown on nutrient agar plates and inoculated into 10 ml of Nutrient Broth containing 0.1% w/v substrate in sterile Falcon Tubes (100ml); the tops were loosened to allow for gas exchange and then incubated with vigorous shaking for 7 days at 37°C. The contents were then allowed to settle and an aliquot was transferred to a HP-1050 DAD HPLC SYSTEM for analysis of resultant ions, or dipped with the relevant ion dipstick (Figure 6.1).



Figure 6.1: (A). Dipsticks container with dipsticks used for ion determination showing concentration chart, (B), Dipsticks used for ion determination showing concentration chart set used for determination of phosphate.

6.3 Results and Discussion

A comment on Dipsticks as a method for ion analysis. The measurement of ions (which are important in environmental transformations) in this laboratory (such as nitrate, phosphate and sulphate) has generally involved the use of colorimetry. These approaches have worked extremely well and, as result have proved to be very useful; of late however, it has been turned to the use of analytical-ion Dipsticks which are less expensive and are generally less- influenced by interference. They also do not involve the preparation and use of dangerous chemicals (for example when they are used to replace corrosive chromotropic acid in the analysis of nitrate) and can be used to rapidly screen a wide number of

samples. This replacement of chromotropic acid with Nitrate-Dipsticks provides a good example of the advantages in changing this analytical approach. This analytical reagent is based on concentrated sulphuric acid (Simms and Jackson, 1971) and is therefore highly corrosive and is dangerous to prepare and use. Its use as an analytical reagent is also hindered by the fact that, as well as nitrate, it reacts with sugars, when these are present to give a pink-purple colour which interferes with the reading of the normal yellow colour produced by the interaction of chromotropic acid with the nitrate ion. Because no acids are used when nitrate-ion Dipsticks are used, and because no colour interference occurs using this approach, being quicker and cheaper in the long run, is regarded as an improvement on the use of chromotropic acid.

The only disadvantage with using Dipsticks is that some accuracy is sacrificed. In most cases, however, all that is needed from a nitrate ion analysis is a rapid indication of whether or not the ion in question is present or a relatively accurate estimate of its concentration, both of which can be provided using Dipsticks.

Tests, by others in this laboratory have shown that results from Dipsticks are between 5 percent (plus or minus) of the values achieved using colorimetry, as was previously used.

The results show that all of the bacteria hydrolysed urea to ammonium and were able to oxidize ammonium to nitrate, via nitrite (Table 6.1). The most active urea hydrolyzer was isolate 7, followed by 5 and 4, while isolate 10 showed the least activity in this respect (Table 6.1). All isolates also oxidized ammonium to nitrate

with the formation of small amounts of nitrite as intermediates. Since the oxidation of nitrite to nitrate is rapid, the former ion rarely appears at high concentration in the environment (Killham, 1994). In terms of nitrate production, bacterium 7 was the most active, followed by 10 and 5, while bacteria 1 and 2 were the least active (Table 6.2).

Table 6.1. Hydrolysis of urea to ammonium.

Substrate-Urea	Ammonium (μgml^{-1})
1	27.5 (1.9) 30
2	25.3 (2.7) 15
3	26.7 (0.9) 25
4	30.6 (3.6) 30
5	43.6 (5.2) 50
6	13.1 (2.1) 15
7	50.9 (2.3) 60
8	10.8 (8.6) 5
9	17.3 (1.9) 20
10	4.4 (0.2) 10

1) *Bacillus licheniformis*, 2) *Bacillus subtilis*, 3) *Bacillus thuringiensis*
 4) *Enterococcus mundtii*, 5) *Citrobacter freundii*, 6) *Pseudomonas luteola*
 7) *Arthrobacter sanguinis*, 8) *Klebsiella oxytoca*, 9) *Kocuria rhizophila*, 10) *Rothia amarae*.

Figures in **bold** relate to Dipstick analysis. Means of triplicate flasks (+/- standard deviation), 7day incubation at 37°C, values in excess of control value.

Table 6.2. Oxidation of ammonium to nitrite and nitrate.

Substrate-Urea		Product ($\mu\text{g ml}^{-1}$)	
Nitrite		Nitrate	
1.	1.8. (1.9) All- trace	5.2 (0.66)	8
2.	0.7 (0.1)	3.2 (0.8)	Trace
3.	0.9 (0.9)	11.9 (1.2)	10
4.	1.4 (0.60)	15.9 (2.0)	20
5.	0.3 (0.0.2)	30.2 (0.9)	25
6.	1.1 (0.1)	10.2 (0.1)	15
7.	0.5 (0.2)	50.3 (07)	35
8.	0 (0.1)	7.9 (1.3)	10
9.	1.9 (0.1)	20.9 (1.7)	20
10.	1.4 (0.2)	30.2 (5.4)	50

1) *Bacillus licheniformis*, 2) *Bacillus subtilis*, 3) *Bacillus thuringiensis*
4) *Enterococcus mundtii*, 5) *Citrobacter freundii*, 6) *Pseudomonas luteola*
7) *Arthrobacter sanguinis*, 8) *Klebsiella oxytoca*, 9) *Kocuria rhizophila*, 10) *Rothia amarae*.

Figures in **bold** relate to Dipstick analysis. Means of triplicate flasks (+/- standard deviation), 7day incubation at 37 °C, values in excess of control value.

All of the isolates were able to solubilize insoluble phosphate, notably isolate 7 followed by isolate 5, with isolate 10 being the least effective. Similarly, all isolates were able to oxidize iron sulphide to release Fe^{2+} and oxidize copper to sulphide to release Cu^{2+} ions. All of the isolates, with the exception of isolate 2,

released similar amounts of Fe²⁺, while isolate 7 and 5 were particularly active at oxidizing copper sulphide (Table 6.3).

Table 6.3. Solubilization of insoluble P and oxidation of FeS and CuS.

Substrate-			Product (µg ml ⁻¹)		
Insoluble-P to Phosphate			FeS to Fe ²⁺	CuS to Cu ²⁺	
1	26.5 (1.3)	30	3.1 (0.9) All- trace	4.2 (1.6)	4
2	5.3 (2.7)	15	1.7 (0.2)	3.6 (1.8)	4
3	4.7 (1.9)	20	4.9 (1.9)	5.9 (1.2)	7
4	25.6 (4.6)	30	4.4 (1.6)	14.6 (1.8)	10
5	45.6 (4.2)	40	3.0 (1.2)	23.2 (1.9)	20
6	18.1 (1.1)	10	5.0 (0.5)	10.4 (1.1)	10
7	45.9 (1.3)	50	3.5 (0.2)	25.3 (2.7)	15
8	20.8 (8.5)	15	4.0 (0.9)	17.9 (1.5)	20
9	15.3 (1.6)	15	5.0 (0.2)	12.9 (1.7)	10
10	6.4 (1.2)	5	2.4 (2.2)	12.2 (1.4)	30

1) *Bacillus licheniformis*, 2) *Bacillus subtilis*, 3) *Bacillus thuringiensis*
 4) *Enterococcus mundtii*, 5) *Citrobacter freundii*, 6) *Pseudomonas luteola*
 7) *Arthrobacter sanguinis*, 8) *Klebsiella oxytoca*, 9) *Kocuria rhizophila*, 10) *Rothia amarae*.

Figures in **bold** relate to dipstick analysis. Means of triplicate flasks (+/- standard deviation), 7day incubation at 37°C, values in excess of control value.

The results obtained using proprietary Dipsticks were in most cases similar enough to this obtained using HPLC to point to their use in experiment where an exact, but reasonably close figure for an ion-concentration is required. As mentioned above, this loss of accuracy is acceptable for convenience, low cost and safety of using Dipsticks instead of the colorimetric assay. The results show that the bacteria isolated from healthcare environments are capable of participating in some important metabolic process, whether such participation is relevant to such environments, especially where carbon is limiting, is not immediately clear. Despite this, the experience of conducting these experiments was, in any case, highly valuable.

CHAPTER 7

FINAL DISCUSSION

The work provided in this Thesis clearly shows that bacteria and other microbes commonly contaminate everyday objects and surfaces, a fact which does not generally cause problems in normal life (except for the occasional food poisoning episode). Such contamination does, of course, have major implications in health care settings such as hospitals and in particular for immunocompromised patients. These patients have their immune systems inhibited either intestinally (e.g. during organ transplantation), or as the result of a natural intervention, such as AIDS. The microbes which cause these infections are often generally not pathogenic and the immune systems of non-immunocompromised patients can generally cope with them, even when the patient is otherwise ill. The stark fact is that any microbe can act as pathogen in people with a compromised immune system. It is therefore extremely important to make certain that all surfaces in health care settings are as clean and microbe-free as possible and that this hygiene status includes non-pathogens as well as the generally recognized pathogens. Such a relatively microbe-free environment can be achieved using biocides, but there is no substitute for thorough “deep cleaning”.

The cleaning of hospital surfaces is the main defence against the threat of antibiotic resistant bacteria and nosocomial infections. While the importance of deep cleaning is universally recognized, this does not mean that such cleaning programmes are theoretically effective or always well-managed. Bleach is by far the most commonly used biocide in both hospital and domestic settings and, when used correctly, can be

extremely efficient and act as an effective standby for all disinfecting purposes.

Although it is more expensive, hydrogen peroxide provides a useful alternative to bleach especially for use on so-called “non-critical surfaces” since it is fact acting and exhibits a broad spectrum antimicrobial activity against bacterial and fungal spores and viruses; steam vapour systems and microfiber- containing biocides can also be effectively employed for routine cleaning purposes.

An essential requirement for successful disinfection is the provision of an adequate contact time between a disinfectant and the object being disinfected. Such contact times vary in addition to a number of environmental factors, notably the degree of microbial loading. Pathogens, and other microbes, can be transmitted in a number of ways in the hospital environment, including:

Droplet contact transmission: A large number of diseases are spread inside droplets which make contact with the eye, nose or mouth. Examples of these droplet infections include SARS, the common cold, Legionnaires' disease and MRSA.

Such infected droplets are produced by infected persons when coughing, sneezing or talking.

Airborne Transmission: Suspended dust particles which contain microbes often remain in the air over long periods and from here can gain entry into the upper and lower respiratory tracts and thereby bring about infections like Aspergillosis, chickenpox and measles.

Faecal-oral transmission: Pathogenic and non-pathogenic microbes infect patients following the consumption of food and water which has been contaminated with

faeces. Bad hygiene and poor sanitation allow such microbes to contaminate food, water and environmental surfaces. These organisms then multiply inside the digestive system then released from the body in faeces and urine, so that the infection-cycle then begins again. Diseases which are disseminated in this way include: rotavirus, hepatitis A virus, *E. coli*, *Cryptosporidium*, *C. difficile*, *H. pylori* and *Candida*.

Direct contact transmission: This disease-transmission process involves the direct physical contact between an infected person and the new host; examples include, kissing and sexual intercourse, or merely close contact. Diseases which are spread by direct contact transmission include H1N1 virus and hepatitis A virus, *Acinetobacter*, *E. coli*, SARS, the common cold, ringworm and other yeast infections, scarlet fever, norovirus, foot and mouth disease, *H. pylori*, and MRSA.

Indirect contact transmission: This process of disease spread takes place when a susceptible person makes direct physical contact with the body surfaces of an infected person via their hands, and then touches their own face, eyes or mouth, allowing the pathogen to gain access into the body and initiate infection. Organisms which use this transmission route include: *rotavirus*, hepatitis A virus, influenza, the common cold, *H. pylori* and tuberculosis, norovirus, *C. difficile*, MRSA, SARS, *E. coli*, *Cryptosporidium*, ringworm and other yeast infections and scarlet fever. The fact that these microbes can survive and be replenished on body surfaces for long periods demonstrates the obvious need for hand washing and other aspects of body hygiene amongst patients, staff and hospital visitors.

Use of disinfectants and detergents in healthcare settings

The above discussion demonstrates the clear need for effective sterilizing agents to reduce the pathogen load on surfaces in hospitals. The use of such agents varies based upon factors such as efficiency and cleaning ability, environmental impact, cost, and associated toxic effects.

Detergents

Detergents generally need effort in regard to rubbing and scrubbing to be fully effective but are good at removing chemicals, food spills, and other commercial wastes. They are not, however, generally suitable for use in cleaning blood or other body fluids from contaminated surfaces and are also usually not effective against bacterial spores, particularly those of *C. difficile*.

Sterilizing agents

Sterilizing agents are generally expensive and highly toxic and are therefore not used in routine household or day to day use. They are however, effective on so-called “critical surfaces” in hospital, industrial and laboratory settings. These agents are however, very effective at killing pathogens, including spores of *C. difficile*, and are particularly effective for use in disinfecting and removing blood and other contaminating body fluids; they include agents such as chemicals, heat, irradiation, filtration and high pressure sterilization.

Guidelines for use in the decontamination of healthcare associated surfaces

Consideration needs to be given to the sterilization of the numerous surfaces found in health care environments, simply because each site varies in its surface properties and the environmental conditions to which it is exposed. An obvious example is provided by the differing conditions found on the surface of an oven compared to that of a refrigerator.

Non-critical medical surfaces

These areas can be adequately served by routine, low maintenance, detergent-based cleaning which will prevent the transmission of microbes from non-critical surfaces, such as furniture and floors. The cleaning process should begin the removal of soil and other debris by wiping or deep scrubbing before a cleaning/disinfecting agent is applied. The survival of bacteria such as *Staphylococcus* spp., MRSA, *Acinetobacter*, and other airborne fungi is encouraged by the presence of dust on surfaces and damp-dusting with cloths wetted with detergents recommended for use on such non-critical surfaces, followed by a period thorough drying. All objects within the ward or room including radiators, air conditioning units, fans, switches, sockets and of course computer areas should be thoroughly wiped with disinfectant-rich cloths. Doors, which include their edges, should be cleaned and special attention should be given to areas which are frequently touched such as handles and door-push plates. In order to control Legionnaires' disease, any infrequently used taps and shower heads should be run weekly for around 10 minutes and all air conditioning and ventilation grills and

associated extractors and inlets should be dusted at least weekly and should be fully cleaned on a yearly basis. Walls and ceilings however, need to be washed only every 6 months or so, by the use of hot water and detergent.

The following factors affect the choice and use of disinfectants. Attention should always be given to the most important factor in their use, namely contact–exposure time:

- The need to comply with the appropriate chemical safety regulations.
- Some disinfectants corrode or discolour surfaces, so compatibility should be considered and compatibility tests conducted.
- The required antimicrobial activity- this relates to the need to determine if the proposed has a wide antimicrobial spectrum?
- Contact times- the necessary contact time needed to kill nearly are hundred percent of the pathogens present needs to be determined by laboratory tests.
- Is the disinfecting agent effective when organic matter is present? This is a very important point because organic matter rapidly deactivates many otherwise potent disinfecting agents?
- Storage and disposal -Is the agent stable when stored for long periods? This is an important point because such agents are often bought in bulk to reduce costs and are stored for long periods; storage should of course be done at room temperature in the dark. Issues regarding the disposal of an agent and its environmental toxicity obviously need to be considered; some healthcare authorities may wish to impose ethical environmental tests on the sterilizing products they employ.

- Environmental factors- Do common environmental factors such as temperature and pH affect the disinfectant's effectiveness?
- Costs-Finally, cost is an essential factor especially since healthcare facilities worldwide are increasingly being effected by Government cost-cutting.

Disinfectants used in routine cleaning of healthcare environments

The following is a description of the most frequently used, EPA-registered disinfectants:

- Phenolic and iodophore-based germicides.
- Quaternary ammonium germicidal detergents.
- Hydrogen peroxide (3-7.5%).
- Ethyl or isopropyl alcohol (70-90%); sodium hypochlorite, i.e. bleach (5.25-6.15% household bleach diluted 1:500 gives >100 ppm available chlorine).

Environmentally safe disinfectants

So-called environmentally safe disinfectants include baking soda, vinegar, eucalyptus oil, grapefruit seed extract borax, liquid detergent, alcohol and tea tree oil. With the exception of undiluted vinegar and eucalyptus oil, most of these disinfectants do not kill *E. coli*.

Antibacterial coating of surfaces

Antimicrobial coatings are available for use on commonly used domestic fabrics, including, linen (curtains); furniture (chairs and tables); office equipment (computers and printers); hand-touch sites (handles and water taps) and generally used surfaces such as walls, floors, walls and doors. Any product that can be impregnated or coated with a micro-biocidal paint or chemical can be referred to as being 'antibacterial'. Such bioactive surfaces or coatings generally provide heavy metals, antiseptics and biocide to the surface undergoing treatment. Nano-silver particles, like titanium dioxide, are particularly useful since they form an invisible protective nano-coating which can act as a very effective biocide for use on a wide range of surfaces, even under extreme temperatures.

CONCLUSIONS

A wide range of bacteria were isolated during the course of this study from computer keyboards, sinks, used toothbrushes and vacuum cleaner dust. Bacteria were also shown to be emitted by toilet hand dryers; these are becoming increasingly popular in public buildings because of reasons of cost and convenience. The results of work presented here shows, however, that paper towels should be used in healthcare environments instead of these dryers, simply because hand dryers can release potential pathogens into the surrounding air, into what is often a small, enclosed space, with little in the way of ventilation. Studies are also reported on the survivability of bacteria on smooth and rough tiles, essentially the same similar to those found in homes and

healthcare settings. Seemingly paradoxically, bacteria were shown to survive for longer on rough, compared to smooth tiles as well as on plastic plumbing fittings than on copper fittings. These findings confirm those of other works, by showing that copper fittings are desirable over plastic fittings because they dramatically reduce the survival of surface-growing, potential pathogens. The results presented here also show that bacteria are important nosocomial pathogens and, since they have been shown to be widely isolatable from the hospital and other indoor environments studied here, they will likely cause life-altering diseases in hospital specially for immunocompromised patients. Clearly the existence and survival in healthcare settings of these potentially pathogenic bacteria is a highly important research area which deserves continued investigation.

SUGGESTIONS FOR FURTHER STUDIES

The following important areas of research which have been examined in this Thesis need to be further studied: a) the survival of potentially pathogenic bacteria on tiles, b) the ways in which potentially pathogenic bacteria can survive on different types of piping used in plumbing in hospitals and c) the part played by hand dryers in distributing bacteria and other microbes inside the environment of hospitals and other healthcare facilities.

REFERENCES

- AHMED, I., YOKOTA, A., YAMAZOE, A. AND FUJIWARA, T. (2007) 'Proposal of *Lysinibacillus boronitolerans* gen. Nov. Sp. Nov., and transfer of *Bacillus fusiformis* to *Lysinibacillus fusiformis* comb. Nov. And *Bacillus sphaericus* to *Lysinibacillus sphaericus* comb. Nov. *International Journal of Systematic and Evolutionary Microbiology*, 57: 1117–1125.
- AL-GHAMDI, A. K., ABDELMALEK, S. M. ABDELMALEK, A., ASHSHI, A. M., FAIDAH, H. H., SHUKRI A. AND JIMAN-FATANI, A. A. (2011). Bacterial contamination of computer keyboards and mice, elevator buttons and shopping carts. *African Journal of Microbiology Research*. 5: 3998-4003
- ALEXANDER, M. 1977. *Introduction to Soil Microbiology*, New York
- ALTOMARE, C., NORVELL, W. A., RKMAN, T. B., AND HARMAN, G. E. (1999). Solubilization of phosphates and micronutrients by the plant-growth-promoting and biocontrol fungus *Trichoderma harzianum* Rifai 1295-22. *Applied and Environmental Microbiology* 65: 2926–2933.
- AMANN, R. I., LUDWIG, W. AND SCHLEIFER, K. H. (1995). Phylogenetic identification and *in situ* detection of individual microbial cells without cultivation. *Microbiology and Molecular Biology Reviews* 59: 143-169.
- ANASTASIADES, P., PRATT, TL., ROUSSEAU, L.H., STEINBERG, W.H., JOUBERT, G. (2009). *Staphylococcus aureus* on computer mice and keyboards in intensive care units of the universitas academic hospital, Bloemfontein and ICU staff's knowledge of its hazards and cleaning practices. *The Southern African journal of Epidemiology & Infection* 24: 22-26.

- BADGER, J. L., STINS, M. F., & SIK KIM, K. (1999).** *Citrobacter freundii* Invades and Replicates in Human Brain Microvascular Endothelial Cells. *Infection and Immunity*, 67:4208–4215.
- BADIEE, P., & HASHEMIZADEH, Z. (2014).** Opportunistic invasive fungal infections: diagnosis & clinical management. *The Indian Journal of Medical Research*, 139: 195–204.
- BALAPPANAVAR, A.Y., NAGESH, L., ANKOLA, A.V., TANGADE, P.S., KAKODKAR, P., AND VARUN, S. (2009).** Antimicrobial efficacy of various disinfecting solutions in reducing the contamination of the toothbrush - a comparative study. *Oral Health and Preventive Dentistry* 7: 137-145.
- BARBOZA-SILVA, E., CASTRO, A. C. D. AND MARQUIS, R. E. (2005).** Mechanisms of inhibition by fluoride of urease activities of cell suspensions and biofilms of *Staphylococcus epidermidis*, *Streptococcus salivarius*, *Actinomyces naeslundii* and of dental plaque. *Oral Microbiology and Immunology*, 20: 323–332.
- BARD, J. D., DEVILLE, J. G., SUMMANEN, P. H., and LEWINSKI, M. A. (2010).** *Roseomonas mucosa* Isolated from Bloodstream of Pediatric Patient . *Journal of Clinical Microbiology*, 48: 3027–3029.
- BAUER, T., OFNER, E., JUST, H. AND DASCHNER, F. (1990).** An epidemiological study assessing the relative importance of airborne and direct contact transmission of microorganisms in a medical intensive care unit. *Hospital Infection* 15: 301-309.
- BEHERA, B., SINGH, R. I., XESS, I., MATHUR, P., HASAN, F. AND MISRA, M. C. (2010)** *Candida rugosa*: a possible emerging cause of candidaemia in trauma patients. *Infection* 38: 387.

- BERGOGNE-BÉREZIN, E. AND TOWNER, K. J. (1996).** *Acinetobacter spp.* As nosocomial pathogens: Microbiological, clinical, and epidemiological features, *Clinical Microbiology Reviews.*, 9: 148–165.
- BECKER, K., RUTSCH, F., UEKÖTTER, A., KIPP, F., KÖNIG, J., MARQUARDT, T., PETERS, G., AND VON EIFF, C. (2008).** *Kocuria rhizophila* adds to the emerging spectrum of micrococcal species involved in human infections. *Journal of Clinical Microbiology*, 46: 3537–3539.
- BERDITSCH, M., AFONIN, S., AND ULRICH, A. S. (2007).** The Ability of *Aneurinibacillus migulanus* (*Bacillus brevis*) to produce the Antibiotic Gramicidin S. Is Correlated with Phenotype Variation? *Applied and Environmental Microbiology*, 73: 6620–6628.
- BEST E. L., PARNELL P., AND WILCOX M. H. (2014).** Microbial comparisons of hand-drying methods; the potential for contamination of the environment, user, and bystander. *Hospital Infection* 88:199–206.
- BEST, E. L., W. N. FAWLEY, P. PARNELL, AND M. H. WILCOX. (2010).** The potential for airborne dispersal of *Clostridium difficile* from symptomatic patients. *Clinical Infectious Disease*. 50:1450-1457.
- BLACKMORE, M. A. (1989).** A comparison of hand drying methods. *Catering & Health* 1:189-198.
- BOURAFI, N., LOUCIF, L., BOUTEFNOUCHET, N. AND ROLAIN, J. (2015).** ‘*Enterococcus hirae*, an unusual pathogen in humans causing urinary tract infection in a patient with benign prostatic hyperplasia: First case report in Algeria’, *New Microbes New Infections*, 8: 7–9.

- BOYCE, J. M., GAIL, P.-B., CLAIRE, C., AND THOMAS, K. (1997).** Environmental contamination due to methicillin-resistant *staphylococcus aureus* possible infection control implications. *Infection Control and Hospital Epidemiology* 18:622-627.
- BOYLE, M.A., O'DONNELL M.J., MILLER, A., RUSSELL, R.J. AND COLEMAN, D.C. (2012).** Control of bacterial contamination of washbasin taps and output water using Ecasol: a one-year study. *Journal of Hospital Infection*, 80: 288-292.
- BRADY, C., CLEENWERCK, I., VENTER, S., ENGELBEEN, K., DE VOS, P. AND COUTINHO, T. (2010).** Emended description of the genus *Pantoea*, description of four species from human clinical samples, *Pantoea septica* sp. nov., *Pantoea eucrina* sp. nov., *Pantoea brenneri* sp. nov. and *Pantoea conspicua* sp. nov., and transfer of *Pectobacterium cypripedii* (Hori 1911) Brenner *et al.* 1973 emend. Hauben *et al.* 1998 to the genus as *Pantoea cypripedii* comb. nov. *International Journal of Systematic and Evolutionary Microbiology* 60:2430-2440
- BRENNER, D.J., MCWHORTER, A., KAI, A., STEIGERWALT, A., AND FARMER, J. J. (1986).** *Enterobacter asburiae* sp. nov., a new species found in clinical specimens, and reassignment of *Erwinia dissolvens* and *Erwinia nimipressuralis* to the genus *Enterobacter* as *Enterobacter dissolvens* comb. nov. and *Enterobacter nimipressuralis* comb. nov. *Journal of Clinical Microbiology*. 23 :1114-20
- BROOKE, J. S. (2012).** *Stenotrophomonas maltophilia*: An Emerging Global Opportunistic Pathogen. *Clinical Microbiology Reviews*, 25: 2–41.
- BUNETEL, L., TRICOT-DOLEUX, S., AGNANI, G., AND BONNAURE-MALLET, M. (2000).** *In vitro* evaluation of the retention of three species of pathogenic microorganisms by three different types of toothbrush. *Oral Microbiology and Immunology* 15: 313–316.

- BURES, S., JOEL, T. F., CATHERINE, F. T., JOSEPH, M., AND BENJAMIN, W. (2000).** Computer keyboards and faucet handles as reservoirs of nosocomial pathogens in the intensive care unit. *Infection Control* 28:465–471.
- BURKE, A. C. (2010).** *Infectious Diseases in Critical Care Medicine*. Third Edition. Informa Healthcare, New York, USA
- CATALANOTTO, F. A., SHKLAIR, I. L., AND KEENE, H. J. (1975).** Prevalence and localization of *Streptococcus mutans* in infants and children. *Journal of the American Dental Association*, 91: 606-609.
- CESAR, P. G.-C., MARIA, J. N. A., AND OMAR, E. A. H. (2012).** Fungal and bacterial contamination on indoor surfaces of a hospital in Mexico. *Jundishapur Journal Of Microbiology* 5: 460-464.
- CERVERA, C., M. ALMELA, J. A. MARTINEZ-MARTINEZ, A. MORENO, AND MIRO, J. M. (2009).** Risk factors and management of Gram-positive bacteraemia. *International Journal of Antimicrobial Agents* 34:26-30.
- CHEMALY, R. F., SIMMONS, S., DALE, C., GHANTOJI, S. S., RODRIGUEZ, M., GUBB, J., STACHOWIAK, J. AND STIBICH, M. (2014).** The role of the healthcare environment in the spread of multidrug-resistant organisms: update on current best practices for containment. *Therapeutic Advances in Infectious Disease*, 2: 79–90.
- DANCER, S. J. (2009).** The role of environmental cleaning in the control of hospital acquired infection. *Hospital Infection* 73: 378-385.
- DANCER, S. J. (2014).** Controlling Hospital-Acquired Infection: Focus on the Role of the Environment and New Technologies for Decontamination. *Clinical Microbiology Reviews*, 27:665–690.

- DAVIES, R. M. (2008).** Toothpaste in the control of plaque/gingivitis and periodontitis. *Periodontology* 48: 23-30.
- DAYOUB, M. B., RUSILKO, D., AND GROSS, A. (1977).** Microbial contamination of toothbrushes. *Journal of Dental Research*. 56:706.
- DE ANGELIS, M. AND GOBBETTI, M. (2004).** Environmental stress responses in *Lactobacillus*: a review, *Proteomics*, 4: 106–22.
- DE ROSSI, A., FERREIRA, D., DA SILVA, R., DE QUEIROZ, A., DA SILVA, L., NELSON-FILHO, P. (2014).** Antimicrobial activity of toothpastes containing natural extracts, Chlorhexidine or Triclosan. *Brazilian Dental Journal* 25: 186-190.
- DIBA, K., RHAIMIRAD, M., MAKHDOOMI, K. AND KHORSHIDVAND, Z. (2012).** Identification of *Candida* species isolated from hospital acquired infections patients and hospital indoor environments. *African Journal of Microbiology Research* 6:4164-4168.
- DOWNES, J., HOOPER, S. J., WILSON, M. J., WADE, W. G. (2008).** *Prevotella histicola* sp. nov., isolated from the human oral cavity. *International Journal of Systematic and Evolutionary Microbiology*, 58: 1788-1791.
- EDWAY, K. AND FAWDAR, S. (2008).** *A comparative study of three different hand drying methods: paper towel, warm air dryer, jet air dryer.* European Tissue Symposium. University of Westminster, UK.
- ESPÍRITO SANTO, C., ELOWSKY, C. G., D.W., CHANG, C. J., GRASS, G., QUARANTA, D., AND TRAVIS, K. (2011).** Mechanisms of contact-mediated killing of yeast cells on dry metallic copper surfaces. *Applied and Environmental Microbiology* 77: 416–426.

ESPIRITO SANTO, C., NADINE, T., DIETRICH, H., NIES, AND GREGOR, G. (2008).

Contribution of copper ion resistance to survival of *Escherichia coli* on metallic copper surfaces. *Applied and Environmental Microbiology* 74:977-986.

FALVEY D. G., AND STREIFEL A. J. (2007). Ten-year air sample analysis of *Aspergillus*

prevalence in a university hospital. *ScienceDirect, Hospital Infection* 67: 35–41.

FAN, Y., JIN, Z., TONG, J., LI, W., PASCIAK, M., GAMIAN, A., LIU, Z. AND HUANG, Y.

(2002). *Rothia amarae* sp. Nov., from sludge of a foul water sewer, *International Journal of Systematic and Evolutionary Microbiology*. 52: 2257–60.

FILHO, P.N., FARIA, G., ASSED, S., AND ITO, I.Y. (2000). Microbial contamination of

toothbrushes and their decontamination. *American Academy of Pediatric Dentistry* 22: 381- 384.

FRAZELLE, M. R. AND MUNRO, C. L. (2012) “Toothbrush Contamination: A Review of the

Literature,” *Nursing Research and Practice*, 2012:6.

FRITZ, S., CASSIR, N., NOUDEL, R., DE LA ROSA, S., ROCHE, P.-H., & DRANCOURT, M.

(2014). Postsurgical *Pantoea calida* meningitis: a case report. *Journal of Medical Case Reports*, 8, 195.

FUJIWARA, T, SASADA, E, MIMA, N, AND OOSHIMA, T. (1991). Caries prevalence and

salivary mutans *streptococci* in 0-2 years old children of Japan. *Community Dentistry and Oral Epidemiology* 19:151–154.

- GASTMEIERA, P., SCHWABB, F. ARWOLFFB, S. B., RÜDENB, H. AND GRUNDMANNC, H. (2006).** Correlation between the genetic diversity of nosocomial pathogens and their survival time in intensive care units. *Journal of Hospital Infection* 62: 181–186.
- GRIFFITHS, M. (2005).** Understanding Pathogen Behavior Virulence, stress response and resistance. Woodhead Publishing Limited, Cambridge: England.
- GLASS RT, LARE MM., (1986).** Toothbrush contamination: a potential health risk? *Quintessence International, Europe PMC* 17: 39-42.
- GLASS RT., (1992).** The infected toothbrush, the infected denture, and transmission of disease. *US National Library of Medicine National Institutes of Health*, 3: 592-598.
- GOLDMAN, J. M., MICHAEL, W. N., DEININGER, J., MELO, V. (2000).** The molecular biology of chronic myeloid leukemia. *American Society of Hematology, Blood* 96: 3343-3356.
- GRASS, G., RENSING, C., SOLIOZ, M. (2011).** Metallic copper as an antimicrobial surface. *Applied and Environmental Microbiology* 77: 1541–1547.
- GRASS G., ANDRÉ M., SUSANNE H., LADJI T., JÖRG B., DIETRICH H. N. (2010).** Survival of bacteria on metallic copper surfaces in a hospital trial. *Applied Microbial and Cell Physiology, Applied Microbiology and Biotechnology* 87: 1875-1879.
- GUPTA, D. AND CHAUHAN P. (2016).** Fungicidal activity of silver particles against *Alternaria brassicicola*. *Nanoscale Research Letters* 10: 1-15.

- HAILS, J., KWAKU, F., WILSON, A.P. AND BELLINGAN G, SINGER M. (2003).** Large variation in MRSA policies, procedures and prevalence in English intensive care units: a questionnaire analysis. *Intensive Care Medicine*. 29:481–483.
- HARRISON, J., (2003).** The nitrogen cycle of microbes and men. Module Library, *Vision learning* 2: 1-6.
- HARTMANN, B., BENSON, M., JUNGER, A. QUINZIO L, RÖHRIG R, FENGLER B, FÄRBER UW, WILLE B, AND HEMPELMANN G. (2004).** Computer keyboard and mouse as a reservoir of pathogens in an intensive care unit. *Journal of Clinical Monitoring and Computing*.18:7–12.
- HESS, B., BURCHETT, A., AND HUNTINGTON, M. K. (2008).** *Leclercia adecarboxylata* in an immunocompetent patient. *Journal of Medical Microbiology* 57: 896-898.
- HIDRON, A., EDWARDS, J., PATEL, J., HORAN, T., SIEVERT, D., POLLOCK, D., & FRIDKIN, S. (2008).** Antimicrobial-Resistant Pathogens Associated with Healthcare-Associated Infections: Annual Summary of Data Reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006–2007. *Infection Control & Hospital Epidemiology*, 29: 996-1011.
- HILL, G., MITKOWSKI, N., ALDRICH-WOLFE, L., EMELE, L., JURKONIE, D., FICKE, A., MALDONADO-RAMIREZ, S., LYNCH, S. AND NELSON, E. (2000).** Methods for assessing the composition and diversity of soil microbial communities. *Applied Soil Ecology* 15: 25–36.

- HÖGENAUER, C., LANGNER, C., BEUBLER, E., LIPPE, I.T., SCHICHO, R., GORKIEWICZ, G., KRAUSE, R., GERSTGRASSER, N., KREJS, G.J. AND HINTERLEITNER, T.A. (2006)** '*Klebsiella oxytocaas* a causative organism of antibiotic-associated Hemorrhagic Colitis', *New England Journal of Medicine*, 355: 2418–2426.
- HOLAH, J. (2011).** *Interview on Hygienic Materials*. Interviewed by Mikael Andersson. Chipping Campden, England.
- HOTA, B., (2004).** Contamination, disinfection and cross contamination-Are hospital surfaces reservoirs for nosocomial infection? *Healthcare Epidemiology, Clinical Infectious Diseases* 39:1182-1189.
- HUANG C, WENJA MA AND STACK, S., (2012).** The hygienic efficacy of different hand drying methods: A review of the evidence. *Mayo Clinic Proceedings* 87: 791–798.
- HUGONNET, S., PERNEGER, T.V., AND PITTET, D. (2002).** Alcohol-Based Handrub Improves Compliance with Hand Hygiene in Intensive Care Units. *Archives of Internal Medicine*. 162:1037-1043.
- INCANI, R.N., HERNÁNDEZ, M, CORTEZ, J., GONZÁLEZ, M., DOREL, S. Y. (2010)** *Staphylococcus warneri* meningitis in a patient with Strongyloides stercoralis hyperinfection and lymphoma: first report of a case. *Revista do Instituto de Medicina Tropical de São Paulo* 52: 169-170.
- ISAACS D., DALEY A., DALTON, D. (1998).** Swabbing computers in search of nosocomial bacteria. *Pediatric Infectious Disease Journal* 17: 533.
- JONES C G. (1997).** Chlorhexidine: is it still the gold standard? *Periodontology* 2000 15: 55–62.

- JUNG, W. K., KOO, H. C., KIM, K. W., SHIN, S., KIM, S. H., & PARK, Y. H. (2008).** Antibacterial Activity and Mechanism of Action of the Silver Ion in *Staphylococcus aureus* and *Escherichia coli*. *Applied and Environmental Microbiology*, 74: 2171–2178.
- KALWASIŃSKA, A., BURKOWSKA, A. AND WILK, I. (2012).** Microbial air contamination in indoor environment of a university library. *Annals of Agricultural and Environmental Medicine* 19: 25–9.
- KARBOWSKA-BERENT, J., GÓRNY, R. L., STRZELCZYK, A. B., WLAZŁO, A. (2011)** Airborne and dust borne microorganisms in selected Polish libraries and archives, *Building and Environment*, 46: 1872-1879.
- KATZ, J.D. (2004).** Hand washing and hand disinfection. *Anesthesiology Clinics of North America* 22: 457-471.
- KAYABAS, U., BAYRAKTAR, M., OTLU, B., UGRAS, M., ERSOY, Y., BAYINDIR, Y. AND DUMAZ, R. (2008).** An outbreak of *Pseudomonas aeruginosa* because of inadequate disinfection procedures in a urology unit: A pulsedfield gel electrophoresis-based epidemiologic study. *Infection Control* 36: 33-38.
- KHAN, H., AHMAD, A., MEHBOOB, R. (2015).** Nosocomial infections and their control strategies. *Asian Pacific Journal of Tropical Biomedicine*, 5: 509-514.
- KHAN, S., SISTLA, S., DHODAPKAR, R., AND PARIJA, S. C. (2012).** Fatal *Delftia acidovorans* infection in an immunocompetent patient with empyema. *Asian Pacific Journal of Tropical Biomedicine*, 2: 923-924.
- KILLHAM, K. S. (1994).** Soil Ecology, Cambridge, Cambridge University Press.

- KIM K. Y., JORDAN D., MCDONALD G. A. (1997).** Effect of phosphate-solubilizing bacteria and vesicular-arbuscular mycorrhizae on tomato growth and soil microbial activity. *Biology and Fertility of Soils* 26: 79-87.
- KIM K. Y., JORDAN D., MCDONALD G. A. (1998).** *Enterobacter agglomerans*, phosphate solubilizing bacteria, and microbial activity in soil: Effect of carbon sources. *Soil Biology and Biochemistry* 30: 995–1003.
- KIMOULI, MARIA & VRIONI, GEORGIA & PAPADOPOULOU, MAGDALINI & KOUMAKI, VASILIKI & PETROPOULOU, DIMITRA & GOUNARIS, ANTONIOS & FRIEDRICH, ALEXANDER & TSAKRIS, ATHANASSIOS. (2011).** Two cases of severe sepsis caused by *Bacillus pumilus* in neonatal infants. *Journal of medical microbiology*. 61: 596-599.
- KLEVENS, R. M., EDWARDS, J. R., RICHARDS, C. L., HORAN, T. C., GAYNES, R. P., POLLOCK, D. A., & CARDO, D. M. (2007).** Estimating Health Care-Associated Infections and Deaths in U.S. Hospitals, 2002. *Public Health Reports*, 122: 160–166.
- KNIGHTS B., EVANS C., BARRASS S., MCHARDY B. (1993).** *Hand Drying: An Assessment of Efficiency and Hygiene of Different Methods: A Survey Carried Out by the Applied Ecology Research Group for the Association of Makers of Soft Tissue Papers.* University of Westminster; London, UK.
- KRAMER, A., SCHWEBKE, I., KAMPF, G. (2006).** How long do nosocomial pathogens persist on inanimate surfaces? A systematic review. *BioMed Central, Infectious Diseases*, DOI: 10.1186/1471-2334-6-130.

- KUROKI, R., KAWAKAMI, K., QIN, L., KAJI, C., WATANABE, K., KIMURA, Y., ISHIGURO, C., TANIMURA, S., TSUCHIYA, Y., HAMAGUCHI, I., SAKAKURA, M., SAKABE, S., TSUJI, K. AND INOUE, M. (2009).** ‘Nosocomial bacteremia caused by biofilm-forming *Bacillus cereus* and *Bacillus thuringiensis*’. *Internal medicine* 48:791–6.
- LAHIRI, D., BYA, S., JR, J., HODES, M. AND CRISP, M. (1992).** A non-organic and non-enzymatic extraction method gives higher yields of genomic DNA from whole-blood samples than do nine other methods tested. *Biochemical and Biophysical Methods* 25: 193-205.
- LANGLEY, J. (2002).** From soap and water, to waterless agents: Update on hand hygiene in health care settings. *The Canadian Journal of Infectious Diseases*, 13: 285–286.
- LAI K. K. (2001).** A cluster of invasive aspergillosis in a bone marrow transplant unit related to construction and the utility of air sampling. *Infection Control* 29: 333–337.
- LIU, D. (2011).** Molecular Detection of Human Bacterial Pathogens. CRC Press YU, S.J., YIN.
- MADIGAN, M. T., MARTINKO, J. M., DUNLAP, P. V. AND CLARK, D. P. (2012).** Cell structure and function in bacteria and archaea. *Biology of Microorganisms*. pp 1-31.
- MAGES, I.S., FRODL, R., BERNARD, K.A. AND FUNKE, G. (2008).** Identities of *Arthrobacter* spp. And *Arthrobacter*-Like bacteria encountered in human clinical specimens. *Journal of Clinical Microbiology* 46: 2980–2986.
- MAIER, R. M., PEPPER, I. L. AND GERBA, C. P. (2009).** *Environmental Microbiology*. London, Academic Press.
- MARSH, P.D. (2010).** Controlling the oral biofilm with antimicrobials. *Journal of Dentistry* 38: S11–S15.

- MARTIN, M. A., PFALLER, M. A. AND WENZEL, R. P. (1989).** Coagulase-negative *Staphylococcal* bacteremia. Mortality and hospital stay. *Annals of Internal Medicine* 110: 9-16.
- MARQUIS, R.E. (1995).** Antimicrobial actions of fluoride for oral bacteria. *Canadian Journal of Microbiology* 41: 955-964.
- MCBRYDE, E.S., BRADLEY, L.C., WHITBY, M., MCELWAIN, D.L.S. (2004).** An investigation of contact transmission of methicillin-resistant, *Journal of Hospital Infection*, 58:104-108.
- MCCARTHY, C., SNYDER, M.L, PARKER, R.B. (1965).** The indigenous flora of man. I -The newborn to the 1 year old infant. *Archives of Oral Biology* 10: 61-70.
- MEDINA, M., RODRÍGUEZ, M., ARENAS, M. AND GALLEG0, G. (1997).** Nosocomial infections in surgical patients: comparison of two measures of intrinsic patient risk. *Infection Control and Hospital Epidemiology* 18: 19-23.
- MEHTA, G. (1990).** *Aspergillus* endocarditis after open heart surgery: an epidemiological investigation. *Hospital Infection* 15: 245-253.
- MEHTAR, S., WIID, I., AND TODOROV, S.D. (2008).** The antimicrobial activity of copper and copper alloys against nosocomial pathogens and *Mycobacterium tuberculosis* isolated from healthcare facilities in the Western Cape: an in vitro study. *Journal of Hospital Infection*. 68: 45–51.
- METRI, B. C., JYOTHI, P., & PEERAPUR, B. V. (2013).** Antibiotic resistance in *Citrobacter spp.* isolated from urinary tract infection. *Urology Annals*, 5: 312–313.
- MISHRA, K. N., AAGGARWAL, A., ABDELHADI, E. AND SRIVASTAVA, P. C. (2010).** An efficient horizontal and vertical method for online DNA sequence compression. *International Journal of Computer Applications* 3: 39-46
- MULLIS, K. (1990).** PCR-the polymerase chain reaction. *Data Science* 11: 249.

NASCIMENTO, C., SCARABEL, T.T., MIANI, P.K., WATANABE, E., AND PEDRAZZI, V.

(2012). In vitro evaluation of the microbial contamination on new toothbrushes: a preliminary study. *Microscopy Research and Technique* 75: 42–45.

NASCIMENTO, C., TRINCA, N., PITA, M., AND PEDRAZZI, V. (2015). Genomic

identification and quantification of microbial species adhering to toothbrush bristles after disinfection: A cross-over study, *In Archives of Oral Biology*, 60: 1039-1047

NEELY A. N., MALEY M. P. (2000). Survival of *Enterococci* and *Staphylococci* on hospital

fabrics and plastic. *Journal of Clinical Microbiology* 38:724-726.

NEELY A. N., MATTHEW P. M., GLENN D. W. (1999). Computer keyboards as reservoirs for

Acinetobacter baumannii in a burn hospital. *Oxford Journals, Medicine and Health, Clinical Infectious Diseases* 29: 1358-1359.

NOYCE, J.O., MICHELS, H., KEEVIL, C.W. (2006). Potential use of copper surfaces to reduce

survival of epidemic meticillin-resistant *Staphylococcus aureus* in the healthcare environment. *Hospital Infection* 63: 289–297.

ORTEGA, M., MARCO, F., SORIANO, A. (2010). *Candida* spp. bloodstream infection: influence

of antifungal treatment on outcome. *Journal of Antimicrobial Chemotherapy*. 65: 562-568.

OTOIKHIAN, C. S. O., AND OKOROR, L. O. (2012). Resistance of Oral Bacterial Species to

Varied Toothpastes Effects. *International Journal of Engineering Research and Science & Technology* 1:1-10

OTTER, J. A., AND FRENCH, G. L. (2009). Survival of Nosocomial Bacteria and Spores on

Surfaces and Inactivation by Hydrogen Peroxide Vapor. *Journal of Clinical Microbiology*, 47:205–207.

- PANNUTI, C.M., MATTOS, J.P., RANOYA, P.N., JESUS, A.M., LOTUFO, R.F., ROMITO, G.A. (2003).** Clinical effect of a herbal dentifrice on the control of plaque and gingivitis: a double-blind study. *Brasileira de Pesquisa Odontológica* 17: 314-318.
- PATTABI, R. M., AND PATTABI, M. (2013).** Antibacterial Applications of Silver Nanoparticles. *Materials Science Forum*, 754:131-142.
- PAUL, E.A AND CLARK, F.E. (1989).** *Soil Microbiology and Biochemistry*. Academic Press, London.
- PERLROTH, J., CHOI, B., AND SPELLBERG, B. (2007).** Nosocomial fungal infections: epidemiology, diagnosis, and treatment. *Medical Mycology* 45:321-46.
- PFALLER, M. A., DIEKEMA, D. J., COLOMBO, A. L., KIBBLER, C., NG, K. P., GIBBS, D. L., NEWELL, V. A AND THE GLOBAL ANTIFUNGAL SURVEILLANCE GROUP. (2006).** *Candida rugosa*, an Emerging Fungal Pathogen with Resistance to Azoles: Geographic and Temporal Trends from the ARTEMIS DISK Antifungal Surveillance Program. *Journal of Clinical Microbiology*, 44: 3578–3582.
- PRASANTH, M. (2011).** Antimicrobial efficacy of different toothpastes and mouth rinses: an in vitro study. *Dental Research Journal* 8: 85-94.
- PRIEST, F. G., GOODFELLOW, M., SHUTE, L. A. AND BERKELEY, R. C. W. (1987).** *Bacillus amyloliquefaciens* sp. nov. norn. rev. *International Journal of Systematic Bacteriology* 37:69-71
- POPOVICH, K. J., R. A. WEINSTEIN, AND B. HOTA. (2008).** Are community-associated methicillin resistant *Staphylococcus aureus* (MRSA) strains replacing traditional nosocomial MRSA strains? *Clinical Infectious Diseases* 46:787-794.

- POYART, C., LAMBERT, T., MORAND, P., ABASSADE, P., QUESNE, G., BAUDOUY, Y. AND TRIEU-CUOT, P. (2002).** ‘Native valve Endocarditis due to *Enterococcus hirae*’, *Journal of Clinical Microbiology*, 40: 2689–2690.
- QUARANTA, D., KRANS, T., ESPIRITO SANTO, C., ELOWSKY, C.G., DOMAILLE, D.W., CHANG, C.J. AND GRASS, G. (2011).** Mechanisms of yeast contact-killing on dry metallic copper surfaces. *Applied and Environmental Microbiology* 77:416–426.
- RAJESH, B., USHA, D. C., BINDU, R. A. D. AND REDDY, B. I. (2013).** Screening and microbial characterization of Lipase producing organic solvent tolerant *Lysinibacillus Fusiformis* C5 (MTCC 11801). *International Journal of Scientific & Engineering Research*, 4:427-432.
- RAMANAN, P., BARRETO, J.N. AND OSMON, D.R. (2014).** *Rothia Bacteraemia: A 10-Year experience at Mayo clinic, Rochester, Minnesota*, *Journal of Clinical Microbiology*, 52: 3184–3189.
- RAMIREZ, M.S., VAZQUEZ, M., TANAKA, N., TURCO, M., ALMUZARA, M., LOPEZ, E.L., PASTERAN, F., RAPOPORT, M., PROCOPIO, A. AND VAY, C.A. (2010).** First case of human infection due to *Pseudomonas fulva*, an environmental bacterium isolated from Cerebrospinal fluid, *Journal of Clinical Microbiology*, 48: 660–664.
- RANDALL, J., SEOW, W. AND WALSH, L. (2015).** Antibacterial activity of fluoride compounds and herbal toothpastes on *Streptococcus mutans*: an in vitro study. *Australian Dental Journal*, 60: 368–374.
- REGALADO, N.G., MARTIN, G. AND ANTONY, S. J. (2009).** *Acinetobacter lwoffii*: Bacteraemia associated with acute gastroenteritis’. *Travel Medicine and Infectious Disease*, 7: 316–317.

- RIBERA, G., ROMANO, F., GIULIANO, M. (1994).** A study of a hospital cluster of systemic candidiosis using DNA typing methods. *Epidemiology and Infection* 112: 93-398.
- ROUX, D., AUBIER, B., COCHARD, H., QUENTIN, R., VAN DER MEE-MARQUET, N. (2013).** Contaminated sinks in intensive care units: an underestimated source of extended-spectrum beta-lactamase-producing Enterobacteriaceae in the patient environment. *Journal of Hospital Infection*; 85:106-111.
- RUTALA, W. A., WHITE, M. S., GERGEN, M. F., AND WEBER, D. J. (2006).** Bacterial contamination of keyboards: efficacy and functional impact of disinfectants. *Infection Control & Hospital Epidemiology* 27:372-376.
- SAMMONS, R., KAUR, D. AND NEAL, P. (2004).** Bacterial survival and biofilm formation on conventional and antibacterial toothbrushes. *Biofilms* 1: 123-130.
- SANDERS, M. E., MORELLI, L. AND TOMPKINS, T. A. (2003).** *Sporeformers* as Human Probiotics: *Bacillus*, *Sporolactobacillus*, and *Brevibacillus*. *Institute of Food Technologists* 2:101-110.
- SAVINI, V., BONFINI, T., MARROLLO, R., ARGENTIERI, A., RICCIONI, S., ASTOLFI, D., FAZIL, P., D'ANTONIO, D. AND GHERARDI, G. (2013).** 'Enterococcus hirae: A zoonotic microorganism in human umbilical cord blood'. *World Journal of Microbiology and Biotechnology*, 30: 1423–1426.
- SCHOLLE, M. D., WHITE, C. A., KUNNIMALAIYAAN, M., & VARY, P. S. (2003).** Sequencing and Characterization of pBM400 from *Bacillus megaterium* QM B1551. *Applied and Environmental Microbiology*, 69: 6888–6898.
- SCOTT, E., AND BLOOMFIELD, S.F. (2008).** The survival and transfer of microbial contamination via cloths, hands and utensils. *Journal of Applied Microbiology* 68: 271-278.

- SEHULSTER, L. AND CHINN, R. (2003).** Guidelines for environmental infection control in health-care facilities. *Morbidity and Mortality Weekly Report (MMWR)* 52: 1-42.
- SHIDA, O.; TAKAGI, H.; KADOWAKI, K.; UDAKA, S.; NAKAMURA, L.; KOMAGATA, K. (1995).** "Proposal of *Bacillus reuszeri* sp. nov., *Bacillus formosus* sp. nov., nom. rev., and *Bacillus borstelensis* sp. nov., nom. rev.". *International Journal of Systematic Bacteriology* 43:93–100.
- SHIVAJI, S., CHATURVEDI, P., SURESH, K., REDDY, G.S.N., DUTT, C.B.S., WAINWRIGHT, M., NARLIKAR, J.V. AND BHARGAVA, P.M. (2006)** *Bacillus aerius* sp. Nov., *Bacillus aerophilus* sp. Nov., *Bacillus stratosphericus* sp. Nov. And *Bacillus altitudinis* sp. Nov., isolated from cryogenic tubes used for collecting air samples from high altitudes, *International Journal of Systematic and Evolutionary Microbiology*, 56: 1465–1473.
- SIMS, J. R. AND JACKSON, G. D. (1971)** *Soil Sci. Soc. Am. Proc.*, 35, 603-606.
- SIMS, J.R. AND JACKSON, G.D. (1971).** Rapid analysis of soil nitrate with chromic acid. *Soil Science Society of America Proceedings* 35, 603-606.
- SMITH, D. (2009).** *Hand hygiene: guidelines for best practice*. Campden BRI Guideline No. 62. Chipping Campden: Campden BRI.
- SNYDER, O. P. (1998).** Hand Washing for Retail Food Operations- A Review. *Dairy, Food and Environment Sanitation*, 18:149-162.
- SOCRANSKY, S. S., AND MANGANIELLO, S. D. (1971).** The oral microbiota of man from birth to senility. *Periodontology* 42: 485-496.

- SPAULDING, A. R., SALGADO-PABÓN, W., KOHLER, P. L., HORSWILL, A. R., LEUNG, D. Y. M., & SCHLIEVERT, P. M. (2013).** Staphylococcal and Streptococcal Superantigen Exotoxins. *Clinical Microbiology Reviews*, 26: 422–447.
- SULLIVAN, D., HAYNES, K., BILLE, J., BOERLIN, P., RODERO, L., LLOYD, S., HENMAN, M. AND COLEMAN, D. (1997).** Widespread geographic distribution of oral *Candida dubliniensis* strains in human immunodeficiency virus-infected individuals. *Clinical Microbiology* 35: 960-964.
- TAKAGI, H., O. SHIDA, K. KADOWAKI, K. KOMAGATA, AND S. UDAKA. (1993).** Characterization of *Bacillus brevis* with descriptions of *Bacillus migulanus* sp. nov., *Bacillus choshinensis* sp. nov., *Bacillus parabrevis* sp. nov., and *Bacillus galactophilus* sp. nov. *International Journal of Systematic Bacteriology*. 43:221-231.
- TALON, D. (1999).** The role of the hospital environment in the epidemiology of multi-resistant bacteria. *Journal of Hospital infection* 43: 13–17.
- TAM, V., & NAYAK, S. (2012).** Isolation of *Leclercia adecarboxylata* from a wound infection after exposure to hurricane-related floodwater. *BMJ Case Reports*, 2012:1-3.
- TRAORÉ O., SPRINGTHORPE V.S., SATTAR S.A. (2002).** A quantitative study of the survival of two species of *Candida* on porous and non-porous environmental surfaces and hands. *Applied Microbiology* 92: 549-555
- THOMAS A. C., SALLY A. S., JULIETTE M., TIMOTHY L., BETH A. A-S., MARY E. B., RISA M. W., MARY C., RICHARD H. F., SCOTT K. F., AND RANA A. H. (2004).** Epidemiologic and molecular characterization of an outbreak of *Candida parapsilosis* bloodstream infections in a community hospital. *Clinical Microbiology* 42: 4468–4472.

- TRAORE, O., SPRINGTHORPE, V. AND SATTAR, S. (2002).** A quantitative study of the survival of two species of *Candida* on porous and non-porous environmental surfaces and hands. *Journal of Applied Microbiology* 92: 549-555.
- TWETMAN, S. (2004).** Antimicrobials in future caries control? A review with special reference to chlorhexidine treatment. *Caries Research* 38: 223-229.
- VALONES, M. A. A., GUIMARÃES, R. L., BRANDÃO, L. A. C., DE SOUZA, P. R. E., DE ALBUQUERQUE TAVARES CARVALHO, A., & CROVELA, S. (2009).** Principles and applications of polymerase chain reaction in medical diagnostic fields: a review. *Brazilian Journal of Microbiology*, 40: 1–11.
- VERHILLE S, BAIDA N, DABBOUSSI F, IZARD D, LECLERC H (1999).** Taxonomic study of bacteria isolated from natural mineral waters: proposal of *Pseudomonas jessenii* sp. nov. and *Pseudomonas mandelii* sp. nov. *Systematic and Applied Microbiology* 22: 45–58
- VERKAIK, M. J., BUSSCHER, H. J., JAGER, D., SLOMP, A. M., ABBAS, F., AND VAN DER MEI, H. C. (2011).** Efficacy of natural antimicrobials in toothpaste formulations against oral biofilms *in vitro*. *Journal of Dentistry* 39: 218–224.
- VERRAN, J., AND LEAHY-GILMARTIN, A. A. (1996).** Investigations into the microbial contamination of toothbrushes. *Europe PMC, Microbios* 85: 231-238.
- WARNES, S. L., GREEN, S. M., MICHELS, H. T. AND KEEVIL, C. W. (2010).** Biocidal efficacy of copper alloys against pathogenic enterococci involves degradation of genomic and plasmid DNAs. *Applied and Environmental microbiology* 76: 5390-5401.
- WEAVER, L. MICHELS, H.T. AND KEEVIL, C.W. (2010).** Potential for preventing spread of fungi in air-conditioning systems constructed using copper instead of aluminium. *Letters in Applied Microbiology*, 50: 8-23.

WEBER DJ, RUTALA WA, MILLER MB, HUSLAGE K, SICKBERT-BENNETT E. (2010).

Role of hospital surfaces in the transmission of emerging health care-associated pathogens: *Norovirus, Clostridium difficile, and Acinetobacter species*. *American Journal of Infection Control*. 38:25-33.

WENZLER, E., KAMBOJ, K. AND BALADA-LLASAT, J. (2015). ‘Severe sepsis secondary to

persistent and Bacteremia’. *International Journal of Infectious Diseases*, 35: 93–95.

WILKS, S. A., MICHELS, H., KEEVIL, C. W. (2005). The survival of *Escherichia coli* O157 on a

range of metal surfaces. *Intentional Journal of Food Microbiology* 105:445–454.

WISPLINGHOFF, H., BISCHOFF, T., TALLENT, S. M., SEIFERT, H., WENZEL, R. P.,

EDMOND, M. B. (2004). Nosocomial Bloodstream Infections in US Hospitals:

Analysis of 24,179 Cases from a Prospective Nationwide Surveillance Study.

Clinical Infectious Disease 39: 309-317.

WOLF, A., FRITZE, A., HAGEMANN, M., AND BERG, G. (2002). *Stenotrophomonas*

rhizophila sp. nov., a novel plant-associated bacterium with antifungal

properties. *International Journal of Systematic and Evolutionary Microbiology*, 52:

1937–1944.

XIONG, Z., ZHANG, J., ZHANG, D., ZHOU, Z., LIU, M., ZHU, W., ZHAO, L., XU, L. AND

L.I, W. (2013). *Rothia endophytica* sp. Nov., an actinobacterium isolated from

Dysophylla stellata (Lour.) Benth. *International Journal of Systematic and*

Evolutionary Microbiology, 63: 3964–3969.

YANG. D., AND ZHANG, Z. (2008). Biofilm-forming *Klebsiella pneumoniae* strains have greater

likelihood of producing extended-spectrum β -lactamases. *Journal of Hospital*

Infection. 68:369–371.

- YAP, D.Y.H., TSE, H., MOK, M.M.Y., CHAN, G.C.W., YIP, T., LUI, S.L., LO, W.K. AND CHAN, T.M. (2015).** *Arthrobacter sanguinis*: An uncommon cause of peritonitis in a peritoneal dialysis patient, *Nephrology*, 20: 868–869.
- YAZGI, H., UYANIK M. H., ERTEK, M., AKTAŞ, A. E., İGAN, H. AND AYYILDIZ, A. (2009).** Survival of certain nosocomial infectious agents on the surfaces of various covering materials. *Turkish Journal of Medical Science*; 39: 619-622.
- YU, S. J., YIN, Y. G, AND LIU, J. F. (2013).** Silver nanoparticles in the environment. *Environmental Science: Processes & Impacts*. 15:78-92.

APPENDIX 1

Publications Arising From This Thesis

Bacterial contamination of used manual toothbrushes and effects of toothpastes on isolated potential pathogens

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Toothbrushes play an essential role in oral hygiene and are commonly found in community and hospital settings. The toothbrushes may act as a reservoir for potential pathogens transferred from the oral cavity and from the bathroom environment. The purpose of this study is to determine the bacterial contamination of used toothbrushes and determine the antibacterial effect of toothpastes. Scanning electron microscopy (SEM) was used to visualize biofilms on toothbrush bristles. 50 used toothbrushes obtained from volunteers were analysed bacteriologically using standard microbiological techniques. Bacteria present on all toothbrushes heads were cultured to determine the presence of bacteria and scanned by SEM. The antibacterial effect of toothpastes was determined using seven types of commercial toothpastes and chlorhexidine toothpaste by inoculation bacteria on the toothpaste plates. The result showed that all the toothbrushes were contaminated with the following bacteria: *Roseomonas mucosa*, *Stenotrophomonas maltophilia*, *Pseudomonas aeruginosa*, *Leclercia adecarboxylata*, *Enterobacter asburiae*, *Candidatus Roseomonas massiliae*, *Pseudomonas parafulva*, *Bacillus licheniformis*, *Pseudomonas aeruginosa*, *Agrobacterium larrymoorei*, *Pantoea septica*, *Stenotrophomonas rhizophila*, *Citrobacter freundii* and *Pseudomonas frederiksbergensis*. The bristle surfaces, being rough, provided ample sites for trapping organisms. Examination of a brush revealed a biofilm on the brush head. The biofilm seen on the surface of the head to be composed of a compacted mixed community of microorganisms, including cocci, bacilli and filamentous organisms, together with cellular and debris. The toothpaste used proved antibacterial and inhibited bacterial growth, based mainly in the activity of fluoride which is widely used as an effective anticaries agent. In conclusion the isolated organisms are potentially pathogenic, particularly in relation to immunocompromised patients. The appropriate rinsing and drying of the toothbrushes before storage will however, likely reduce the incidence of these bacteria and the health risk associated with these pathogens.

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7th World Congress on Microbiology, November 28-29, 2016 Valencia, Spain

Influence of copper and plastic surfaces on the survival of bacteria in relation to the health care environment

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Metallic copper (Cu) surfaces have antimicrobial properties against a variety of different microorganisms and copper touch surfaces are likely to be increasingly used in public places including hospitals. Studies in the literature show that molecular mechanisms result in the rapid killing of Cu surface-exposed bacteria and yeasts result from a sharp shock of extreme and immediate Cu-ion overload combined with severe membrane and cell envelope damage, although similar low mutation rates have been observed in cells obtained from both Cu and control surfaces. The aim of this study was to determine the survival of bacteria on the surfaces of copper and plastic plumbing surfaces. The antibacterial activity of copper surfaces was determined by overlying suspensions of *Staphylococcus aureus* and *Escherichia coli* on copper and plastic surfaces. All pipes were sterilized and bacterial suspensions from colonies were prepared and then the pipes were contaminated by the bacterial suspension. The experiments were performed at 18-23°C and the results were assessed after a 20-day exposure. The numbers of viable bacteria in the suspension were determined by serial dilution and plating on Nutrient Agar plates; the plates being incubated at 37°C for 48 h. The results showed that low counts of *Staphylococcus aureus* were seen on copper surfaces, as compared with those obtained on the plastic, control surfaces, i.e., the results show that *E. coli* failed to survive on copper pipes. The number of bacteria isolated from the plastic surfaces was consistently higher than the number isolated from copper surfaces. The survival rate of bacteria on the copper surfaces was low and none of the inoculated bacteria survived after 20 days of exposure. Copper is well known to be an antibacterial, and its use in medical environments is likely to lead to the continuous reduction of environmental microbial contamination, including MRSA. The studies presented here show that the incorporation of Cu in healthcare facilities may dramatically help reduce the environmental microbial burden and act as a useful adjunct to current infection prevention and control systems, despite the fact that bacteria will eventually acquire resistance to the ion.

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8th CLINICAL MICROBIOLOGY CONFERENCE October 26-28, 2017 | Paris, France

Abstract

Bacteria and yeast can survive for 3 days on ceramic tiles.
Toilet air hand dryers can spread bacteria around the room and on the hands of the user.
The number of hand-contaminating bacteria from the hand dryer increases with time of use

Introduction

The hospital environment is a potential reservoir of bacterial pathogens living in associated with patients with diverse pathogenic microorganisms and a large number of susceptible individuals linked to significant morbidity and mortality. Bacterial pathogens have an innate ability to survive on surfaces in the hospital environment, often for long periods of time (1). Bacterial pathogens isolated from hospital environment are continuing to develop resistance to multiple antimicrobial agents. The emergence of multi-drug resistant organisms in hospital causes major problems in the treatment of nosocomial infections. The environment of patients is heavily contaminated by infectious multidrug resistant organisms, including methicillin-resistant *S. aureus* (MRSA), vancomycin-resistant enterococci (VRE), *Clostridium difficile*, *Acinetobacter baumannii* and *Pseudomonas aeruginosa* are an emerging, which caused well-known problem in healthcare systems (3).

Aims of the project

The aim of the work done in this study was a) to determine the occurrence of various bacteria and fungi on surfaces in the built environment, b) to determine the survival rates of these organisms on such surface environments, and c) determine factors that influence such survival.

Methods

- 1- Determine the survival rates of *Staphylococcus aureus* and *Escherichia coli* on types of sterilized ceramic tiles . The number of bacteria was determined from 3rd day, then 5, 15, and 20 days(2).
- 2- Assessment of the number of microorganisms on hands after washing and drying by warm air hand dryers. Then hands were touch the surface of nutrient agar plate and incubated at 37°C for 48 h and scored for the presence or absence of growing bacteria
- 3- Quantification of bacteria transferred from hand warm air dryers. The bacteria were counted and measured the effect of drying times (4).

References

- 1) Dancer, S. J. (2009). The role of environmental cleaning in the control of hospital -acquired infection. *J.Hosp.Infec.* 73, 378-385
- 2) Neely, A.N. and Maley, M.P. (2000). Survival of Enterococci and Staphylococci on Hospital Fabrics and Plastic. *J. Clin. Microbiol.* 38,724.
- 3) Talon, D. (1999). The role of the hospital environment in the epidemiology of multi-resistant bacteria. *J. Hosp. Infect.* 43: 13–17
- 4) Redway, K. and Fawdar, S. (2008) European Tissue Symposium: A Comparative Study of Three Different Hand Drying Methods: Paper Towel, Warm Air Dryer, Jet Air Dryer.

Results

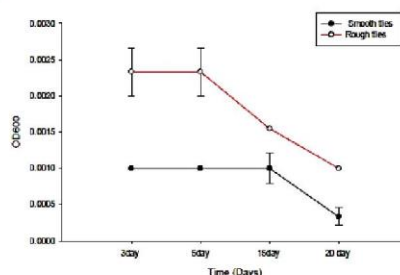


Figure 1: Survival of *S. aureus* on ceramic tiles under dry conditions

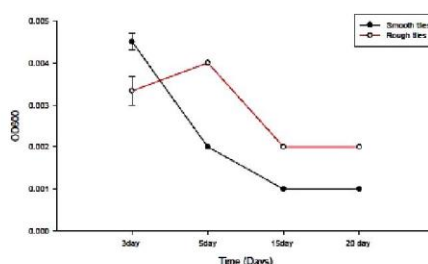


Figure 2: Survival of *E. coli* on ceramic tiles under dry conditions

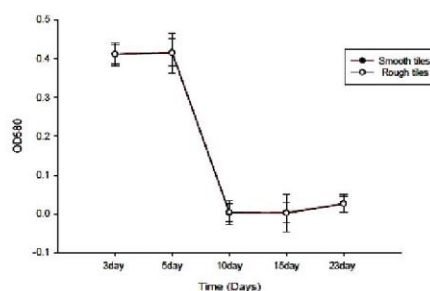


Figure 3: Survival of *Candida rugosa* on ceramic tiles under dry conditions

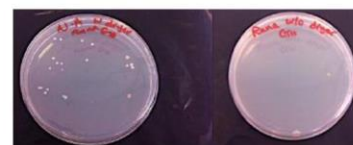


Figure 4: Agar plates used to assess the occurrence of bacteria on hands. (Left) hand washed and dried with a warm air dryer, (Right) hand washed and dried without a warm air dryer.

	WAD1	WAD2	WAD3
5 Sec	0	12	19
10 Sec	2	17	20
15 Sec	5	18	15
20 Sec	6	22	24
25 Sec	3	18	10
30 Sec	9	16	8
35 Sec	12	50	11
40 Sec	10	37	20
45 Sec	23	15	18
50 Sec	41	49	30
55 Sec	31	44	21
60 Sec	30	63	19

Table 1: The mean bacterial count using three different types warm air dryer (WAD) for different drying times.

Conclusion

To conclude, these environmental surfaces have been shown to carry both non-pathogenic and pathogenic bacteria. Therefore, a single hand contact with a contaminated surface could result in a variable degree of pathogen transfer. In this age of increasing antibiotic resistance, these survival data indicate that the appropriate disinfection of the environment and control procedures should be used to control of infections in hospitals.

Future work

- In the future, I will continue to isolate bacteria and *Candida* from environmental surface.
- Determine their survival rates and also the factors which influence this survival.
- I will also determine if antibiotic resistant bacteria are better able to survive than non-resistant types

Abstract

All the toothbrushes were contaminated with bacteria. Toothpastes which contains fluoride or chlorhexidine acts as an antibacterial and inhibits bacterial growth. The biofilm on the brush head to be composed of a compacted mixed community of microorganisms, including cocci, bacilli and filamentous organisms, together with cellular and other debris.

Introduction

Toothbrushes play an essential role in oral hygiene and are commonly found in community and hospital settings. The toothbrushes may act as a reservoir for potential pathogens transferred from the oral cavity, the bathroom environment, from storage containers and the water used for rinsing and the users. Contaminated toothbrushes have been suggested to play a role in both systemic and localized diseases. The possibility of toothbrushes being associated with the transmission of heart diseases, arthritis, bacteremia and stroke have also been reported.

Aims of the project

The aim of the work done in this study was to a) isolate, characterize and identify the bacterial contaminants on used manual toothbrushes. b) determine the antibacterial effect of toothpastes. C) Scanning electron microscopy (SEM) was used to visualise biofilms on toothbrush bristles.

Methods

- 1- Fifty used toothbrushes obtained from students and volunteers were analysed bacteriologically using standard microbiological techniques.
- 2- Scanning toothbrushes heads using SEM by immersing them in 2.5% glutaraldehyde in 0.1M sodium cacodylate buffer, pH7.3, for 60min. Brushes was rinsed, dehydrated in ethanol and dried from liquid CO₂. examined using a JEOL JSM-5300lv scanning electron microscope at an accelerating voltage of 10–30 kV.
- 3- Determination the antibacterial effect of toothpastes using seven types of commercial toothpastes and chlorhexidine toothpaste by inoculation *Staphylococcus aureus* and *Escherichia coli* on the toothpaste plates.

References

- 1- Frazelle, M.R. and Munro, C.L. (2012). Toothbrush contamination: a review of the literature. *Nursing Research and Practice*. Vol. 2012
- 2- Samuel, O. and Ifeanyi, O. (2015). Bacterial Contamination of Used Manual Toothbrushes Obtained from Some Students of Nnamdi Azikiwe University Awka, Nigeria. *Universal Journal of Microbiology Research* 3(4): 56-59
- 3- De Rossi, A., Ferreira, D., da Silva, R., de Queiroz, A., da Silva, L., Nelson-Filho, P. (2014). Antimicrobial Activity of Toothpastes Containing Natural Extracts, Chlorhexidine or Triclosan. *Brazilian Dental Journal* 25(3): 186-190
- 4- Marquis, R.E. (1995). Antimicrobial actions of fluoride for oral bacteria. *Canadian Journal of Microbiology*. 41: 955-964
- 5- Munir, A., Umar, J., Hameed, A. and Ahmed, S. (2005). The effect of commercially available local brand of toothpastes against oral bacteria. *Pakistan Oral and Dental Journal*. 25 (1):35-40

Results

Table (1): Bacterial Isolates from the Used Manual Toothbrushes

<i>Roseomonas mucosa</i>
<i>Stenotrophomonas maltophilia</i>
<i>Leclercia adedecarboxylata</i>
<i>Enterobacter asburiae</i>
<i>Candidatus Roseomonas massiliae</i>
<i>Pseudomonas parafulva</i>
<i>Bacillus licheniformis</i>
<i>Pseudomonas aeruginosa</i>
<i>Agrobacterium larrymoorei</i>
<i>Pantoea septic</i>
<i>Stenotrophomonas rhizophila</i>
<i>Citrobacter freundii</i>
<i>Pseudomonas frederiksbergensis</i>

Figure (1): Scanning electron microscopy of toothbrush biofilms on manual toothbrushes used for approximately 3 months. A) showing debris and cocci bacteria; B) showing debris and biofilm; C) image of tip of bristle, showing indentations and crevasses; down right) cocci embedded in the biofilm.

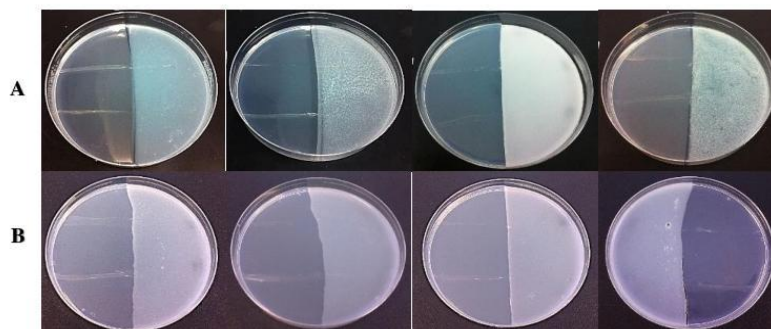
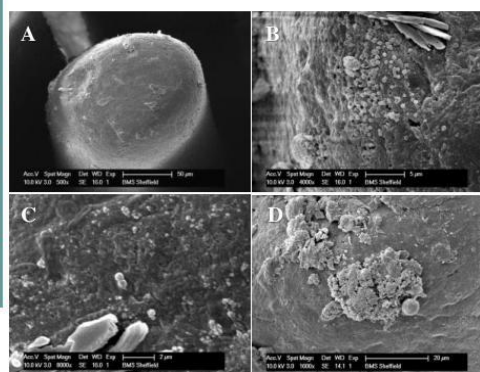


Figure (2) : A) commercial toothpastes agar plates that inoculated by *Staphylococcus aureus* and *Escherichia coli* and B) chlorhexidine toothpaste agar plates that inoculated by the same bacterial strains.

Conclusion

To conclude, the isolated organisms are potentially pathogenic, particularly in relation to immunocompromised patients. The appropriate rinsing and drying of the toothbrushes before storage will however, likely reduce the incidence of these bacteria and the health risk associated with these pathogens.

Future work

- In the future, I will continue to isolate bacteria and *Candida* from environmental surface.
- Determine their survival rates and also the factors which influence this survival.
- I will also determine if antibiotic resistant bacteria are better able to survive than non-resistant types.

APPINDIX 2

The phylogenetic analysis of bacteria isolated from environment samples

1- The phylogenetic analysis of bacteria isolated from sink samples

Klebsiella oxytoca CP011618.1

Klebsiella oxytoca strain CAV1335, complete genome

Sequence ID: [gb|CP011618.1](#) Length: 6229565 Number of Matches: 8

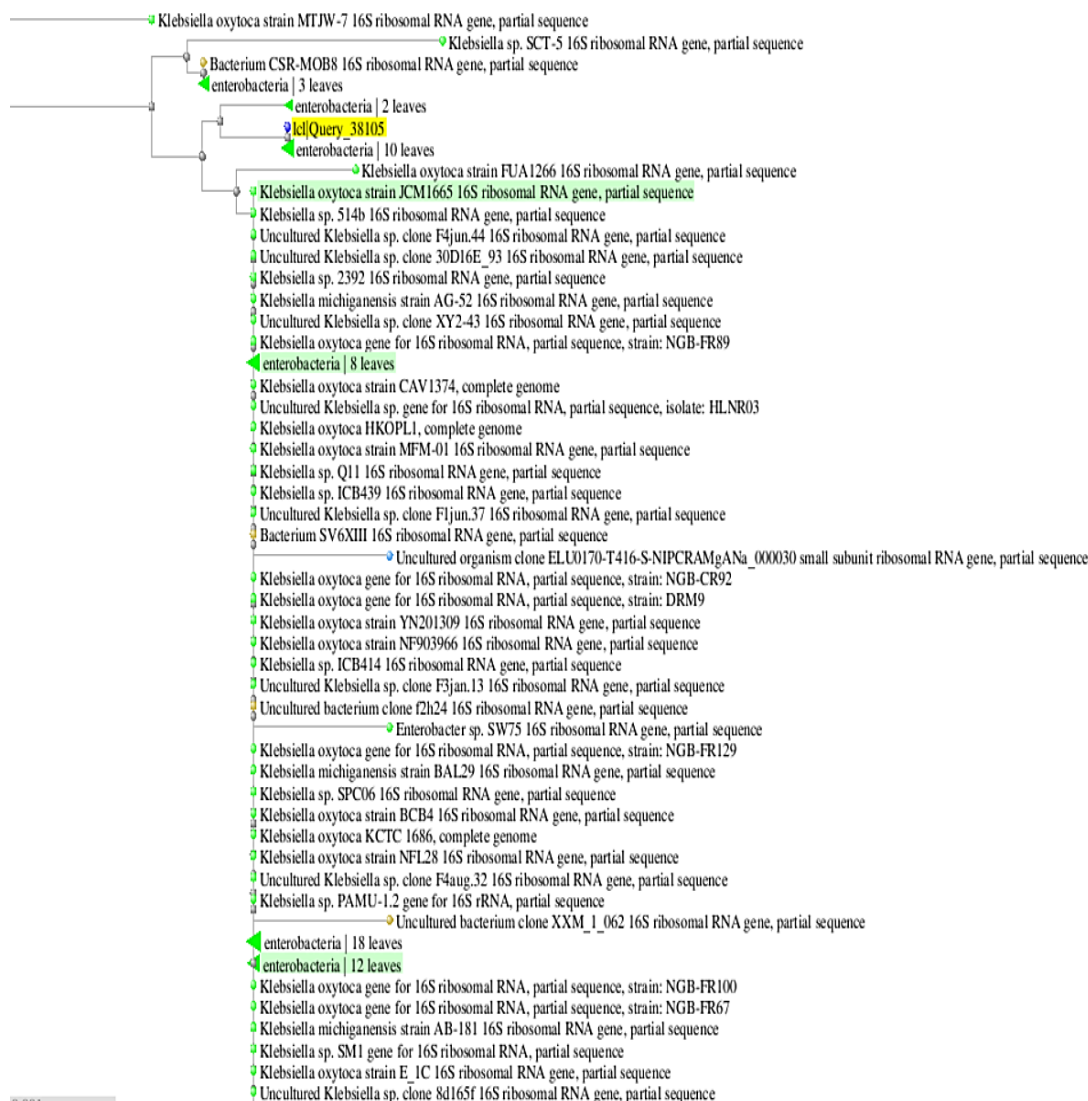
Range 1: 2928343 to 2929103 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1373 bits(1522)	0.0	761/761(100%)	0/761(0%)	Plus/Plus

Features: [SAM-dependent methyltransferase](#)
[rRNA-16S ribosomal RNA](#)

Query	1	TTTTGCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCGGGAACGTATTCA	60
Sbjct	2928343	TTTTGCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCGGGAACGTATTCA	2928402
Query	61	CCGTGGCATTCTGATCCACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGAC	120
Sbjct	2928403	CCGTGGCATTCTGATCCACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGAC	2928462
Query	121	TCCAATCCGGACTACGACATACTTTATGAGGTCGCTTGTCTCGCGAGGTCGCTTCTCT	180
Sbjct	2928463	TCCAATCCGGACTACGACATACTTTATGAGGTCGCTTGTCTCGCGAGGTCGCTTCTCT	2928522
Query	181	TTGTATATGCCATTGTAGCACGTGTGTAGCCCTACTCGTAAGGGCCATGATGACTTGACG	240
Sbjct	2928523	TTGTATATGCCATTGTAGCACGTGTGTAGCCCTACTCGTAAGGGCCATGATGACTTGACG	2928582
Query	241	TCATCCCCACCTTCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGACCTAATCG	300
Sbjct	2928583	TCATCCCCACCTTCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGACCTAATCG	2928642
Query	301	CTGGCAACAAAGGATAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTACAAACAC	360
Sbjct	2928643	CTGGCAACAAAGGATAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTACAAACAC	2928702
Query	361	GAGCTGACGACAGCCATGCAGCACCTGTCTCAGAGTTCCCGAAGGCACCAAAGCATCTCT	420
Sbjct	2928703	GAGCTGACGACAGCCATGCAGCACCTGTCTCAGAGTTCCCGAAGGCACCAAAGCATCTCT	2928762
Query	421	GCTAAGTTCTCTGGATGTCAAGAGTAGGTAAGGTTCTTCGCGTTGCATCGAATTAAACCA	480
Sbjct	2928763	GCTAAGTTCTCTGGATGTCAAGAGTAGGTAAGGTTCTTCGCGTTGCATCGAATTAAACCA	2928822
Query	481	CATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCATTTGAGTTTTAACCTTGCGGCCGT	540
Sbjct	2928823	CATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCATTTGAGTTTTAACCTTGCGGCCGT	2928882
Query	541	ACTCCCCAGGCGGTGCGACTTAACGCGTTAGCTCCGGAAGCCACTCCTCAAGGGAACAACC	600
Sbjct	2928883	ACTCCCCAGGCGGTGCGACTTAACGCGTTAGCTCCGGAAGCCACTCCTCAAGGGAACAACC	2928942
Query	601	TCCAAGTCGACATCGTTTACAGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCA	660
Sbjct	2928943	TCCAAGTCGACATCGTTTACAGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCA	2929002
Query	661	CGCTTTCGCACCTGAGCGTCAGTCTTTGTCCAGGGGGCCGCCTTCGCCACCGGTATTCTCT	720
Sbjct	2929003	CGCTTTCGCACCTGAGCGTCAGTCTTTGTCCAGGGGGCCGCCTTCGCCACCGGTATTCTCT	2929062
Query	721	CCAGATCTCTACGCATTTACCGCTACACCTGGAATTCTAC	761
Sbjct	2929063	CCAGATCTCTACGCATTTACCGCTACACCTGGAATTCTAC	2929103



Bacillus subtilis KP340123.1

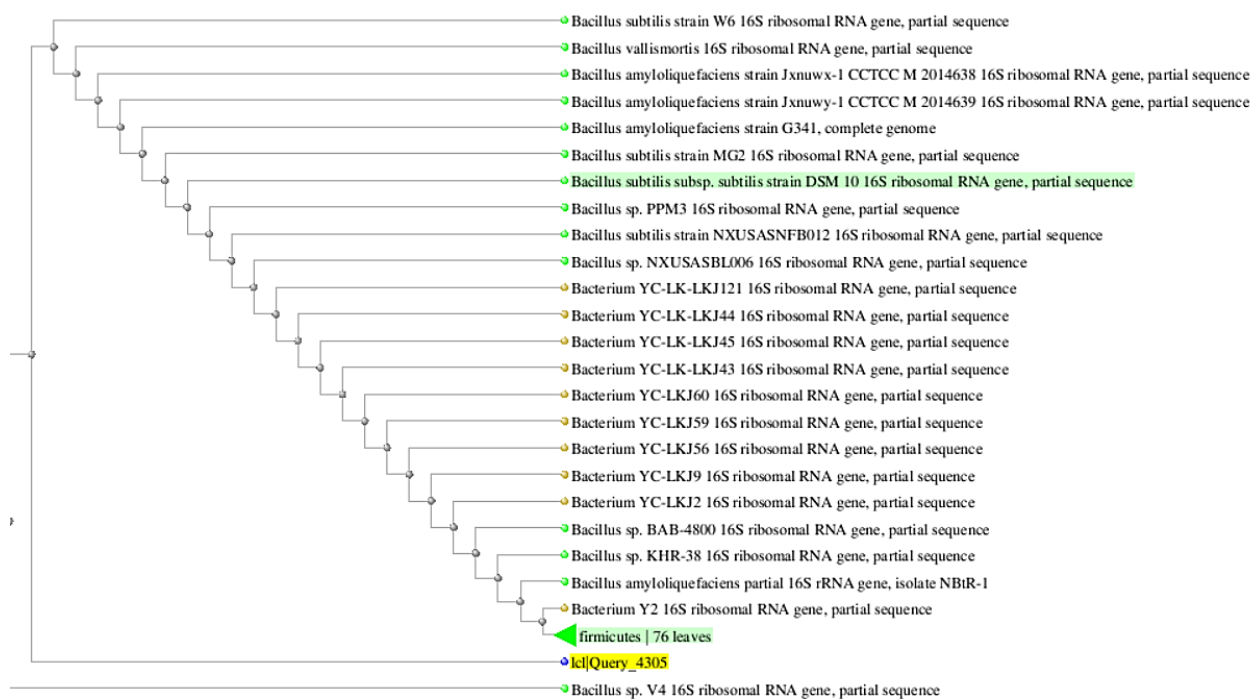
Bacillus subtilis strain W6 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP340123.1|](#) Length: 1431 Number of Matches: 1

Range 1: 467 to 1367 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1626 bits(1802)	0.0	901/901(100%)	0/901(0%)	Plus/Minus
Query 1	TGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTGATCCGCGATT	60		
Sbjct 1367	TGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTGATCCGCGATT	1308		
Query 61	ACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACTGAGAACAGAT	120		
Sbjct 1307	ACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACTGAGAACAGAT	1248		
Query 121	TTGTGGGATTGGCTTAACCTCGCGGTTTCGCTGCCCTTTGTTCTGTCCATTGTAGCACGT	180		
Sbjct 1247	TTGTGGGATTGGCTTAACCTCGCGGTTTCGCTGCCCTTTGTTCTGTCCATTGTAGCACGT	1188		
Query 181	GTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCCGGTTT	240		
Sbjct 1187	GTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCCGGTTT	1128		
Query 241	GTCACGGCGAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGATCAAGGGTTGC	300		
Sbjct 1127	GTCACGGCGAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGATCAAGGGTTGC	1068		
Query 301	GCTCGTTGCGGGACTTAACCCAACATCTCAGACACGAGCTGACGACAACCATGCACCAC	360		
Sbjct 1067	GCTCGTTGCGGGACTTAACCCAACATCTCAGACACGAGCTGACGACAACCATGCACCAC	1008		
Query 361	CTGTCACTCTGCCCCGAAGGGGACGTCCTATCTCTAGGATTGTCAGAGGATGTCAAGAC	420		
Sbjct 1007	CTGTCACTCTGCCCCGAAGGGGACGTCCTATCTCTAGGATTGTCAGAGGATGTCAAGAC	948		
Query 421	CTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGGCC	480		
Sbjct 947	CTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGGCC	888		
Query 481	CCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGCGGAGTGCTTAATG	540		
Sbjct 887	CCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGCGGAGTGCTTAATG	828		
Query 541	CGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCATCGTTACGGC	600		
Sbjct 827	CGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCATCGTTACGGC	768		
Query 601	GTGGACTACCAGGGTATCTAATCCTGTTTCGCTCCCCACGCTTTCGCTCCTCAGCGTCAGT	660		
Sbjct 767	GTGGACTACCAGGGTATCTAATCCTGTTTCGCTCCCCACGCTTTCGCTCCTCAGCGTCAGT	708		
Query 661	TACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACATCTCTACGCATTTACCG	720		
Sbjct 707	TACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACATCTCTACGCATTTACCG	648		
Query 721	CTACACGTGGAATTCCACTCTCCTCTTCTGCACTCAAGTTCCCGAGTTTCCAATGACCCT	780		
Sbjct 647	CTACACGTGGAATTCCACTCTCCTCTTCTGCACTCAAGTTCCCGAGTTTCCAATGACCCT	588		
Query 781	CCCCGGTTGAGCCGGGGGCTTTACATCAGACTTAAGAAACCGCCTGCGAGCCCTTTACG	840		
Sbjct 587	CCCCGGTTGAGCCGGGGGCTTTACATCAGACTTAAGAAACCGCCTGCGAGCCCTTTACG	528		
Query 841	CCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAGTT	900		
Sbjct 527	CCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAGTT	468		
Query 901	A 901			
Sbjct 467	A 467			



Kocuria rhizophila KM978822.1

Kocuria rhizophila strain AHT-1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KM978822.1](#) Length: 1491 Number of Matches: 1

Range 1: 475 to 1394 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1653 bits(1832)	0.0	920/921(99%)	1/921(0%)	Plus/Minus
Query 1	CCAACCTTTCGTGACTTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCAGCG	60		
Sbjct 1394	CCAACCTTTCGTGACTTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCAGCG	1335		
Query 61	TTGCTGATCTGCGATTACTAGCGACTCCGACTTCACGTGGTCGAGTTGCAGACCACGATC	120		
Sbjct 1334	TTGCTGATCTGCGATTACTAGCGACTCCGACTTCACGTGGTCGAGTTGCAGACCACGATC	1275		
Query 121	CGAACTGAGACCAGCTTTTTGGGATTAGCTCCACCTCGCGGCATCGCAACCCATTGTACT	180		
Sbjct 1274	CGAACTGAGACCAGCTTTTTGGGATTAGCTCCACCTCGCGGCATCGCAACCCATTGTACT	1215		
Query 181	GGCCATTGTAGCATGCGTGAAGCCCAAGACATAAGGGGCATGATGATTGACGTCATCCT	240		
Sbjct 1214	GGCCATTGTAGCATGCGTGAAGCCCAAGACATAAGGGGCATGATGATTGACGTCATCCT	1155		
Query 241	CACCTTCCTCCGAGTTGACCCCGGCGAGTCTCCTATGAGTCCCCACCATCACGTGCTGGCA	300		
Sbjct 1154	CACCTTCCTCCGAGTTGACCCCGGCGAGTCTCCTATGAGTCCCCACCATCACGTGCTGGCA	1095		
Query 301	ACATAGAACGAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTG	360		
Sbjct 1094	ACATAGAACGAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTG	1035		
Query 361	ACGACAACCATGCACCACCTGTACACCAGCCCCACAAGGGGGAAGACCATCTCTGGCCC	420		
Sbjct 1034	ACGACAACCATGCACCACCTGTACACCAGCCCCACAAGGGGGAAGACCATCTCTGGCCC	975		
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Query 481	CTCCGCCGCTTGTGCGGGCCCCCGTCAATTCCCTTTAGTCTTTAGCCTTGCGGCCGTACTC	540		
Sbjct 914	CTCCGCCGCTTGTGCGGGCCCCCGTCAATTCCCTTTAGTCTTTAGCCTTGCGGCCGTACTC	855		
Query 541	CCCAGGCGGGGCACCTTAATGCGTTAGCTACGGCGCGGAAAACGTGGAATGTTCCCCACAC	600		
Sbjct 854	CCCAGGCGGGGCACCTTAATGCGTTAGCTACGGCGCGGAAAACGTGGAATGTTCCCCACAC	795		
Query 601	CTAGTGCCCAACGTTTACGGCATGGACTACCAGGGTATCTAATCCTGTTGCTCCCCATG	660		
Sbjct 794	CTAGTGCCCAACGTTTACGGCATGGACTACCAGGGTATCTAATCCTGTTGCTCCCCATG	735		
Query 661	CTTTCGCTCCTCAGCGTCAGTAACAGCCCAGAGACCTGCCTTCGCCATCGGTGTTCTCTCC	720		
Sbjct 734	CTTTCGCTCCTCAGCGTCAGTAACAGCCCAGAGACCTGCCTTCGCCATCGGTGTTCTCTCC	675		
Query 721	TGATATCTGCGCATTTACCGCTACACCAGGAATTCAGTCTCCCCCTACTGCACTCAAGT	780		
Sbjct 674	TGATATCTGCGCATTTACCGCTACACCAGGAATTCAGTCTCCCCCTACTGCACTCAAGT	615		
Query 781	CTGCCCCTACCCACTGCACACCCGGGGTTAAGCCCCGGGCTTTCACAGCAGACGCGACAA	840		
Sbjct 614	CTGCCCCTACCCACTGCACACCCGGGGTTAAGCCCCGGGCTTTCACAGCAGACGCGACAA	555		
Query 841	ACCGCCTACGAGCTCTTTACGCCCAATAATTCCCGGACAACGCTTGCGCCCTACGTATTA	900		
Sbjct 554	ACCGCCTACGAGCTCTTTACGCCCAATAATTCCCGGACAACGCTTGCGCCCTACGTATTA	496		
Query 901	CCGCGGCTGCTGGCACGTAGT	921		
Sbjct 495	CCGCGGCTGCTGGCACGTAGT	475		



Bacillus cereus KC731425.1

Bacillus cereus strain VITSH1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KC731425.1](#) Length: 1369 Number of Matches: 1

Range 1: 513 to 1353		GenBank	Graphics			Next Match	Previous Match
Score	Expect	Identities	Gaps	Strand			
1283 bits(1422)	0.0	789/841(94%)	0/841(0%)	Plus/Minus			
Query	1	ACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTGATCCGCGATTAC					60
Sbjct	1353	ACGGGCGGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTGATCCGCGATTAC					1294
Query	61	TAGCGATTCCAGCTTCATGTATGCAAGTTGCAACCGACAATCCAACTGAAAACGGTTTT					120
Sbjct	1293	TAGCGATTCCAGCTTCAGGTAGGCAAGTTGCAGCTACAATCCAACTGAAAACGGTTTT					1234
Query	121	ATGATATTAGCTCCACCTCGCGGTCTTGCACTCTTTGTACCGTCCATTGTAACACGTGT					180
Sbjct	1233	ATGAAATTAGCTCCACCTCGCGGTCTTGCACTCTTTGTACCGTCCATTGTAGCACGTGT					1174
Query	181	GTACCCAGGTCATAGGGGGCATGATGATTGACGTATCCACCTTCTCCGGTTTGT					240
Sbjct	1173	GTAGCCAGGTCATAAGGGGCATGATGATTGACGTATCCACCTTCTCCGGTTTGT					1114
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Sbjct	1113	CACCGGCAGTCACCTTAAAGTGCCCAACTAAATGATGGCAACTAAAATCAAGGGTTGCGC					1054
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Sbjct	1053	TCGTTGCGGGACTTAACCCAAACATCTCACAACACAAGCTGACAACAACCATGCACCACCT					994
Query	361	GTCACCTCTGTCCCGAAGGAAAAGACCTATCTCTAGGGTTTTACAGGATGTCAAGAGCT					420
Sbjct	993	GTCACCTCTGTCCCGAAGGAAAAGCCCTATCTCTAGGGTTGTCAAAGGATGTCAAGACCT					934
Query	421	GGTAAGGGTCTTCCCGTTGCTTCCAATTAAACCACATGCTCCACCGCTTGGGCGGGCCCC					480
Sbjct	933	GGTAAGGTTCTTCGCGTTGCTTCAAATTAAACCACATGCTCCACCGCTTGTGCGGGCCCC					874
Query	481	CGTCAATTCCTTTGAATTTCAACCTTGCGGACGTACTCCCCAGGGGAATGGTTAATGCG					540
Sbjct	873	CGTCAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCCAGGCGGAGTGCTTAATGCG					814
Query	541	TTAACTTCGCACTAAAGGACGGAACCCCTTAACACTTACCACCTCATCGTTACGGCGG					600
Sbjct	813	TTAACTTCAGCACTAAAGGGCGGAAACCCCTTAACACTTAGCACTCATCGTTACGGCGT					754
Query	601	GAAC TACCAGGGTATCTAATCCGGTTTGCTCCCCACGCTTTCGCGCCTCACTGTCAAGTTA					660
Sbjct	753	GAAC TACCAGGGTATCTAATCCGGTTTGCTCCCCACGCTTTCGCGCCTCATGTCAAGTTA					694
Query	661	GAGACCAGAAAGTCCCTTCGCCACTGGGGTTCCTCCATATCTCTACACATTTACCGCT					720
Sbjct	693	CAGACCAAAAGTCCCTTCGCCACTGGTGTTCCTCCATATCTCTACGCATTTACCGCT					634
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Sbjct	633	ACACATGGAATTCACCTTTCTCTCTGCACTCAAGTCTCCAGTTTCCAATGACCTCC					574
Query	781	GGGGTTGACCCGGGGGCTTTCCCTCAAACCTAAAAAACACCTGCGCGCGCTTTACCCC					840
Sbjct	573	ACGGTTGAGCCGTGGGCTTTCACATCAGACTTAAGAAACACCTGCGCGCGCTTTACGCC					514
Query	841	C 841					
Sbjct	513	C 513					

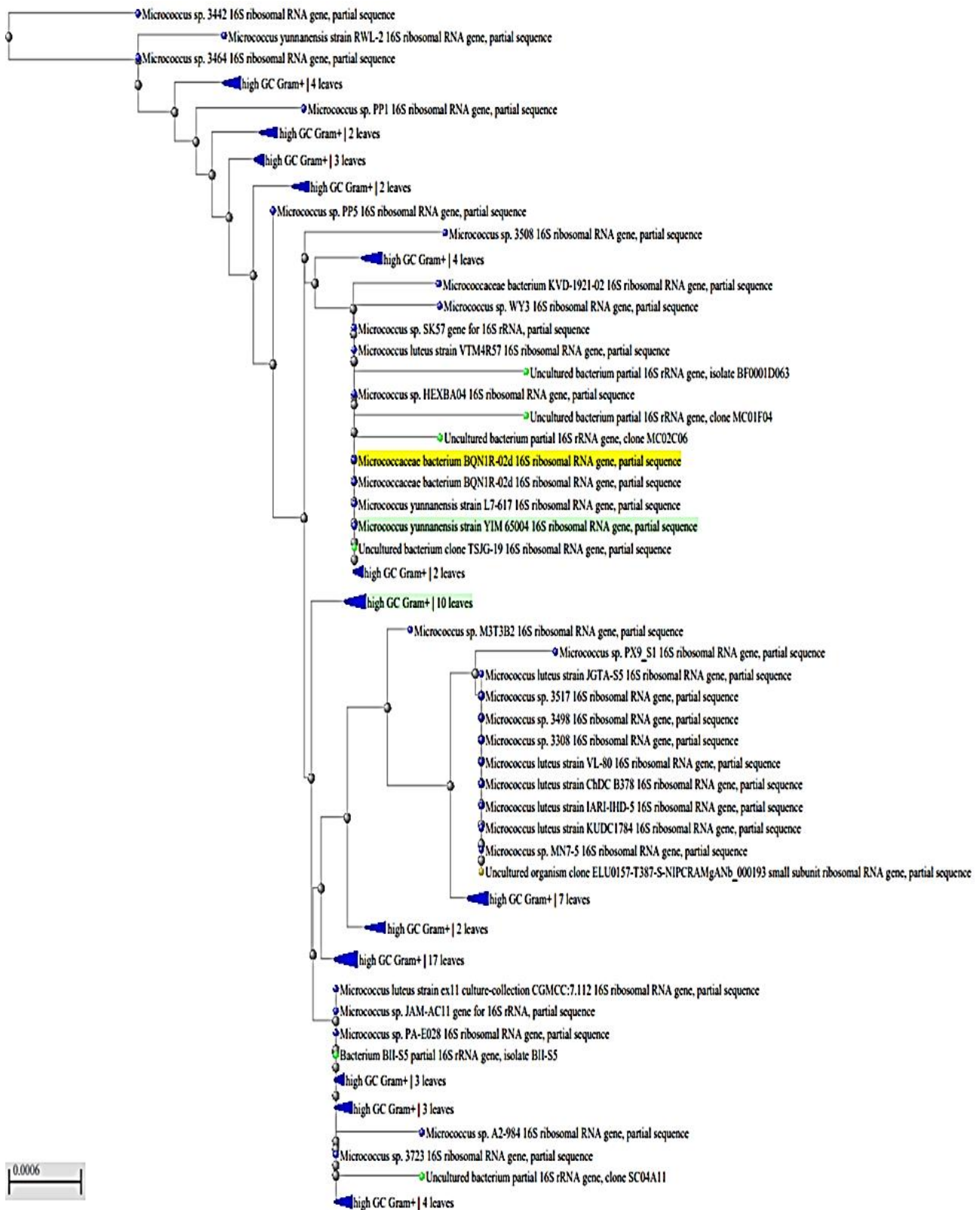


Micrococcus luteus KT339390

Micrococcus luteus strain DYPSBB RPF YRP01 16S ribosomal RNA gene, partial sequence
Sequence ID: [KT339390.1](#) Length: 1462 Number of Matches: 1

Range 1: 30 to 1424 [GenBank](#) [Graphics](#) [Next Match](#) [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
2560 bits(1386)	0.0	1393/1396(99%)	2/1396(0%)	Plus/Plus
Query 2	ACATGCAAGTCGAACGATGAAGCCCAGCTTGCTGGGTGGATTAGTGGCGAACGGGTGAGT	61		
Sbjct 30	ACATGCAAGTCGAACGATGAAGCCCAGCTTGCTGGGTGGATTAGTGGCGAACGGGTGAGT	89		
Query 62	AACACGTGAGTAACCTGCCCTTAACCTCTGGGATAAGCCTGGGAACTGGGTCTAATACCG	121		
Sbjct 90	AACACGTGAGTAACCTGCCCTTAACCTCTGGGATAAGCCTGGGAACTGGGTCTAATACCG	149		
Query 122	GATAGGAGCGTCCACCGCATGGTGGGTGTTGGAAAGATTATCGGTTTTGGATGGACTCG	181		
Sbjct 150	GATAGGAGCGTCCACCGCATGGTGGGTGTTGGAAAGATTATCGGTTTTGGATGGACTCG	209		
Query 182	CGGCCTATCAGCTTGTTGGTGAGGTAATGGCTCACCAAGGCGACGACGGGTAGCCGGCCT	241		
Sbjct 210	CGGCCTATCAGCTTGTTGGTGAGGTAATGGCTCACCAAGGCGACGACGGGTAGCCGGCCT	269		
Query 242	GAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCA	301		
Sbjct 270	GAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCA	329		
Query 302	GTGGGGAATATTGCACAATGGGCGAAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACG	361		
Sbjct 330	GTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACG	389		
Query 362	GCCTTCGGGTTGTAAACCTCTTTAGTAGGGGAAGCGAAAGTGACGGTACCTGCAGAA	421		
Sbjct 390	GCCTTCGGGTTGTAAACCTCTTTAGTAGGGGAAGCGAAAGTGACGGTACCTGCAGAA	449		
Query 422	GAAGCACCAGGCTAACTACGTGCCAGCAGCCGCGGTAAATACGTAGGGTGCGAGCGTTATCC	481		
Sbjct 450	GAAGCACCAGGCTAACTACGTGCCAGCAGCCGCGGTAAATACGTAGGGTGCGAGCGTTATCC	509		
Query 482	GGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGTTTGTGCGCTCTGTCGTGAAAGTCCGG	541		
Sbjct 510	-GGGAATTATTGGGCGTAAAGAGCTCGTAGGCGGTTTGTGCGCTCTGTCGTGAAAGTCCGG	568		
Query 542	GGCTTAACCCCGGATCTGCGGTGGGTACGGGAGACTAGAGTGAGTGGGGAGACTGGA	601		
Sbjct 569	GGCTTAACCCCGGATCTGCGGTGGGTACGGGAGACTAGAGTGAGTGGGGAGACTGGA	628		
Query 602	ATTCTGGTGTAGCGGTGGAATGCGCAGATATCAGGAGGAACACCGATGGCGAAGGCAGG	661		
Sbjct 629	ATTCTGGTGTAGCGGTGGAATGCGCAGATATCAGGAGGAACACCGATGGCGAAGGCAGG	688		
Query 662	TCTCTGGGCTGTAACTGACGCTGAGGAGCGAAAGCATGGGGAGCGAACAGGATTAGATAC	721		
Sbjct 689	TCTCTGGGCTGTAACTGACGCTGAGGAGCGAAAGCATGGGGAGCGAACAGGATTAGATAC	748		
Query 722	CCTGGTAGTCCATGCCGTAAACGTTGGGCACTAGGTGTGGGGACCATTCACCGGTTTCCG	781		
Sbjct 749	CCTGGTAGTCCATGCCGTAAACGTTGGGCACTAGGTGTGGGGACCATTCACCGGTTTCCG	808		
Query 782	CGCCGCAGCTAACGCATTAAAGTCCCCCGCTGGGGAGTACGGCCGCAAGGCTAAAACTCA	841		
Sbjct 809	CGCCGCAGCTAACGCATTAAAGTCCCCCGCTGGGGAGTACGGCCGCAAGGCTAAAACTCA	868		
Query 842	AAGGAATTGACGGGGGCCGACAAAGCGGCGGAGCATGCGGATTAAATTCGATGCAACGCG	901		
Sbjct 869	AAGGAATTGACGGGGGCCGACAAAGCGGCGGAGCATGCGGATTAAATTCGATGCAACGCG	928		
Query 902	AAGAACCTTACCAAGGCTTGACATGTTCTCGATCGCCGTAGAGATACGGTTTCCCCTTTG	961		
Sbjct 929	AAGAACCTTACCAAGGCTTGACATGTTCTCGATCGCCGTAGAGATACGGTTTCCCCTTTG	988		
Query 962	GGGCGGGTTTACAGGTTGGTGCATGGTTGTGCTGAGCTCGTGTGCTGAGATGTTGGGTTAA	1021		
Sbjct 989	GGGCGGGTTTACAGGTTGGTGCATGGTTGTGCTGAGCTCGTGTGCTGAGATGTTGGGTTAA	1048		
Query 1022	GTCCCGCAACGAGCGCAACCTCTGTTCCATGTTGCCAGCACGTAATGGTGGGGACTCATG	1081		
Sbjct 1049	GTCCCGCAACGAGCGCAACCTCTGTTCCATGTTGCCAGCACGTAATGGTGGGGACTCATG	1108		
Query 1082	GGAGACTGCCGGGGTCAACTCGGAGGAAGGTGAGGACGACGTCAAATCATCATGCCCCCTT	1141		
Sbjct 1109	GGAGACTGCCGGGGTCAACTCGGAGGAAGGTGAGGACGACGTCAAATCATCATGCCCCCTT	1168		
Query 1142	ATGTCTTGGGCTTACGCATGCTACAATGGCCGGTACAATGGGTTGCGATACTGTGAGGT	1201		
Sbjct 1169	ATGTCTTGGGCTTACGCATGCTACAATGGCCGGTACAATGGGTTGCGATACTGTGAGGT	1228		
Query 1202	GGAGCTAATCCCAAAAAGCCGGTCTCAGTTTCGGATTGGGGTCTGCAACTCGACCCCATGA	1261		
Sbjct 1229	GGAGCTAATCCCAAAAAGCCGGTCTCAGTTTCGGATTGGGGTCTGCAACTCGACCCCATGA	1288		
Query 1262	AGTCGGAGTCGCTAGTAATCGCAGATCAGCAACGCTGCGGTGAATACGTTCCCGGGCCTT	1321		
Sbjct 1289	AGTCGGAGTCGCTAGTAATCGCAGATCAGCAACGCTGCGGTGAATACGTTCCCGGGCCTT	1348		
Query 1322	GTACACACCGCCCGTCAAGTCACGAAAGTTGGTAACACCCGAAGCCGGTGGCCTAACCTT	1381		
Sbjct 1349	GTACACACCGCCCGTCAAGTCACGAAAGTTGGTAACACCCGAAGCCGGTGGCCTAACCTT	1408		
Query 1382	TGTGGGGG-AGCCGTC	1396		
Sbjct 1409	TGTGGGGGGAAGCCGTC	1424		



2- The phylogenetic analysis of bacteria isolated from mirror samples

Bacillus cereus KP100400.1

Bacillus cereus 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP100400.1|](#) Length: 1179 Number of Matches: 1

Range 1: 55 to 1014 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1683 bits(1866)	0.0	949/960(99%)	0/960(0%)	Plus/Plus
Query 1	GGCGGACGGGTGAGTAACACGTGGGTAACCTGCCATAAGACTGGGATAACTCCGGGAAA	60		
Sbjct 55	GGCGGACGGGTGAGTAACACGTGGGTAACCTGCCATAAGACTGGGATAACTCCGGGAAA	114		
Query 61	CCGGGGCTAATACCGGATAACATTTTGAACCGCATGGTTCGAAATTGAAAGGCGGCTTCG	120		
Sbjct 115	CCGGGGCTAATACCGGATAACATTTTGAACCGCATGGTTCGAAATTGAAAGGCGGCTTCG	174		
Query 121	GCTGTCACTTATGGATGGACCCGCGTCGCATTAGCTAGTTGGTGAGGTAACGGCTCACCA	180		
Sbjct 175	GCTGTCACTTATGGATGGACCCGCGTCGCATTAGCTAGTTGGTGAGGTAACGGCTCACCA	234		
Query 181	AGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGC	240		
Sbjct 235	AGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGC	294		
Query 241	CCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGGA	300		
Sbjct 295	CCAAACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGGA	354		
Query 301	GCAACGCCGCGTGAGTGATGAAGGCTTTCGGGTCGTAAACTCTGTTGTTAGGGAAGAAC	360		
Sbjct 355	GCAACGCCGCGTGAGTGATGAAGGCTTTCGGGTCGTAAACTCTGTTGTTAGGGAAGAAC	414		
Query 361	AAGTGCTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACGAGAAAGCCACGGCTAACT	420		
Sbjct 415	AAGTGCTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACGAGAAAGCCACGGCTAACT	474		
Query 421	ACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATGGGCGTA	480		
Sbjct 475	ACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATGGGCGTA	534		
Query 481	AAGCGCGCGCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCACGGCTCANCCGTGGAGGG	540		
Sbjct 535	AAGCGCGCGCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGAGGG	594		
Query 541	TCATTGGAAGCTGGGAGACTTGAGTGCAGAAGAGGAAAGTGAATTCCATGTGTAGCGGT	600		
Sbjct 595	TCATTGGAAGCTGGGAGACTTGAGTGCAGAAGAGGAAAGTGAATTCCATGTGTAGCGGT	654		
Query 601	GAAATGCGTAGAGATATGGAGGAACACCACTGGCGAAGGCGACTTTCTGGTCTGTAAGTG	660		
Sbjct 655	GAAATGCGTAGAGATATGGAGGAACACCACTGGCGAAGGCGACTTTCTGGTCTGTAAGTG	714		
Query 661	ACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCG	720		
Sbjct 715	ACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCG	774		
Query 721	TAAACGATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGCATT	780		
Sbjct 775	TAAACGATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGCATT	834		
Query 781	AAGCACTCCGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGACGGGGGCC	840		
Sbjct 835	AAGCACTCCGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGACGGGGGCC	894		
Query 841	CGCACAAAGCGGGGAGCATGTGGTTTAATTGGAAGCAACGCAAAAAACCTTACCAGGTCT	900		
Sbjct 895	CGCACAAAGCGGTGGAGCATGTGGTTTAATTGGAAGCAACGCAAAAAACCTTACCAGGTCT	954		
Query 901	TGACATCCTCCGACAACCCCTAAAGAAAGGGCTTCCCTTCCGGAACAAAATGACAGGGGG	960		
Sbjct 955	TGACATCCTCTGAAAACCCCTAAAGAAAGGGCTTCCCTTCCGGAACAAAATGACAGGTGG	1014		

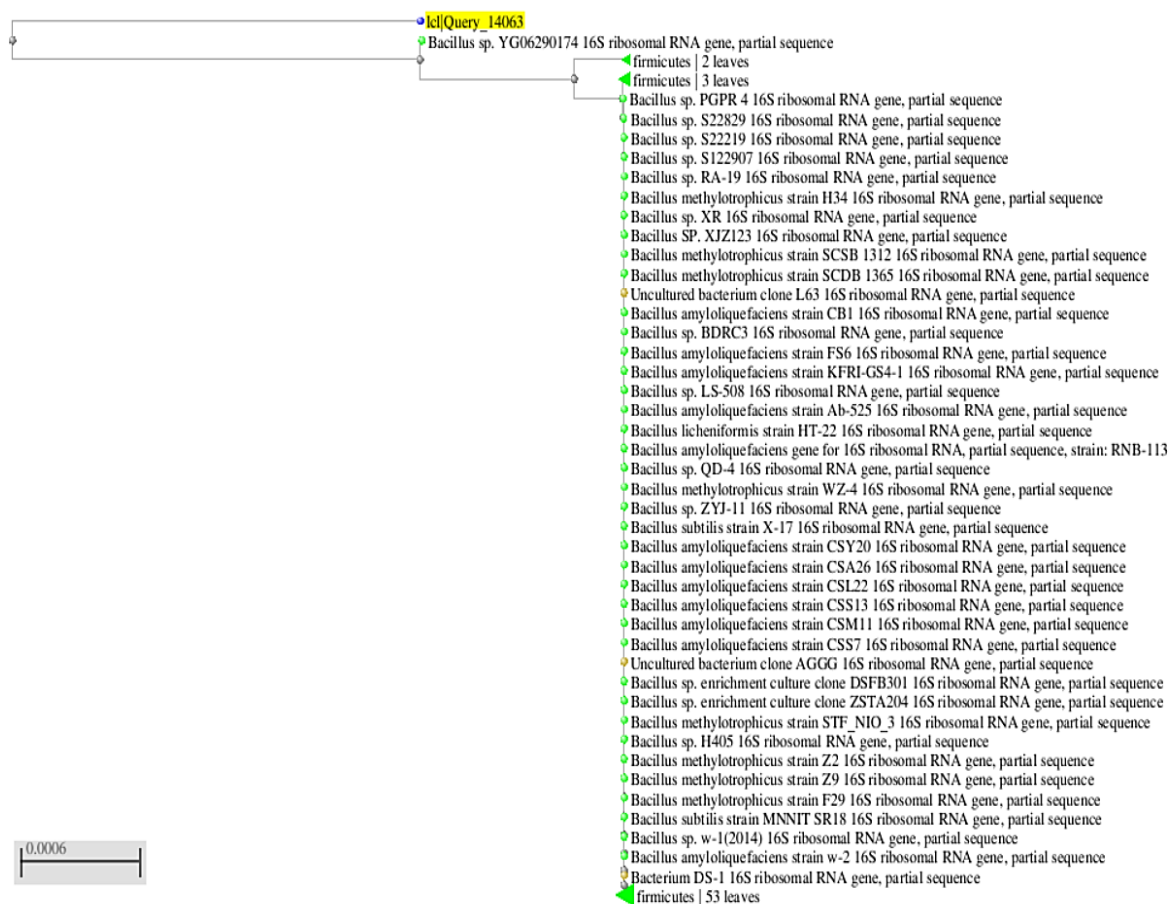


Bacillus subtilis DQ683077.1

Bacillus subtilis strain GB03 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|DQ683077.1](#) Length: 1124 Number of Matches: 1

Range 1: 26 to 1004		GenBank	Graphics	▼ Next Match ▲ Previous Match	
Score	Expect	Identities	Gaps	Strand	
1739 bits(1928)	0.0	973/979(99%)	0/979(0%)	Plus/Plus	
Query	1	AGATGGGAGCTTGCTCCCTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTG			60
Sbjct	26	AGATGGGAGCTTGCTCCCTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTG			85
Query	61	CCTGTAAGACTGGGATAACTCCGGGAAACCGGGGCTAATACCGGATGGTTGTTTGAACCG			120
Sbjct	86	CCTGTAAGACTGGGATAACTCCGGGAAACCGGGGCTAATACCGGATGGTTGTTTGAACCG			145
Query	121	CATGGTTTCAGACATAAAAGGTGGCTTCGGCTACCACTTACAGATGGACCCGCGGCGCATT			180
Sbjct	146	CATGGTTTCAGACATAAAAGGTGGCTTCGGCTACCACTTACAGATGGACCCGCGGCGCATT			205
Query	181	AGCTAGTTGGTGAGGTAACGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTG			240
Sbjct	206	AGCTAGTTGGTGAGGTAACGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTG			265
Query	241	ATCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCAGTAGGGAAT			300
Sbjct	266	ATCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCAGTAGGGAAT			325
Query	301	CTTCCGCAATGGACGAAAGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTCGGA			360
Sbjct	326	CTTCCGCAATGGACGAAAGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTCGGA			385
Query	361	TCGTAAAGCTCTGTTGTTAGGGAAGAACAAGTGCCGTTCAAATAGGGCGGCACCTTGACG			420
Sbjct	386	TCGTAAAGCTCTGTTGTTAGGGAAGAACAAGTGCCGTTCAAATAGGGCGGCACCTTGACG			445
Query	421	GTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGG			480
Sbjct	446	GTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGG			505
Query	481	CAAGCGTTGTCCGGAATTATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATG			540
Sbjct	506	CAAGCGTTGTCCGGAATTATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATG			565
Query	541	TGAAAGCCCCCGGCTCAACCGGGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCAAG			600
Sbjct	566	TGAAAGCCCCCGGCTCAACCGGGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCAAG			625
Query	601	AGGAGAGTGGAATTCACGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCACTG			660
Sbjct	626	AGGAGAGTGGAATTCACGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCACTG			685
Query	661	GCGAAGGCGACTCTCTGGTCTGTAACGTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACA			720
Sbjct	686	GCGAAGGCGACTCTCTGGTCTGTAACGTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACA			745
Query	721	GGATTAGATACCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTC			780
Sbjct	746	GGATTAGATACCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTC			805
Query	781	CGCCCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGA			840
Sbjct	806	CGCCCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGA			865
Query	841	CTGAAACTCAAAGGAATTGACGGGGGCCGCACAAGCGGTGGAGCATGTGGTTTAATTCTG			900
Sbjct	866	CTGAAACTCAAAGGAATTGACGGGGGCCGCACAAGCGGTGGAGCATGTGGTTTAATTCTG			925
Query	901	AAGCAACGCGAGAAACCTTACCAGGTCTTGACATCCTCTGACATCCTAAAGAAAGGACGTC			960
Sbjct	926	AAGCAACGCGAGAAACCTTACCAGGTCTTGACATCCTCTGACATCCTAGAGATAGGACGTC			985
Query	961	CCCTTCGGGGGCAAAATGA	979		
Sbjct	986	CCCTTCGGGGGCGAGAGTGA	1004		



Bacillus cereus JQ659737.1

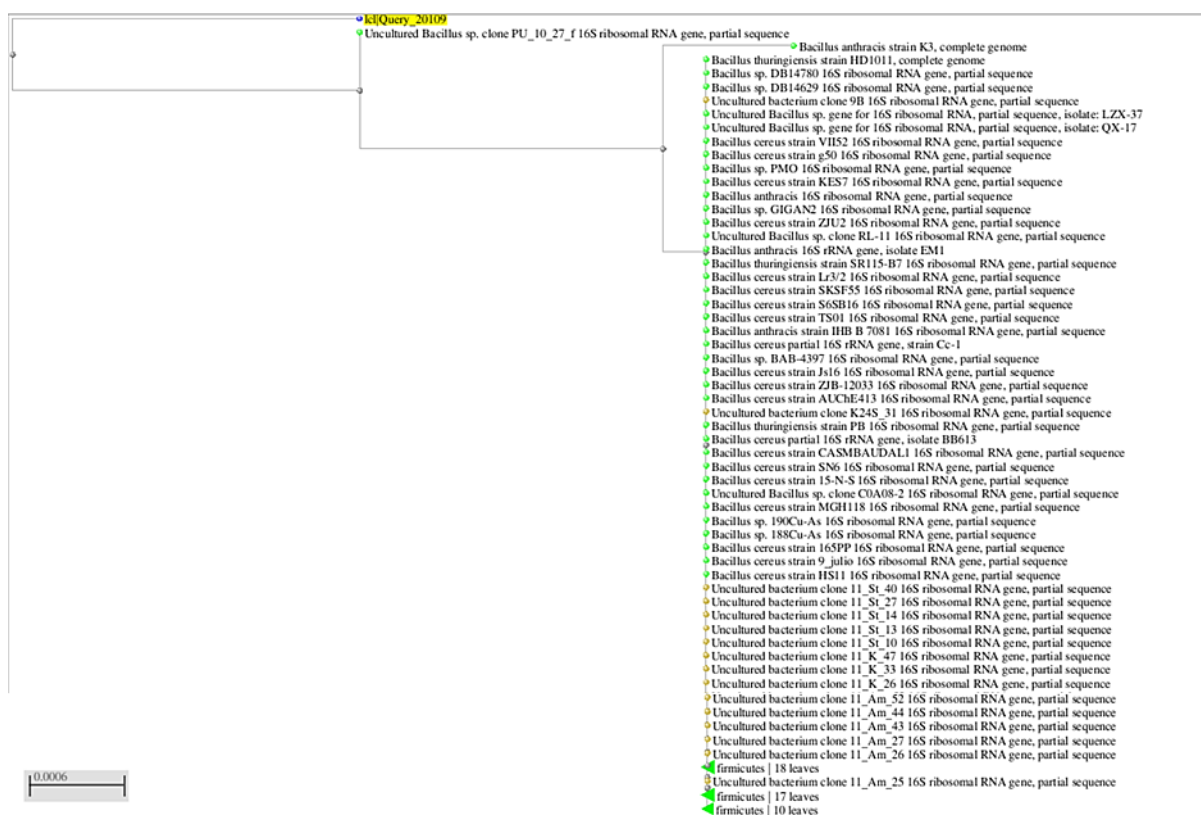
Bacillus cereus strain R5-339 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|JQ659737.1](#) Length: 1492 Number of Matches: 1

Range 1: 100 to 1025 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1631 bits(1808)	0.0	918/926(99%)	1/926(0%)	Plus/Plus
Query 1	ACGGGTGAGTAACACGTGGGTAACCTGCCCATAGACTGGGATAACTCCGGGAAACCGGG	60		
Sbjct 100	ACGGGTGAGTAACACGTGGGTAACCTGCCCATAGACTGGGATAACTCCGGGAAACCGGG	159		
Query 61	GCTAATACCGGATAACATTTTGAACCGCATGGTTCGAAATTGAAAGGCGGCTTCGGCTGT	120		
Sbjct 160	GCTAATACCGGATAACATTTTGAACCGCATGGTTCGAAATTGAAAGGCGGCTTCGGCTGT	219		
Query 121	CACTTATGGATGGACCCGCGTCGCATTAGCTAGTTGGTGAGGTAACGGCTCACCAAGGCA	180		
Sbjct 220	CACTTATGGATGGACCCGCGTCGCATTAGCTAGTTGGTGAGGTAACGGCTCACCAAGGCA	279		
Query 181	ACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGCCCAGA	240		
Sbjct 280	ACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGCCCAGA	339		
Query 241	CTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGGAGCAAC	300		
Sbjct 340	CTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGGAGCAAC	399		
Query 301	GCCGCGTGAGTGATGAAGGCTTTTCGGGTGTAAGGCTCTGTTGTTAGGGAAGAACAAGTG	360		
Sbjct 400	GCCGCGTGAGTGATGAAGGCTTTTCGGGTGTAAGGCTCTGTTGTTAGGGAAGAACAAGTG	459		
Query 361	CTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACGTG	420		
Sbjct 460	CTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACGTG	519		
Query 421	CCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCGTAAAGCG	480		
Sbjct 520	CCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCGTAAAGCG	579		
Query 481	CGCGCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGAGGGTCATT	540		
Sbjct 580	CGCGCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGAGGGTCATT	639		
Query 541	GGAAACTGGGAGACTTGAGTGCAGAAGAGGAAAGTGAATTCCATGTGTAGCGGTGAAAT	600		
Sbjct 640	GGAAACTGGGAGACTTGAGTGCAGAAGAGGAAAGTGAATTCCATGTGTAGCGGTGAAAT	699		
Query 601	GCGTAGAGATATGGAGGAACACAGTGGCGAAGGCGACTTTCTGGTCTGTAAGTACACT	660		
Sbjct 700	GCGTAGAGATATGGAGGAACACAGTGGCGAAGGCGACTTTCTGGTCTGTAAGTACACT	759		
Query 661	GAGGCGCGAAAGCGTGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAAC	720		
Sbjct 760	GAGGCGCGAAAGCGTGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAAC	819		
Query 721	GATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCA	780		
Sbjct 820	GATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCA	879		
Query 781	CTCCGCCCTGGGAGTACGGCCGCAAGGCTGAAACTCCAAGGAATTGACGGGGGCCCGCA	840		
Sbjct 880	CTCCGCCCTGGGAGTACGGCCGCAAGGCTGAAACTCCAAGGAATTGACGGGGGCCCGCA	939		
Query 841	CAAGCGGTGGAGCATGTGGTTTAATTCGAAGC-ACCCGAAAAACCTTACCAGGTCTTGAC	899		
Sbjct 940	CAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACCTTACCAGGTCTTGAC	999		
Query 900	TTCCTCTGACAACCCTAAAAATAGGG	925		
Sbjct 1000	ATCCTCTGAAAACCCTAGAGATAGGG	1025		



Staphylococcus haemolyticus KC139455.1

Staphylococcus haemolyticus strain B-20 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KC139455.1|](#) Length: 1428 Number of Matches: 1

Range 1: 508 to 1366 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1503 bits(1666)	0.0	852/862(99%)	3/862(0%)	Plus/Minus
Query 1	TGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTCACCGTAGCATGCTGATCTA	60		
Sbjct 1366	TGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTCACCGTAGCATGCTGATCTA	1307		
Query 61	CGATTACTAGCGATTCCAGCTTCATGTAGTCGAGTTGCAGACTACAATCCGAACCTGAGAA	120		
Sbjct 1306	CGATTACTAGCGATTCCAGCTTCATGTAGTCGAGTTGCAGACTACAATCCGAACCTGAGAA	1247		
Query 121	CAACTTTATGGGATTTGCTTGACCTCGCGGTTTCGCTACCCCTTGTATTGTCCATTGTAG	180		
Sbjct 1246	CAACTTTATGGGATTTGCTTGACCTCGCGGTTTCGCTACCCCTTGTATTGTCCATTGTAG	1187		
Query 181	CACGTGTGTAGCCCAAATCATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCC	240		
Sbjct 1186	CACGTGTGTAGCCCAAATCATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCC	1127		
Query 241	GGTTTGTACCGGCAGTCAACTTAGAGTGCCCAACTTAATGATGGCAACTAAGCTTAAGG	300		
Sbjct 1126	GGTTTGTACCGGCAGTCAACTTAGAGTGCCCAACTTAATGATGGCAACTAAGCTTAAGG	1067		
Query 301	GTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAACCATGC	360		
Sbjct 1066	GTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAACCATGC	1007		
Query 361	ACCACCTGTCACCTTTGTCCCCGAAGGGGAAAGCTCTATCTCTAGAGTTGTCAAAGGATG	420		
Sbjct 1006	ACCACCTGTCACCTTTGTCCCCGAAGGGGAAAGCTCTATCTCTAGAGTTGTCAAAGGATG	947		
Query 421	TCAAGATTGGTAAGGTTCTTCGCGTTGCTTCGAATTAACACCATGCTCCACCGCTTGT	480		
Sbjct 946	TCAAGATTGGTAAGGTTCTTCGCGTTGCTTCGAATTAACACCATGCTCCACCGCTTGT	887		
Query 481	GCGGGTCCCCGTCAATTCCTTTGAGTTTCANNCTTGCGNTCGTACTCCCCAGGCGGAGTG	540		
Sbjct 886	GCGGGTCCCCGTCAATTCCTTTGAGTTTCACCTTGCGGTCGTACTCCCCAGGCGGAGTG	827		
Query 541	CTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCATCGN	600		
Sbjct 826	CTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCATCGT	767		
Query 601	TTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGATCCCCACNCTTTTCGCACATCAG	660		
Sbjct 766	TTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGATCCCCACGCTTTTCGCACATCAG	707		
Query 661	CGTCAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCATATCTCTGCGCAT	720		
Sbjct 706	CGTCAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCATATCTCTGCGCAT	647		
Query 721	TTCACCGCTACACATGGAAATTCACCTTTCCTCTTCTGCACTCAAGTTTTCCAGTTTCCA	780		
Sbjct 646	TTCACCGCTACACATGG-AATTCACCTTTCCTCTTCTGCACTCAAGTTTTCCAGTTTCCA	588		
Query 781	ATGACCTCCACGGTTGAGCCGTGGGCTTTCCCATCCGACTTAAAAAACCGCCTACGCGC	840		
Sbjct 587	ATGACCTCCACGGTTGAGCCGTGGGCTTTCACATCAGACTTAAAAAACCGCCTACGCGC	528		
Query 841	GCCTTTACGCCCCAATAAATTCC	862		
Sbjct 527	G-CTTTACGCCCCAAT-AATTCC	508		



3- The phylogenetic analysis of bacteria isolated from computer Keyboards and computer mice samples

Bacillus subtilis KJ746466.1

Bacillus subtilis strain ACHSOC779 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ746466.1](#) Length: 1266 Number of Matches: 1

Range 1: 14 to 970 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1701 bits(1886)	0.0	952/957(99%)	1/957(0%)	Plus/Plus
Query 1	CTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGATA	60		
Sbjct 14	CTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGATA	73		
Query 61	ACTCCGGGAAACCGGGGCTAATACCGGATGGTTGTTTGAACCGCATGGTTCAAACATAAA	120		
Sbjct 74	ACTCCGGGAAACCGGGGCTAATACCGGATGGTTGTTTGAACCGCATGGTTCAAACATAAA	133		
Query 121	AGGTGGCTTGYGGCTACCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGTA	180		
Sbjct 134	AGGTGGCTTCGGCTACCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGTA	193		
Query 181	ACGGCTCACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGAC	240		
Sbjct 194	ACGGCTCACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGAC	253		
Query 241	TGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAA	300		
Sbjct 254	TGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAA	313		
Query 301	AGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTGT	360		
Sbjct 314	AGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTGT	373		
Query 361	TAGGGAAGAACAAGTACCGTTTCAATAGGGCGGTACCTTGACGGTACCTAACCAGAAAGC	420		
Sbjct 374	TAGGGAAGAACAAGTACCGTTTCAATAGGGCGGTACCTTGACGGTACCTAACCAGAAAGC	433		
Query 421	CACGGCTAACTACGTGCCAGCAGCCGCGTAATACGTAGGTGGCAAGCGTTGTCCGGAAT	480		
Sbjct 434	CACGGCTAACTACGTGCCAGCAGCCGCGTAATACGTAGGTGGCAAGCGTTGTCCGGAAT	493		
Query 481	TATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCA	540		
Sbjct 494	TATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCA	553		
Query 541	ACCGGGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCCA	600		
Sbjct 554	ACCGGGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCCA	613		
Query 601	CGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCTG	660		
Sbjct 614	CGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCTG	673		
Query 661	GTCTGTAAGTACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT	720		
Sbjct 674	GTCTGTAAGTACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT	733		
Query 721	AGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCCCTTAGTGCTGCA	780		
Sbjct 734	AGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCCCTTAGTGCTGCA	793		
Query 781	GCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAAT	840		
Sbjct 794	GCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAAT	853		
Query 841	TGAC--GGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTTCGAAGCAACGCGAAGAACC	899		
Sbjct 854	TGACGGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTTCGAAGCAACGCGAAGAACC	913		
Query 900	TTACCCGGTCTTGACATCCTCTGACATCCTAAAAATAGGACGTCCCCTTCGGGGGCA	956		
Sbjct 914	TTACCAGGTCTTGACATCCTCTGACATCCTAGAGATAGGACGTCCCCTTCGGGGGCA	970		



Bacillus amyloliquefaciens AB301004.1

Bacillus amyloliquefaciens gene for 16S rRNA, partial sequence, strain: GH3

Sequence ID: [dbj|AB301004.1](#) Length: 1481 Number of Matches: 1

Range 1: 447 to 1408 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1709 bits(1894)	0.0	957/962(99%)	1/962(0%)	Plus/Minus	
Query 1	GTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACC	60			
Sbjct 1408	GTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACC	1349			
Query 61	GCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTG	120			
Sbjct 1348	GCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTG	1289			
Query 121	CGATCCGAACAGATTGTTGGGATTGGCTTAACCTCGCGGTTTCGCTGCCCTTT	180			
Sbjct 1288	CGATCCGAACAGATTGTTGGGATTGGCTTAACCTCGCGGTTTCGCTGCCCTTT	1229			
Query 181	GTTCTGTCCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTC	240			
Sbjct 1228	GTTCTGTCCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTC	1169			
Query 241	ATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTG	300			
Sbjct 1168	ATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTG	1109			
Query 301	GCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAG	360			
Sbjct 1108	GCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAG	1049			
Query 361	CTGACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGACGTCCTATCTCTAGG	420			
Sbjct 1048	CTGACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGACGTCCTATCTCTAGG	989			
Query 421	ATTGTGAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACAT	480			
Sbjct 988	ATTGTGAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACAT	929			
Query 481	GCTCCACCGCTTGTGCGGGCCCCGTCAATTCCCTTGAGTTTCAGTCTTGCGACCGTACT	540			
Sbjct 928	GCTCCACCGCTTGTGCGGGCCCCGTCAATTCCCTTGAGTTTCAGTCTTGCGACCGTACT	869			
Query 541	CCCCAGGCGGAGTGCTTAATGCGTTAGTGCAGCCTAAGGGGCGGAAACCCCTAACAC	600			
Sbjct 868	CCCCAGGCGGAGTGCTTAATGCGTTAGTGCAGCCTAAGGGGCGGAAACCCCTAACAC	809			
Query 601	TTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACG	660			
Sbjct 808	TTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACG	749			
Query 661	CTTTCGCTCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCC	720			
Sbjct 748	CTTTCGCTCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCC	689			
Query 721	ACATCTCTACGCATTTACCGCTACACGTGGAATTCACCTCTCTCTTCTGCACTCAAGT	780			
Sbjct 688	ACATCTCTACGCATTTACCGCTACACGTGGAATTCACCTCTCTCTTCTGCACTCAAGT	629			
Query 781	TCCCCAGTTTCCAATGACCCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTAAAAA	840			
Sbjct 628	TCCCCAGTTTCCAATGACCCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAGACTTAAGAA	569			
Query 841	CCGCCTGCGAGCCCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACC	900			
Sbjct 568	CCGCCTGCGAGCCCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACC	509			
Query 901	GCGGCTGCTGGCACGTAGTTAGCCG-GGCTTTCTGGTTAGGTACCGTCAAGGGGCCCCC	959			
Sbjct 508	GCGGCTGCTGGCACGTAGTTAGCCGTTTCTGGTTAGGTACCGTCAAGGTGCCGCC	449			
Query 960	TA 961				
Sbjct 448	TA 447				

icl|Query_27917

firmicutes | 2 leaves

- ◆ Bacillus amyloliquefaciens strain GR53 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. H405 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain T45 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus methylotrophicus strain Z7 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain D12 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain D48 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus subtilis strain MNNIT SR18 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium DS-1 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain SDS14 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain August M2 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. LG 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium OKR 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain IHB B 7126 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain MSEB 18 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus methylotrophicus strain HB24 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus vallismortis strain JBS-10 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. APG-1 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain KAVK1 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. 825 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium YC-LKJ56 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus methylotrophicus strain WY-3 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus methylotrophicus strain CYJ 4 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. YP1(2015) 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain Jxnuwy-1 CCTCC M 2014639 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus subtilis strain J-18 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens subsp. plantarum strain Pmg-32 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus vallismortis strain NBIF-001 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain XY-1 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium YC-LKJ9 16S ribosomal RNA gene, partial sequence

firmicutes | 21 leaves

- ◆ Bacillus sp. YP5 16S ribosomal RNA gene, partial sequence
- ◆ Uncultured Bacillus sp. clone DDGJ05 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain XS-2 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium YC-LKJ60 16S ribosomal RNA gene, partial sequence

firmicutes | 23 leaves

- ◆ Bacillus amyloliquefaciens subsp. plantarum strain I-1-b-5 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. DB14353 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain WJ-2 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. PPM3 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium YC-LKJ2 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. HU-2012 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain TD-7 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain Lys-1436 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus subtilis strain ND04 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus methylotrophicus strain HB26 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus methylotrophicus strain HB11 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens subsp. plantarum strain IHB B 12506 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus subtilis strain IHB B 7050 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus sp. BAB-650 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus vallismortis strain CM1E3 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain NLB12 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus subtilis strain YA-3 16S ribosomal RNA gene, partial sequence
- ◆ Bacterium NJ2 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain 1 16S ribosomal RNA gene, partial sequence
- ◆ Bacillus amyloliquefaciens strain Jxnuwx-1 CCTCC M 2014638 16S ribosomal RNA gene, partial sequence

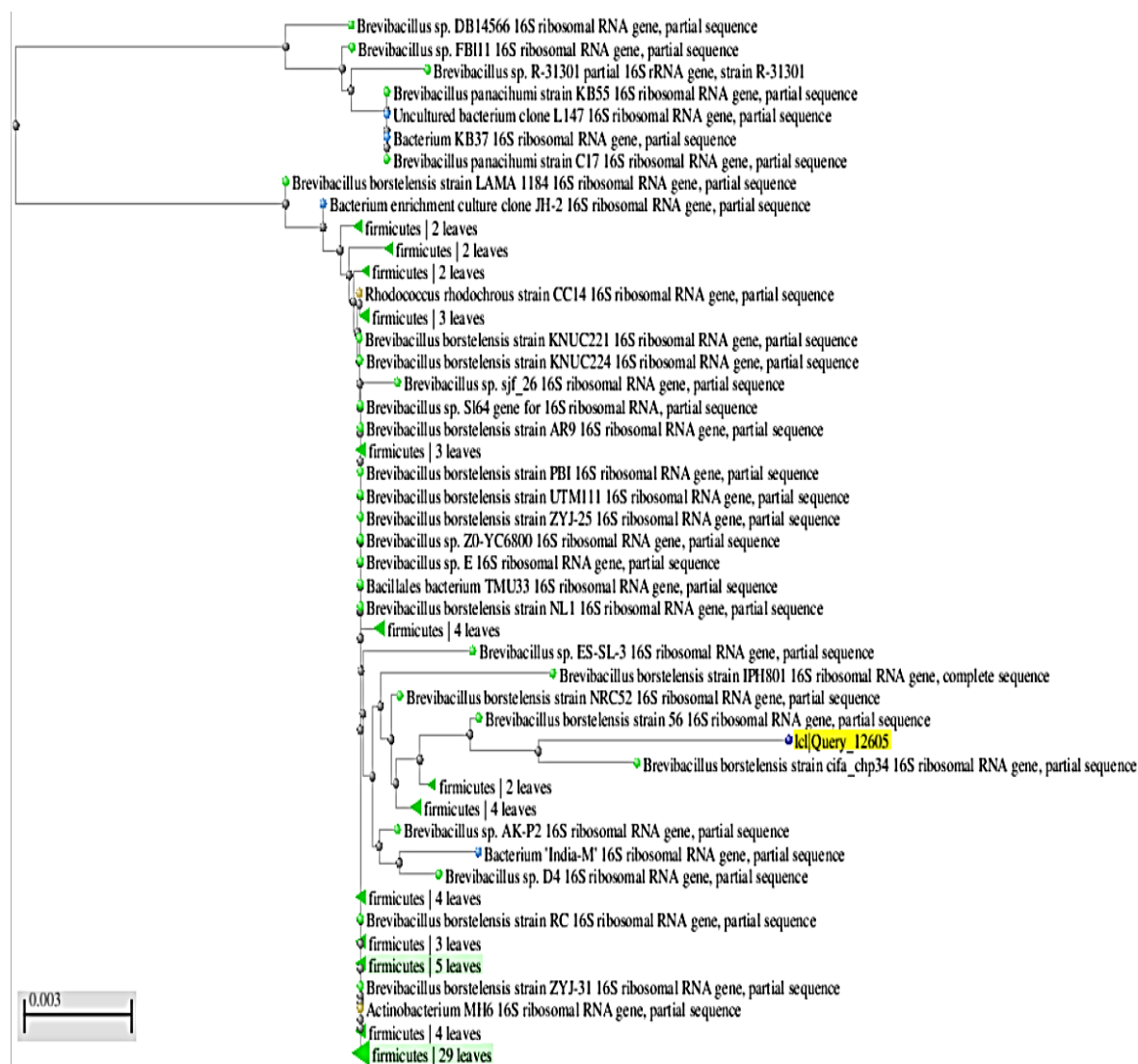
0.0004

Brevibacillus brostelensis EU816699.1

Brevibacillus borstelensis clone US12 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|EU816699.1|](#) Length: 1529 Number of Matches: 1

Range 1: 93 to 1053		GenBank	Graphics			Next Match	Previous Match
Score	Expect	Identities		Gaps	Strand		
1656 bits(1836)	0.0	946/962(98%)		2/962(0%)	Plus/Plus		
Query 1		GTACACGTTAGGCAACCTGCCCCGTAAGCTCGGGATAACATGGGGAAACTCATGCTAATAC					60
Sbjct 93		GTACACGTTAGGCAACCTGCCCCGTAAGCTCGGGATAACATGGGGAAACTCATGCTAATAC					152
Query 61		CGGATAGGGTCTTCTCTCGCATGAGAGGAGACGGAAGGTGGCGCAAGCTACCACTTACG					120
Sbjct 153		CGGATAGGGTCTTCTCTCGCATGAGAGGAGACGGAAGGTGGCGCAAGCTACCACTTACG					212
Query 121		GATGGGCCTGCGGCGCATTAGCTAGTTGGTGGGGTAACGGCCTACCAAGGCGACGATGCG					180
Sbjct 213		GATGGGCCTGCGGCGCATTAGCTAGTTGGTGGGGTAACGGCCTACCAAGGCGACGATGCG					272
Query 181		TAGCCGACCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACG					240
Sbjct 273		TAGCCGACCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCAGACTCCTACG					332
Query 241		GGAGGCAGCAGTAGGGAATTTCCACAATGGACGAAAGTCTGATGGAGCAACGCCGCGTG					300
Sbjct 333		GGAGGCAGCAGTAGGGAATTTCCACAATGGACGAAAGTCTGATGGAGCAACGCCGCGTG					392
Query 301		AACGATGAAGGTCTTCGGATTGTAAAGTTCTGTTGTCAGAGACGAACAAGTACCGTTTCA					360
Sbjct 393		AACGATGAAGGTCTTCGGATTGTAAAGTTCTGTTGTCAGAGACGAACAAGTACCGTTTCA					452
Query 361		ACAGGGCGGTACCTTGACGGTACCTGACGAGAAAGCCACGGCTAACTACGTGCCAGCAGC					420
Sbjct 453		ACAGGGCGGTACCTTGACGGTACCTGACGAGAAAGCCACGGCTAACTACGTGCCAGCAGC					512
Query 421		CGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCGCAGG					480
Sbjct 513		CGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCGCAGG					572
Query 481		CGGCTATGTAAGTCTGGTGTAAAGCCCGGGGCTCAACCCCGGTTTCGCATCGGAAACTGT					540
Sbjct 573		CGGCTATGTAAGTCTGGTGTAAAGCCCGGGGCTCAACCCCGGTTTCGCATCGGAAACTGT					632
Query 541		GTAGCTTGAGTGCAAGAGGAAAGCGGTATTCCACGTGTAGCGGTGAAATGCGTAGAGA					600
Sbjct 633		GTAGCTTGAGTGCAAGAGGAAAGCGGTATTCCACGTGTAGCGGTGAAATGCGTAGAGA					692
Query 601		TGTGGAGGAACACCAAGTGGCGAAGGCGGCTTTCTGGTCTGTAAGTACGCTGAGGCGCGA					660
Sbjct 693		TGTGGAGGAACACCAAGTGGCGAAGGCGGCTTTCTGGTCTGTAAGTACGCTGAGGCGCGA					752
Query 661		AAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGC					720
Sbjct 753		AAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGC					812
Query 721		TAGGTGGTggtgggggTTTCAATACCCTCAGTGCCGCGAGCTAACGCAATAAGCACTCCGCC					780
Sbjct 813		TAGGTGGTGGGGGGTTTCAATACCCTCAGTGCCGCGAGCTAACGCAATAAGCACTCCGCC					872
Query 781		TGGGGAGTACGCTCGCAAGAGTGAAACTCAAAGGAATTGACGGGGGCCCGCACAAAGCGGG					840
Sbjct 873		TGGGGAGTACGCTCGCAAGAGTGAAACTCAAAGGAATTGACGGGGGCCCGCACAAAGCGGT					932
Query 841		GGAGCATGGGGTTTAAATTCGAAGCAACGCGAAGAACCTTACCA-GTCTTGACATCCCGCT					899
Sbjct 933		GGAGCATGTGGTTTAAATTCGAAGCAACGCGAAGAACCTTACCAGGTCTTGACATCCCGCT					992
Query 900		GACCGTCTTAAATAAGGGCTTCCCTTCGGGGCAGCGGGGAAGggggggggggATGGTGGT					959
Sbjct 993		GACCGTCTTAGAGATAGGGCTTCCCTTCGGGGCAGCGGTG-ACAGGTGGTGCATGGTTGT					1051
Query 960		CG	961				
Sbjct 1052		CG	1053				



Staphylococcus epidermidis KF575163.1

Staphylococcus epidermidis strain G0242 16S ribosomal RNA gene, partial sequence
Sequence ID: gb|KF575163.1| Length: 1434 Number of Matches: 1

Range 1: 44 to 1016		GenBank	Graphics			▼ Next Match ▲ Previous Match
Score	Expect	Identities		Gaps	Strand	
1718 bits(1904)	0.0	967/973(99%)		3/973(0%)	Plus/Plus	
Query	1	TTGACGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTACCTATAAGACTGGGATA				60
Sbjct	44	TTGACGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTACCTATAAGACTGGGATA				103
Query	61	ACTTCGGGAAACCGGAGCTAATACCGGATAATATTTTGAACCGCATGGTTCGATAGTGAA				120
Sbjct	104	ACTTCGGGAAACCGGAGCTAATACCGGATAATATTTTGAACCGCATGGTTCGATAGTGAA				163
Query	121	AGATGGTTTTGCTATCACTTATAGATGGACCCGCGCGTATTAGCTAGTTGGTAAGGTAA				180
Sbjct	164	AGATGGTTTTGCTATCACTTATAGATGGACCCGCGCGTATTAGCTAGTTGGTAAGGTAA				223
Query	181	CGGCTTACCAAGGCGACGATACGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGAAC				240
Sbjct	224	CGGCTTACCAAGGCGACGATACGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGAAC				283
Query	241	GAGACACGGTCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGGCGAAA				300
Sbjct	284	GAGACACGGTCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGGCGAAA				343
Query	301	GCCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTCTTCGGATCGTAAACTCTGTTATT				360
Sbjct	344	GCCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTCTTCGGATCGTAAACTCTGTTATT				403
Query	361	AGGGAAGAACATACGTGTAAGTAACATGCACGTCTTGACGGTACCTAATCAGAAAGCCA				420
Sbjct	404	AGGGAAGAACATACGTGTAAGTAACATGCACGTCTTGACGGTACCTAATCAGAAAGCCA				463
Query	421	CGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTAT-CCGGAATT				479
Sbjct	464	CGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCCGGAATT				523
Query	480	ATTGGGCGTAAAGCGCGCGTAGGCGGTTTTTTAAGTCTGATGTGAAAGCCACGGCTCAA				539
Sbjct	524	ATTGGGCGTAAAGCGCGCGTAGGCGGTTTTTTAAGTCTGATGTGAAAGCCACGGCTCAA				583
Query	540	CCGTGGAGGGTCATTGGAAACTGAAAACTTGAGTGCAGAAGAGGAAAGTGGAAATCCAT				599
Sbjct	584	CCGTGGAGGGTCATTGGAAACTGAAAACTTGAGTGCAGAAGAGGAAAGTGGAAATCCAT				643
Query	600	GTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAAGTGGCGAAGGCGACTTTCTGG				659
Sbjct	644	GTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAAGTGGCGAAGGCGACTTTCTGG				703
Query	660	TCTGTAACTGACGCTGATGTGCGAAAGCGTGGGGATCAAACAGGATTAGATACCTGGTA				719
Sbjct	704	TCTGTAACTGACGCTGATGTGCGAAAGCGTGGGGATCAAACAGGATTAGATACCTGGTA				763
Query	720	GTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCTTAGTGCTGCAG				779
Sbjct	764	GTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCTTAGTGCTGCAG				823
Query	780	CTAACGCATTAAGCACTCCGCCGTGGGGAGTACGACCGCAAGGTTGAAACTCAAAGGAATT				839
Sbjct	824	CTAACGCATTAAGCACTCCGCCGTGGGGAGTACGACCGCAAGGTTGAAACTCAAAGGAATT				883
Query	840	GACGGGGACCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGC-ACGCGAAAAACCT				898
Sbjct	884	GACGGGGACCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACCT				943
Query	899	TACCAAATCTTGACATCCTTTGACAACTCTAAAGATAGAGCCTTCCCCTTTTCGGGGGA-A				957
Sbjct	944	TACCAAATCTTGACATCCTTTGACAACTCTAGAGATAGAGCCTTCCCCTTTTCGGGGGACA				1003
Query	958	AAGTGACAGGGG 970				
Sbjct	1004	AAGTGACAGGTG 1016				

kl[Query_21873]

Uncultured bacterium clone nck172601c1 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain G0242 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain Sh29312/L2, complete genome
Staphylococcus sp. BQN2P-01d 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbu277a09c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbu283c12c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbu283g02c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw830c11c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw831h11c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1189c10c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1095d07c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1105d02c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1106f08c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1106h08c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1107b10c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1106d4c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1065d02c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1189f06c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1191b10c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nbw1190f12c1 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain LCR51 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain EG12 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain FR1_68 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncd558b09c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncd577e07c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncd586b11c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncd583b02c1 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. YSL09-4 gene for 16S rRNA, partial sequence
Uncultured bacterium clone ncd924b12c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncd1271c06c1 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. HL-13 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain CIFRI D-TSB2 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain CIFRI P-TSB4 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. CIFRI CH-TSB23 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. CIFRI H-TSB-15-ZMA 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. CIFRI H-TSB-20-ZMA 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. CIFRI H-TSB-6-1A 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain HNMCTRI 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain BP3_2B 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain LE12_2A 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. EB38 16S ribosomal RNA gene, partial sequence
Uncultured Staphylococcus sp. clone T3-3 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. ARB1 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain 20.1 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain 21.1 16S ribosomal RNA gene, partial sequence
Acinetobacter radoresiens strain G16 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain G15 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. RH-29 gene for 16S rRNA, partial sequence
Staphylococcus sp. M17 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus JCS1435 strain KSC1435 16S ribosomal RNA, complete sequence
Staphylococcus haemolyticus strain M2 16S ribosomal RNA gene, partial sequence
Bacterium CulaenoE10F 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain S001N 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain S006N 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain S009b 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain S011b 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain VBW023 16S ribosomal RNA gene, partial sequence
Bacterium KR 1996/1_182 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain L38 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain BP/SU2 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. H-179 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain SH6 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain SH8 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain NIOT-sb3 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain C0181 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. G0271 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain PAH-3 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone g3b23 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone g3b26 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone g3b27 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone g3b33 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone g3b39 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain XB20 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain MJMG7.10 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain KJ1-5-97 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain KJ1599 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck175a12c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck170b04c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck171d06c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncm50d05c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone ncm50g10c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck169d07c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck171g06c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck175c09c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck177d11c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck188a05c1 16S ribosomal RNA gene, partial sequence
Uncultured bacterium clone nck310d02c1 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain DG-4 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain CIFRI P-TSB-72 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain J12 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. 09-A3 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. 09-M3 16S ribosomal RNA gene, partial sequence
Staphylococcus haemolyticus strain DSOAG14 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. Ag08 16S ribosomal RNA gene, partial sequence
Bacterium TRO1 16S ribosomal RNA gene, partial sequence
firmicutes | 4 leaves
Staphylococcus haemolyticus strain DSOAG11 16S ribosomal RNA gene, partial sequence

Pantoea caldia AB907785.1

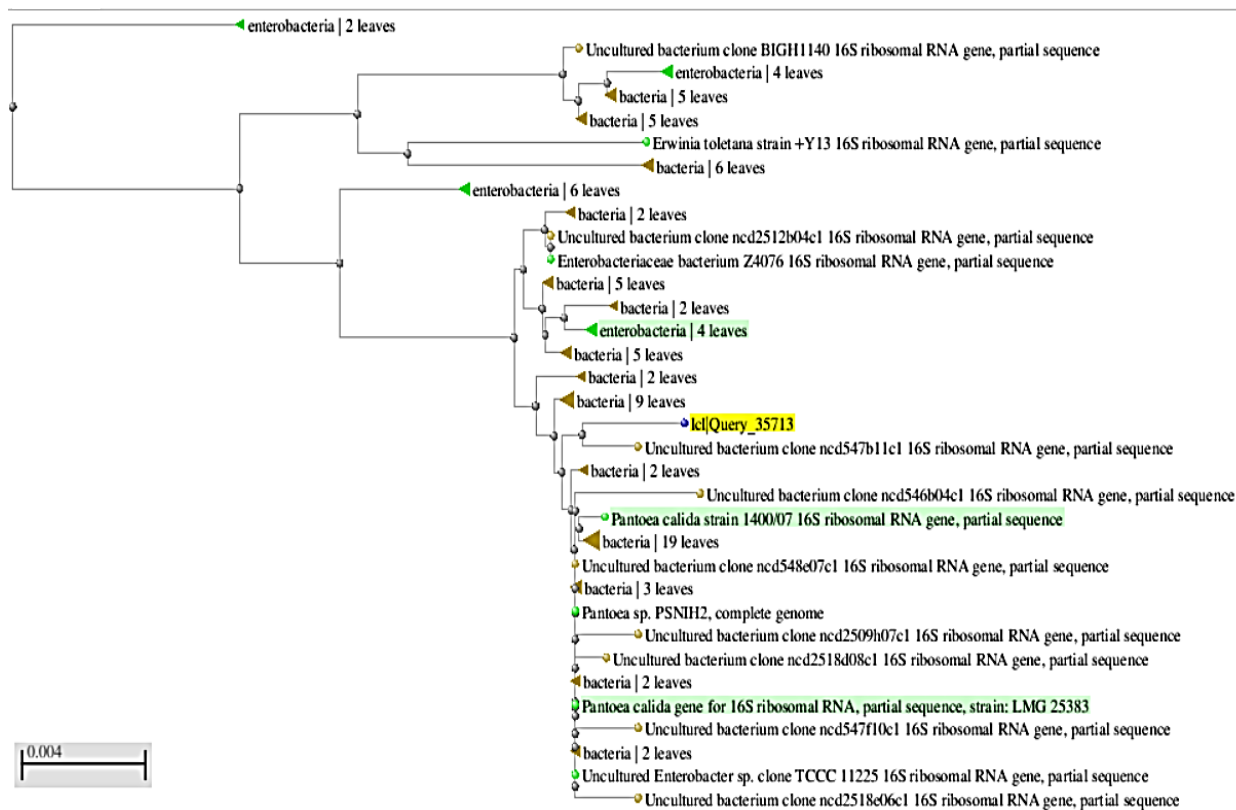
Pantoea caldia gene for 16S ribosomal RNA, partial sequence, strain: LMG 25383

Sequence ID: [dbj|AB907785.1](#) Length: 1495 Number of Matches: 1

Range 1: 51 to 1032 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1745 bits(1934)	0.0	977/982(99%)	1/982(0%)	Plus/Plus
Query 1	AGCAGCTTGCTGCTTCGCTGACGAGTGGCGGACGGGTGAGTAATGTCTGGGGATCTGCCC	60		
Sbjct 51	AGCAGCTTGCTGCTTCGCTGACGAGTGGCGGACGGGTGAGTAATGTCTGGGGATCTGCCC	110		
Query 61	GATGGAGGGGGATAAACCCTGGAACGGTGGCTAATACCGCATAACGTCGCAAGACCAA	120		
Sbjct 111	GATGGAGGGGGATAAACCCTGGAACGGTGGCTAATACCGCATAACGTCGCAAGACCAA	170		
Query 121	GTGGGGGACCTTCGGGCCCTCACACCATCGGATGAACCCAGATGGGATTAGCTAGTAGGTG	180		
Sbjct 171	GTGGGGGACCTTCGGGCCCTCACACCATCGGATGAACCCAGATGGGATTAGCTAGTAGGTG	230		
Query 181	GGGTAACGGCTCACCTAGGCGACGATCCCTAGCTGGTCTGAGAGGATGACCAGCCACACT	240		
Sbjct 231	GGGTAACGGCTCACCTAGGCGACGATCCCTAGCTGGTCTGAGAGGATGACCAGCCACACT	290		
Query 241	GGAAGTGAAGACAGGTCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACAATGG	300		
Sbjct 291	GGAAGTGAAGACAGGTCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACAATGG	350		
Query 301	GCGCAAGCCTGATGCAGCCATGCCGCGTGTATGAAGAAGGCCTTCGGGTTGTAAAGTACT	360		
Sbjct 351	GCGCAAGCCTGATGCAGCCATGCCGCGTGTATGAAGAAGGCCTTCGGGTTGTAAAGTACT	410		
Query 361	TTCAGCGGGGAGGAAGGGATGGTGTCTTAATACGCGCCGTCATTGACGTTACCCGCAGAAG	420		
Sbjct 411	TTCAGCGGGGAGGAAGGGATGGTGTCTTAATACGCGCCGTCATTGACGTTACCCGCAGAAG	470		
Query 421	AAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAATCG	480		
Sbjct 471	AAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAATCG	530		
Query 481	GAATTACTGGGCGTAAAGCGCACGCAGGCGGTCTGTAAAGTCAGATGTGAAATCCCCGGG	540		
Sbjct 531	GAATTACTGGGCGTAAAGCGCACGCAGGCGGTCTGTAAAGTCAGATGTGAAATCCCCGGG	590		
Query 541	CTTAACCTGGGAAGTGCATTTGAAACTGGCAGGCTTGAGTCTCGTAGAGGGGGGTAGAAT	600		
Sbjct 591	CTTAACCTGGGAAGTGCATTTGAAACTGGCAGGCTTGAGTCTCGTAGAGGGGGGTAGAAT	650		
Query 601	TCCAGGTGTAGCGGTGAAATGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCGGCC	660		
Sbjct 651	TCCAGGTGTAGCGGTGAAATGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCGGCC	710		
Query 661	CCTGGACGAAGACTGACGCTCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCC	720		
Sbjct 711	CCTGGACGAAGACTGACGCTCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCC	770		
Query 721	TGGTAGTCCACGCCGTAAACGATGTCGACTTGGAGGCTGTTTCCTTGAGAAGTGGCTTCC	780		
Sbjct 771	TGGTAGTCCACGCCGTAAACGATGTCGACTTGGAGGCTGTTTCCTTGAGAAGTGGCTTCC	830		
Query 781	GGAGCTAACGCGTTAAGTCGACCGCCTGGGGAGTACGGCCGCAAGGTTAAAACCTCAAATG	840		
Sbjct 831	GGAGCTAACGCGTTAAGTCGACCGCCTGGGGAGTACGGCCGCAAGGTTAAAACCTCAAATG	890		
Query 841	AATTGACGGGGGGCCGCACAAGCGGTGGAGCATGTGGTTTAAATTCGATGCAACGCAAGA	900		
Sbjct 891	AATTGACGGGGGGCCGCACAAGCGGTGGAGCATGTGGTTTAAATTCGATGCAACGCAAGA	950		
Query 901	ACCTTACCTGGTCTTGACATCCACGGAATTCGGCAAAAATGCCCTAGTGCCTTCGGGAAC	960		
Sbjct 951	ACCTTACCTGGTCTTGACATCCACGGAATTCGGCAGAGATGCCCTAGTGCCTTCGGGAAC	1010		
Query 961	CGTGAAAAAGG-GCTGCATGGC	981		
Sbjct 1011	CGTGAGACAGGTGCTGCATGGC	1032		



Bacillus cereus DQ923480.1

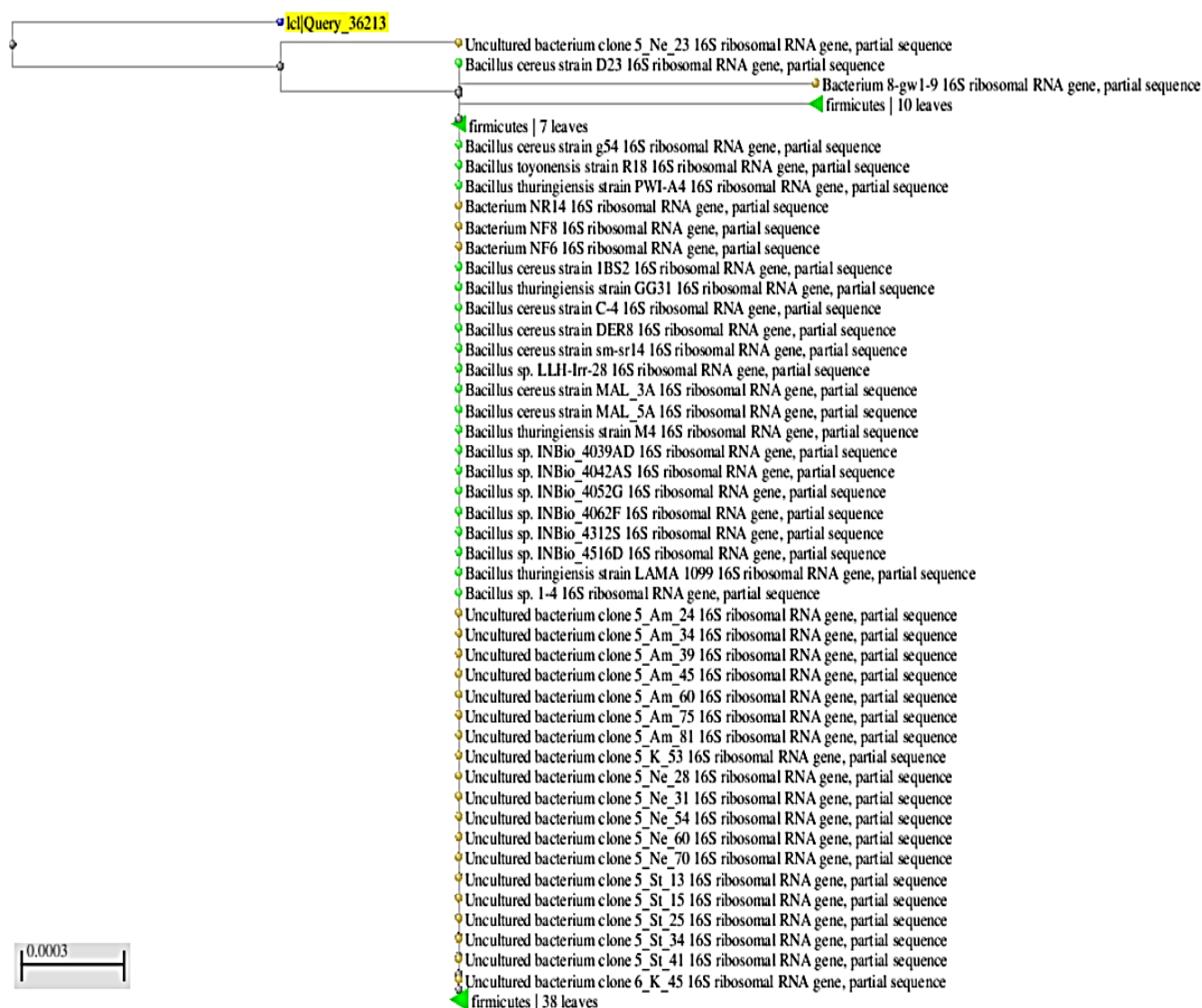
Bacillus cereus strain D23 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|DQ923480.1|](#) Length: 1429 Number of Matches: 1

Range 1: 430 to 1390 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1725 bits(1912)	0.0	959/961(99%)	0/961(0%)	Plus/Minus	
Query 1	TGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCG	60			
Sbjct 1390	TGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCG	1331			
Query 61	CGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTAC	120			
Sbjct 1330	CGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTAC	1271			
Query 121	AATCCGAACTGAGAACGGTTTTATGAGATTAGCTCCACCTCGCGGTCTTGACGCTCTTTG	180			
Sbjct 1270	AATCCGAACTGAGAACGGTTTTATGAGATTAGCTCCACCTCGCGGTCTTGACGCTCTTTG	1211			
Query 181	TACCGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCA	240			
Sbjct 1210	TACCGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCA	1151			
Query 241	TCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGG	300			
Sbjct 1150	TCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGG	1091			
Query 301	CAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCAGCACACGAGC	360			
Sbjct 1090	CAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCAGCACACGAGC	1031			
Query 361	TGACGACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCTAGGG	420			
Sbjct 1030	TGACGACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCTAGGG	971			
Query 421	TTTTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAACCACATG	480			
Sbjct 970	TTTTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAACCACATG	911			
Query 481	CTCCACCGCTTGTGCGGGCCCCGTCGAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTC	540			
Sbjct 910	CTCCACCGCTTGTGCGGGCCCCGTCGAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTC	851			
Query 541	CCCAGGCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCTCTAACACT	600			
Sbjct 850	CCCAGGCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCTCTAACACT	791			
Query 601	TAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGC	660			
Sbjct 790	TAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGC	731			
Query 661	TTTCGCGCCTCAGTGTGAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCA	720			
Sbjct 730	TTTCGCGCCTCAGTGTGAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCA	671			
Query 721	TATCTCTACGCATTTACCGCTACACATGGAATTCACCTTTCCTCTTCTGCACTCAAGTC	780			
Sbjct 670	TATCTCTACGCATTTACCGCTACACATGGAATTCACCTTTCCTCTTCTGCACTCAAGTC	611			
Query 781	TCCCAGTTTCCAATGACCCTCCACGGTTGAGCCGTGGGCTTTCACATCAAACCTTAAGAAA	840			
Sbjct 610	TCCCAGTTTCCAATGACCCTCCACGGTTGAGCCGTGGGCTTTCACATCAGACTTAAGAAA	551			
Query 841	CCACCTGCGCGCGCTTTACGCCCAATAATTTCCGGATAACGCTTGCCACCTACGTATTAC	900			
Sbjct 550	CCACCTGCGCGCGCTTTACGCCCAATAATTTCCGGATAACGCTTGCCACCTACGTATTAC	491			
Query 901	CGCGGCTGCTGGCACGTAGTTAGCCGTGGCTTCTGGTTAGGTACCGTCAAGGGGCCAGC	960			
Sbjct 490	CGCGGCTGCTGGCACGTAGTTAGCCGTGGCTTCTGGTTAGGTACCGTCAAGGTGCCAGC	431			
Query 961	T 961				
Sbjct 430	T 430				



Pseudomonas luteola KC429633.1

Pseudomonas luteola strain XFB-BV 16S ribosomal RNA gene, partial sequence
Sequence ID: [gb|KC429633.1](#) Length: 1388 Number of Matches: 1

Range 1: 42 to 991		GenBank	Graphics			▼ Next Match	▲ Previous Match
Score	Expect	Identities	Gaps	Strand			
1669 bits(1850)	0.0	940/950(99%)	0/950(0%)	Plus/Plus			
Query	1	GGCGGACGGGTGAGTAATGCCTAGGAATCTGCCTGGTAGTGGGGGACAACGTTTCGAAAAG					60
Sbjct	42	GGCGGACGGGTGAGTAATGCCTAGGAATCTGCCTGGTAGTGGGGGACAACGTTTCGAAAAG					101
Query	61	GAACGCTAATACCGCATACGTCCTACGGGAGAAAGTGGGGGATCTTCGGACCTCACGCTA					120
Sbjct	102	GAACGCTAATACCGCATACGTCCTACGGGAGAAAGTGGGGGATCTTCGGACCTCACGCTA					161
Query	121	TCAGATGAGCCTAGGTCGGATTAGCTAGTTGGTGGGGTAACGGCTCACCAAGGCGACGAT					180
Sbjct	162	TCAGATGAGCCTAGGTCGGATTAGCTAGTTGGTGGGGTAACGGCTCACCAAGGCGACGAT					221
Query	181	CCGTAACCTGGTCTGAGAGGATGATCAGTCACACTGGAAGTGGAGACACGGTCCAGACTCCT					240
Sbjct	222	CCGTAACCTGGTCTGAGAGGATGATCAGTCACACTGGAAGTGGAGACACGGTCCAGACTCCT					281
Query	241	ACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCCATGCCGC					300
Sbjct	282	ACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCCATGCCGC					341
Query	301	GTGTGTGAAGAAGGCCCTCGGGTCGTAAAGCACTTTAAGCTGGGAGGAAGGGTTGTAACC					360
Sbjct	342	GTGTGTGAAGAAGGCCCTCGGGTCGTAAAGCACTTTAAGCTGGGAGGAAGGGTTGTAACC					401
Query	361	TAATACGTTGCAGCTTTGACGTTACCAGCAGAATAAGCACCGGCTAACTCTGTGCCAGCA					420
Sbjct	402	TAATACGTTGCAGCTTTGACGTTACCAGCAGAATAAGCACCGGCTAACTCTGTGCCAGCA					461
Query	421	GCCGCGGTAATACAGAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGCGCGCGTA					480
Sbjct	462	GCCGCGGTAATACAGAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGCGCGCGTA					521
Query	481	GGTGGCTTGGTAAGTTGAATGTGAAATCCCCGGGCTCAACCTGGGAAGTGCATCCAAAAC					540
Sbjct	522	GGTGGCTTGGTAAGTTGAATGTGAAATCCCCGGGCTCAACCTGGGAAGTGCATCCAAAAC					581
Query	541	TGCCTGGCTAGAGTACGGTAGAGGGTGGTGGAAATTCCTGTGTAGCGGTGAAATGCGTAG					600
Sbjct	582	TGCCTGGCTAGAGTACGGTAGAGGGTGGTGGAAATTCCTGTGTAGCGGTGAAATGCGTAG					641
Query	601	ATATAGGAAGGAACACCAAGTGGCGAAGGCGACCACCTGGACTGATACTGACACTGAGGTG					660
Sbjct	642	ATATAGGAAGGAACACCAAGTGGCGAAGGCGACCACCTGGACTGATACTGACACTGAGGTG					701
Query	661	CGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGTC					720
Sbjct	702	CGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGTC					761
Query	721	AACTAGCCGTTGGGGTCCTTGAGACTTTAGTGGCGCAGCTAACGCAATAAGTTGACCGCC					780
Sbjct	762	AACTAGCCGTTGGGGTCCTTGAGACTTTAGTGGCGCAGCTAACGCAATAAGTTGACCGCC					821
Query	781	TGGGGAGTACGGCCGCAAGGTTAAAACTCAAATGAATTGACGGGGGCCGCAAGCGGT					840
Sbjct	822	TGGGGAGTACGGCCGCAAGGTTAAAACTCAAATGAATTGACGGGGGCCGCAAGCGGT					881
Query	841	GGAGCATGTGGTTTATTTCAAGCAACGCGAAAAACCTTACCAGGCCTTGACATGCAAAA					900
Sbjct	882	GGAGCATGTGGTTTATTTCAAGCAACGCGAAAAACCTTACCAGGCCTTGACATGCAAG					941
Query	901	AACTTTCCAAAAAAGGATTGGTGCCTTCGGGAACTCTGAACCAGGGGCTG					950
Sbjct	942	AACTTTCCAGAGATGGATTGGTGCCTTCGGGAACTCTGACACAGGTGCTG					991



4- The phylogenetic analysis of bacteria isolated from mobile phones samples

Staphylococcus epidermidis KJ398217.1

Staphylococcus epidermidis strain LH-Y4 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ398217.1](#) Length: 1413 Number of Matches: 1

Range 1: 590 to 1369 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1398 bits(1550)	0.0	778/780(99%)	0/780(0%)	Plus/Minus
Query 3	CTCTCGTGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTACCGTAGCATGCT	62		
Sbjct 1369	CTCTCGTGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTACCGTAGCATGCT	1310		
Query 63	GATCTACGATTACTAGCGATTCCAGCTTCATATAGTCGAGTTGCAGACTACAATCCGAAC	122		
Sbjct 1309	GATCTACGATTACTAGCGATTCCAGCTTCATATAGTCGAGTTGCAGACTACAATCCGAAC	1250		
Query 123	TGAGAACAACCTTTATGGGATTTGCTTGACCTCGCGGTTTCGCTACCCTTTGTATTGTCCA	182		
Sbjct 1249	TGAGAACAACCTTTATGGGATTTGCTTGACCTCGCGGTTTCGCTGCCCTTTGTATTGTCCA	1190		
Query 183	TTGTAGCACGTGTGTAGCCCAAATCATAAGGGGCATGATGATTTGACGTCATCCCCACCT	242		
Sbjct 1189	TTGTAGCACGTGTGTAGCCCAAATCATAAGGGGCATGATGATTTGACGTCATCCCCACCT	1130		
Query 243	TCCTCCGGTTTGTACCGGCAGTCAACTTAGAGTGCCCAACTTAATGATGGCAACTAAGC	302		
Sbjct 1129	TCCTCCGGTTTGTACCGGCAGTCAACTTAGAGTGCCCAACTTAATGATGGCAACTAAGC	1070		
Query 303	TTAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAA	362		
Sbjct 1069	TTAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAA	1010		
Query 363	CCATGCACCACCTGTCACTCTGTCCCCGAAGGGGAAAACTCTATCTCTAGAGGGGTCAG	422		
Sbjct 1009	CCATGCACCACCTGTCACTCTGTCCCCGAAGGGGAAAACTCTATCTCTAGAGGGGTCAG	950		
Query 423	AGGATGTCAAGATTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACC	482		
Sbjct 949	AGGATGTCAAGATTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACC	890		
Query 483	GCTTGTGCGGGTCCCCGTCAATTCTTTGAGTTTCAACCTTGCGGTCGTAATCCCCAGGC	542		
Sbjct 889	GCTTGTGCGGGTCCCCGTCAATTCTTTGAGTTTCAACCTTGCGGTCGTAATCCCCAGGC	830		
Query 543	GGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACT	602		
Sbjct 829	GGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACT	770		
Query 603	CATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGATCCCCACGCTTTGCGA	662		
Sbjct 769	CATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGATCCCCACGCTTTGCGA	710		
Query 663	CATCAGCGTCAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCATATCTCT	722		
Sbjct 709	CATCAGCGTCAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCATATCTCT	650		
Query 723	GCGCATTTACCGCTACACATGGAAATTCACCTTTCTGCTTCTGCACTCAAGTTTTCCAG	782		
Sbjct 649	GCGCATTTACCGCTACACATGGAAATTCACCTTTCTGCTTCTGCACTCAAGTTTTCCAG	590		

klQuery_53347

Bacterium N47 16S ribosomal RNA gene, partial sequence
Bacterium N47 16S ribosomal RNA gene, partial sequence

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Staphylococcus epidermidis strain Y5 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain JDM2_4A 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain LEH7_1A 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain ECNU-UE2 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain ECNU-UI1 16S ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0023-T100-S-NIPCRAMgANb_000153 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000053 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000167 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000384 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0072-T477-S-NIPCRAMgANa_000465 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0157-T387-S-NIPCRAMgANb_000009 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0157-T387-S-NIPCRAMgANb_000108 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0169-T415-S-NIPCRAMgANa_000382 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0169-T415-S-NIPCRAMgANa_000517 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0169-T415-S-NIPCRAMgANa_000534 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0178-T478-S-NIPCRAMgANa_000084 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0026-T115-S-NIPCRAMgANa_000049 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0026-T115-S-NIPCRAMgANa_000054 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0026-T115-S-NIPCRAMgANa_000085 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0026-T115-S-NIPCRAMgANa_000101 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0026-T115-S-NIPCRAMgANa_000378 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0026-T115-S-NIPCRAMgANa_000491 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0028-T175-S-NIPCRAMgANa_000053 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000297 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000440 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000548 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000554 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000563 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0032-T156-S-NIPCRAMgANb_000564 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000076 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000155 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000193 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000217 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000397 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000425 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000467 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000482 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0038-T189-S-NPCRAMgANb_000549 small subunit ribosomal RNA gene, partial sequence
Uncultured organism clone ELU0041-T385-S-NIPCRAMgANb_000188 small subunit ribosomal RNA gene, partial sequence
Staphylococcus epidermidis gene for 16S ribosomal RNA, partial sequence, isolate: B0619
Staphylococcus epidermidis strain 59-627 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain M-S-MRS_6 16S ribosomal RNA gene, partial sequence
Bacterium NLAE-z1-G470 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain XJFH-J-1 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis partial 16S rRNA gene, isolate 0609ALT19Q2-GH
Staphylococcus epidermidis strain HKG 163 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. EllikeE4 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain HKG 182 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain HKG 183 16S ribosomal RNA gene, partial sequence
Bacterium L-20 16S ribosomal RNA gene, partial sequence
Uncultured Staphylococcus sp. clone C139100025 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. W2.10-181 16S ribosomal RNA (16S rRNA) gene, complete sequence
Staphylococcus epidermidis strain 72 (br3) 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain ATHA24 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. DF7n-C small subunit ribosomal RNA gene, partial sequence
Staphylococcus epidermidis partial 16S rRNA gene, isolate OCAT7
Staphylococcus epidermidis partial 16S rRNA gene, isolate OCAT8
Staphylococcus sp. OCAT10 partial 16S rRNA gene, isolate OCAT10
Staphylococcus epidermidis partial 16S rRNA gene, isolate OCAT31
Staphylococcus epidermidis partial 16S rRNA gene, isolate OCAT33
Staphylococcus epidermidis partial 16S rRNA gene, isolate OCOB9
Staphylococcus epidermidis strain Au22 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain B7_3CO2 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. QD53 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis partial 16S rRNA gene, strain JPR-05
Uncultured bacterium clone Ap.ba-F-DM-HN-1-27 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain W 3/4 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain W+5/13 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. PJR124 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis PM221 complete genome
Staphylococcus epidermidis strain ASI 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain C18 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain SEI, complete genome
Staphylococcus sp. RJ7 16S ribosomal RNA gene, partial sequence
Staphylococcus sp. JPR7 16S ribosomal RNA gene, partial sequence
Staphylococcus epidermidis strain P 1/1 16S ribosomal RNA gene, partial sequence
Bacterium EM-2014-133 genomic DNA containing 16S-23S intergenic spacer region, isolate 133
Uncultured bacterium gene for 16S rRNA, partial sequence, clone: smkt_Fir_002_002

firmicutes | 3 leaves

Staphylococcus warneri KP771665.1

Staphylococcus warneri strain SuMS_N03 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP771665.1|](#) Length: 1463 Number of Matches: 1

Range 1: 112 to 563 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
807 bits(894)	0.0	450/452(99%)	0/452(0%)	Plus/Plus
Query 5	ACACGTGGATAACCTACCTATAAGACTGGGATAACTTCGGGAAACCGGAGCTAATACCGG	64		
Sbjct 112	ACACGTGGATAACCTACCTATAAGACTGGGATAACTTCGGGAAACCGGAGCTAATACCGG	171		
Query 65	ATAACCCATTGAACCGCATGGTTCAATAGTGAAAGGCGGCTTTGCTGTCACCTTATAGATG	124		
Sbjct 172	ATAACATATTGAACCGCATGGTTCAATAGTGAAAGGCGGCTTTGCTGTCACCTTATAGATG	231		
Query 125	GATCCGCGCCGTATTAGCTAGTTGGTAAGGTAACGGCTTACCAAGGCAACGATACGTAGC	184		
Sbjct 232	GATCCGCGCCGTATTAGCTAGTTGGTAAGGTAACGGCTTACCAAGGCAACGATACGTAGC	291		
Query 185	CGACCTGAGAGGGTGATCGGCCACACTGGAAGTGAACACGGTCCAGACTCCTACGGGAG	244		
Sbjct 292	CGACCTGAGAGGGTGATCGGCCACACTGGAAGTGAACACGGTCCAGACTCCTACGGGAG	351		
Query 245	GCAGCAGTAGGGAATCTTCGCAATGGGCGAAAGCCTGACGGAGCAACGCCGCGTGAGTG	304		
Sbjct 352	GCAGCAGTAGGGAATCTTCGCAATGGGCGAAAGCCTGACGGAGCAACGCCGCGTGAGTG	411		
Query 305	ATGAAGGTCTTCGGATCGTAAAACTCTGTTATCAGGGAAGAACAATGTGTAAGTAACTG	364		
Sbjct 412	ATGAAGGTCTTCGGATCGTAAAACTCTGTTATCAGGGAAGAACAATGTGTAAGTAACTG	471		
Query 365	TGCACATCTTGACGGTACCTGATCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGG	424		
Sbjct 472	TGCACATCTTGACGGTACCTGATCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGG	531		
Query 425	TAATACGTAGGTGGCAAGCGTTATCCGGAATT	456		
Sbjct 532	TAATACGTAGGTGGCAAGCGTTATCCGGAATT	563		

❖ [K1Query_64441](#)
 ❖ [Staphylococcus sp. HL73 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. Iso-16 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. partial 16S rRNA gene, isolate BA-141](#)
 ❖ [Staphylococcus warneri strain 40A 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri SG1 strain SG1 16S ribosomal RNA, complete sequence](#)
 ❖ [Staphylococcus warneri strain 11BP 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured Staphylococcus sp. clone H 05 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured Staphylococcus sp. clone H 26 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured Staphylococcus sp. clone H 28 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus pasteurii strain CS11 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain SCSIO 04349 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain 7.06E02 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Bacterium LGM64 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Bacterium LGM94 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Bacterium LGM96 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. MRSE4 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. EllikeE5 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain CE83 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. C10b 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. C10c strain C10c 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain 11W6fMR22 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain y357 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain SCD2-4 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone B24-203 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain X66 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain Z52 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain M2C2 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain N7API2 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain 171B \(BR11-15\) 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain BTDF2 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium gene for 16S rRNA, partial sequence, clone: OM-C_C06](#)
 ❖ [Uncultured bacterium gene for 16S rRNA, partial sequence, clone: OM-U_B01](#)
 ❖ [Uncultured bacterium gene for 16S rRNA, partial sequence, clone: OM-U_D01](#)
 ❖ [Staphylococcus sp. HFB0021 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain Co11 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured Staphylococcus sp. clone PF34-58 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. Act36 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain 81b 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus pasteurii strain MDM7.13 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri partial 16S rRNA gene, isolate N25](#)
 ❖ [Staphylococcus warneri partial 16S rRNA gene, isolate N16](#)
 ❖ [Staphylococcus warneri partial 16S rRNA gene, isolate N17](#)
 ❖ [Staphylococcus warneri partial 16S rRNA gene, isolate N33](#)
 ❖ [Staphylococcus sp. F34 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. AK-4 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain LJI-Ka3 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured Staphylococcus sp. clone DMV_ASJ2 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. 8-1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. 8-2 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. 28-4 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. 9-7 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone nbw890d01c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone nbw898f04c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone nbw1181c01c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone nck25c10c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone nck31d06c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone ncd1392b01c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium clone ncm65d05c1 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured Staphylococcus sp. gene for 16S ribosomal RNA, partial sequence, clone: DhS1-5](#)
 ❖ [Staphylococcus sp. EP_S_84 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri genomic DNA containing 16S-23S intergenic spacer region, strain N1B](#)
 ❖ [Bacterium SCSIO13300 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. enrichment culture clone zhuowei 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. PMS08 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus warneri strain RJ24 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus pasteurii strain CS.B10 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. MY-CA13 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Staphylococcus sp. MY-CA92 16S ribosomal RNA gene, partial sequence](#)
 ❖ [Uncultured bacterium gene for 16S rRNA, partial sequence, clone: LpF35_1](#)
 ❖ [firmicutes | 31 leaves](#)

Bacillus subtilis HG764646.1

Bacillus subtilis partial 16S rRNA gene, isolate C737

Sequence ID: [emb|HG764646.1](#) Length: 995 Number of Matches: 1

Range 1: 13 to 951 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1656 bits(1836)	0.0	931/939(99%)	1/939(0%)	Plus/Minus
Query 1	CTTCGGGTGTTACAAGCTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCGGGAACGTA	60		
Sbjct 951	CTTCGGGTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCGGGAACGTA	892		
Query 61	TTCACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGC	120		
Sbjct 891	TTCACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGC	832		
Query 121	AGACTGCGATCCGAACAGAACTGAGAACTGAGAACTGAGAACTGAGAACTGAGAACTG	180		
Sbjct 831	AGACTGCGATCCGAACAGAACTGAGAACTGAGAACTGAGAACTGAGAACTGAGAACTG	772		
Query 181	CCCTTTGTTCTGTCCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTT	240		
Sbjct 771	CCCTTTGTTCTGTCCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTT	712		
Query 241	GACGTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGA	300		
Sbjct 711	GACGTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGA	652		
Query 301	ATGCTGGCAACTAAGATCAAGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGA	360		
Sbjct 651	ATGCTGGCAACTAAGATCAAGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGA	592		
Query 361	CACGAGCTGACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGACGTCCTATC	420		
Sbjct 591	CACGAGCTGACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGACGTCCTATC	532		
Query 421	TCTAGGATTGTGAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAA	480		
Sbjct 531	TCTAGGATTGTGAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAA	472		
Query 481	CCACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGTCTTGCGAC	540		
Sbjct 471	CCACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGTCTTGCGAC	412		
Query 541	CGTACTCCCCANGCGGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCC	600		
Sbjct 411	CGTACTCCCCANGCGGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCC	352		
Query 601	TAACACTTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTCGCTC	660		
Sbjct 351	TAACACTTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTCGCTC	292		
Query 661	CCCACGCTTTCGCTCCTCAGCGTCAGTTACAGANCAGAGAGTCGCCTTCGCCACTGGTGT	720		
Sbjct 291	CCCACGCTTTCGCTCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGT	232		
Query 721	TCCTCCACATCTCTACGCATTTACCGCTACACGTGGAATCCACTCTCCTCTTCTGCAC	780		
Sbjct 231	TCCTCCACATCTCTACGCATTTACCGCTACACGTGGAATCCACTCTCCTCTTCTGCAC	172		
Query 781	TCAAGTTCCCCAGTTTCCAATGACCCTCCCGGTTGAGCCGGGGGCTTTACATCAAAC	840		
Sbjct 171	TCAAGTTCCCCAGTTTCCAATGACCCTCCCGGTTGAGCCGGGGGCTTTACATCAGACT	112		
Query 841	TAAAAAACCGCTGCGAGCCCTTTACGCCCAATAATTCGCGA-AACGCTTGCCCCCTAC	899		
Sbjct 111	TAAGAAACCGCTGCGAGCCCTTTACGCCCAATAATTCGCGA-AACGCTTGCCCCCTAC	52		
Query 900	GTATTACCGCGGCTGCTGGCACGTAGTTAGCCGGGGCTT	938		
Sbjct 51	GTATTACCGCGGCTGCTGGCACGTAGTTAGCCGGGGCTT	13		

lcl/Query_13695

- Bacillus sp. WP-XY19-3 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis partial 16S rRNA gene, isolate C737
- Bacillus tequilensis strain RBEB6 16S ribosomal RNA gene, partial sequence
- Bacillus tequilensis strain Z36S 16S ribosomal RNA gene, partial sequence
- Bacillus sp. K6-15P 16S ribosomal RNA gene, partial sequence
- Bacillus amyloliquefaciens gene for 16S rRNA, partial sequence, strain: CSP
- Bacillus tequilensis strain 111-4 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain YNA13 16S ribosomal RNA gene, complete sequence
- Bacillus tequilensis strain BE-1A 16S ribosomal RNA gene, partial sequence
- Bacillus sp. DC3158 16S ribosomal RNA gene, partial sequence
- Bacillus sp. HYC-1-3 16S ribosomal RNA gene, partial sequence
- Bacillus sp. JR65 16S ribosomal RNA gene, partial sequence
- Bacillus sp. JR64 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain PEBS07031802 16S ribosomal RNA gene, partial sequence
- Bacillus amyloliquefaciens strain ASAG1 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain 1778 16S ribosomal RNA gene, partial sequence
- Bacillus licheniformis strain DJ-2 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium clone Zhuo1 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain DC2-2 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain QD399 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain B6-1 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain A9 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium clone Hswb-29 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain ADG 2 16S ribosomal RNA gene, partial sequence
- Bacterium L3 16S ribosomal RNA gene, partial sequence
- firmicutes | 2 leaves
- Azospirillum lipoferum partial 16S rRNA gene, isolate 17:1
- Bacillus subtilis strain H32 16S ribosomal RNA gene, partial sequence
- firmicutes | 4 leaves
- firmicutes | 2 leaves
- firmicutes | 5 leaves
- Bacillus subtilis subsp. spizizenii strain AB4 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis KCTC 1028, complete genome
- Bacillus subtilis subsp. subtilis strain 3NA, complete genome
- Bacillus subtilis strain PS832, complete genome
- Bacillus subtilis subsp. subtilis str. 168, complete genome
- Bacillus subtilis subsp. subtilis str. AG1839, complete genome
- Bacillus subtilis subsp. subtilis str. JH642 substr. AG174, complete genome
- Bacillus subtilis subsp. subtilis 6051-HGW, complete genome
- Bacillus subtilis subsp. subtilis str. AG1839, complete genome
- Bacillus subtilis subsp. subtilis str. JH642 substr. AG174, complete genome
- Bacillus subtilis subsp. subtilis 6051-HGW, complete genome
- Bacillus subtilis XF-1, complete genome
- Bacillus subtilis BEST7003 DNA, complete genome
- Bacillus subtilis BEST7613 DNA, complete genome
- Uncultured bacterium clone Hswb-2 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis subsp. subtilis str. 168 complete genome
- Bacillus sp. ARBELICrg 16S ribosomal RNA gene, partial sequence
- Bacillus sp. VP 18 16S ribosomal RNA gene, partial sequence
- Bacillus subtilis strain M-11 16S ribosomal RNA gene, partial sequence
- Bacterium YC-LK-LKJ44 16S ribosomal RNA gene, partial sequence
- Bacterium YC-LK-LKJ45 16S ribosomal RNA gene, partial sequence
- Bacterium YC-LK-LKJ43 16S ribosomal RNA gene, partial sequence
- Bacillus sp. KHR-38 16S ribosomal RNA gene, partial sequence
- Bacillus amyloliquefaciens partial 16S rRNA gene, isolate NBtR-1
- Bacterium Y2 16S ribosomal RNA gene, partial sequence
- Bacillus sp. 825 16S ribosomal RNA gene, partial sequence
- Bacterium YC-LK-LKJ121 16S ribosomal RNA gene, partial sequence
- firmicutes | 35 leaves

Bacillus cereus KJ612539.1

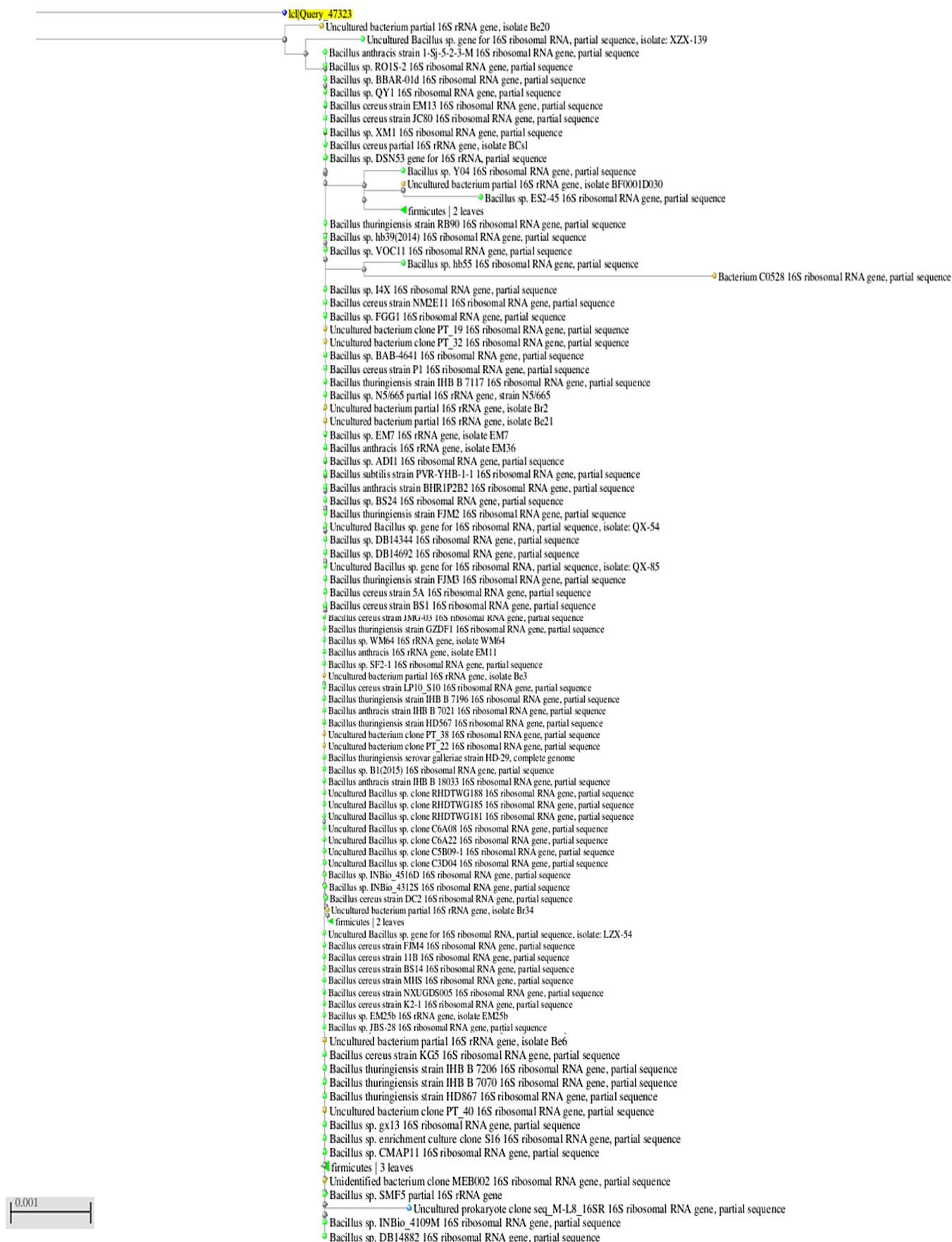
Bacillus cereus strain EM13 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ612539.1](#) Length: 1397 Number of Matches: 1

Range 1: 431 to 1378 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1660 bits(1840)	0.0	941/951(99%)	3/951(0%)	Plus/Minus
Query 1	GTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCCGC	60		
Sbjct 1378	GTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCCGC	1319		
Query 61	GGCATGTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACA	120		
Sbjct 1318	GGCATGTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACA	1259		
Query 121	ATCCGAAGTGAAGCGGTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGT	180		
Sbjct 1258	ATCCGAAGTGAAGCGGTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGT	1199		
Query 181	ACCGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCAT	240		
Sbjct 1198	ACCGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCAT	1139		
Query 241	CCCCACCTTCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGC	300		
Sbjct 1138	CCCCACCTTCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGC	1079		
Query 301	AATAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCAACATCTCAGCACAGAGCT	360		
Sbjct 1078	AATAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCAACATCTCAGCACAGAGCT	1019		
Query 361	GACGACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCTAGGGT	420		
Sbjct 1018	GACGACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCTAGGGT	959		
Query 421	TTTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGC	480		
Sbjct 958	TTTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGC	899		
Query 481	TCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTCC	540		
Sbjct 898	TCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTCC	839		
Query 541	CCAGGCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCCCTCTAACACTT	600		
Sbjct 838	CCAGGCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCCCTCTAACACTT	779		
Query 601	AGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCACGCT	660		
Sbjct 778	AGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCACGCT	719		
Query 661	TTCGCGCCTCAGTGTCAAGTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCAT	720		
Sbjct 718	TTCGCGCCTCAGTGTCAAGTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCAT	659		
Query 721	ATCTCTACGCATTTACCGCTACACATGGAATTCACCTTTCCTCTTCTGCACTCAAGTCT	780		
Sbjct 658	ATCTCTACGCATTTACCGCTACACATGGAATTCACCTTTCCTCTTCTGCACTCAAGTCT	599		
Query 781	CCCAGTTTCCAATGACCTCCACGGGTTGAGCCGTGGGCTTTCACATCAAAGTTAAAAAA	840		
Sbjct 598	CCCAGTTTCCAATGACCTCCAC-GGTTGAGCCGTGGGCTTTCACATCAGACTTAAGAAA	540		
Query 841	CCACCTGCGCGCGCTTTACGCCCAATAATTCCGGAAAACGCTTGCCCCCTACGTATTAC	900		
Sbjct 539	CCACCTGCGCGCGCTTTACGCCCAATAATTCCGGATAACGCTTGCCACCTACGTATTAC	480		
Query 901	CGCGGCTGCTGGCACGTAATTAGCCGGGGCTTTCCTGGTTAAGGTACCGTC	951		
Sbjct 479	CGCGGCTGCTGGCACGTAGTTAGCCGTGGCTTT-CTGGTT-AGGTACCGTC	431		



5- The phylogenetic analysis of bacteria isolated from lift buttons samples

Staphylococcus warneri BCL-34

Staphylococcus warneri strain MBS022 16S ribosomal RNA gene, partial sequence

Sequence ID: [KT582294.1](#) Length: 1486 Number of Matches: 1

Range 1: 820 to 1424 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1077 bits(583)	0.0	599/606(99%)	3/606(0%)	Plus/Minus
Query 19	TAAATGGTTACTCCACCGGCTTCGGGTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTG	78		
Sbjct 1424	TAAATGGTTACTCCACCGGCTTCGGGTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTG	1365		
Query 79	TACAAGACCCGGGAACGTATTACCGTAGCATGCTGATCTACGATTACTAGCGATTCCAG	138		
Sbjct 1364	TACAAGACCCGGGAACGTATTACCGTAGCATGCTGATCTACGATTACTAGCGATTCCAG	1305		
Query 139	CTTCATGTAGTCGAGTTGCAGACTACAATCCGAACTGAGAACAACCTTTATGGGATTGCT	198		
Sbjct 1304	CTTCATGTAGTCGAGTTGCAGACTACAATCCGAACTGAGAACAACCTTTATGGGATTGCT	1245		
Query 199	TGACCTCGCGGTTTAGCTGCCCTTTGTATTGTCCATTGTAGCACGTGTGTAGCCCAAATC	258		
Sbjct 1244	TGACCTCGCGGTTTAGCTGCCCTTTGTATTGTCCATTGTAGCACGTGTGTAGCCCAAATC	1185		
Query 259	ATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCA	318		
Sbjct 1184	ATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCA	1125		
Query 319	ACTTAGAGTGCCCAACTTAATGATGGCAACTAAGCTTAAGGGTTGCGCTCGTTGCGGGAC	378		
Sbjct 1124	ACTTAGAGTGCCCAACTTAATGATGGCAACTAAGCTTAAGGGTTGCGCTCGTTGCGGGAC	1065		
Query 379	TTAACCCAACATCTCACGACACGAGCTGACGACAACCATGCACCACCTGTCACTTTGTCC	438		
Sbjct 1064	TTAACCCAACATCTCACGACACGAGCTGACGACAACCATGCACCACCTGTCACTTTGTCC	1005		
Query 439	CCCGAAGGGGAAGACTCTATCTCTAGAGCGGTCAAAGGATGTCAAGATTGGTAAGGTTC	498		
Sbjct 1004	CCCGAAGGGGAAGACTCTATCTCTAGAGCGGTCAAAGGATGTCAAGATTGGTAAGGTTC	945		
Query 499	TTCGCGTTGCTTCAAATTAACCACATGCTCCACCGTTGGTGCGG--CCCCGTCAATTCT	556		
Sbjct 944	TTCGCGTTGCTTCAAATTAACCACATGCTCCACCGTTGGTGCGGTTCCCGTCAATTCC	885		
Query 557	TTTGATTTTCAACCTTGCGGTCGTACTCCCCAGGCGGAGTGCTTAATGCGTTTAGCTGCA	616		
Sbjct 884	TTTGATTTTCAACCTTGCGGTCGTACTCCCCAGGCGGAGTGCTTAATGCGTT-AGCTGCA	826		
Query 617	GCACTA 622			
Sbjct 825	GCACTA 820			

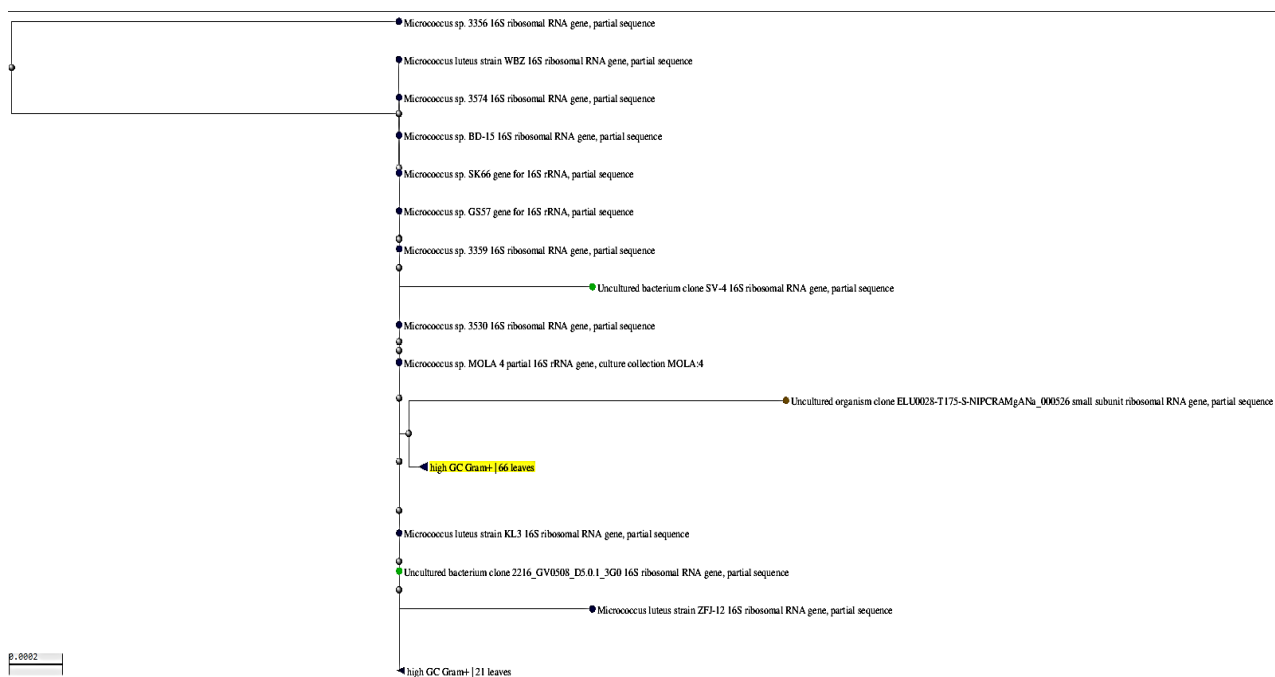


Micrococcus luteus MBS022

Micrococcus luteus strain BCL-34 16S ribosomal RNA gene, partial sequence

Sequence ID: [KM378607.1](#) Length: 1394 Number of Matches: 1

Range 1: 1 to 1393		GenBank	Graphics	▼ Next Match ▲ Previous Match	
Score	Expect	Identities	Gaps	Strand	
2560 bits(1386)	0.0	1391/1393(99%)	1/1393(0%)	Plus/Plus	
Query	1	TGC-AGTCGAACGATGAAGCCCAGCTTGCTGGGTGGATTAGTGGCGAACGGGTGAGTAAC			59
Sbjct	1	TGCAAGTCGAACGATGAAGCCCAGCTTGCTGGGTGGATTAGTGGCGAACGGGTGAGTAAC			60
Query	60	ACGTGAGTAACCTGCCCTTAACTCTGGGATAAGCCTGGGAAACTGGGTCTAATACCGGAT			119
Sbjct	61	ACGTGAGTAACCTGCCCTTAACTCTGGGATAAGCCTGGGAAACTGGGTCTAATACCGGAT			120
Query	120	AGGAGCGCCGACCGCATGGTGGGTGTTGGAAGATTTATCGGTTTTGGATGGACTCGCGG			179
Sbjct	121	AGGAGCGCCTACCGCATGGTGGGTGTTGGAAGATTTATCGGTTTTGGATGGACTCGCGG			180
Query	180	CCTATCAGCTTGTGGGTGAGGTAATGGCTCACCAAGGCGACGACGGGTAGCCGGCCTGAG			239
Sbjct	181	CCTATCAGCTTGTGGGTGAGGTAATGGCTCACCAAGGCGACGACGGGTAGCCGGCCTGAG			240
Query	240	AGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTG			299
Sbjct	241	AGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTG			300
Query	300	GGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCC			359
Sbjct	301	GGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCCGCGTGAGGGATGACGGCC			360
Query	360	TTCGGGTTGTAAACCTCTTTCAGTAGGGGAAGAAGCGAAAGTGACGGTACCTGCAGAAGAA			419
Sbjct	361	TTCGGGTTGTAAACCTCTTTCAGTAGGGGAAGAAGCGAAAGTGACGGTACCTGCAGAAGAA			420
Query	420	GCACCGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGGTGCGAGCGTTATCCGGA			479
Sbjct	421	GCACCGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGGTGCGAGCGTTATCCGGA			480
Query	480	ATTATTGGGCGTAAAGAGCTCGTAGGCGGTTTGTGCGCTCTGTCTGTGAAAGTCCGGGGCT			539
Sbjct	481	ATTATTGGGCGTAAAGAGCTCGTAGGCGGTTTGTGCGCTCTGTCTGTGAAAGTCCGGGGCT			540
Query	540	TAACCCCGGATCTGCGGTGGGTACGGGCAGACTAGAGTGCAGTAGGGGAGACTGGAATTC			599
Sbjct	541	TAACCCCGGATCTGCGGTGGGTACGGGCAGACTAGAGTGCAGTAGGGGAGACTGGAATTC			600
Query	600	CTGGTGTAGCGGTGGAATGCGCAGATATCAGGAGGAACACCGATGGCGAAGGCAGGTCTC			659
Sbjct	601	CTGGTGTAGCGGTGGAATGCGCAGATATCAGGAGGAACACCGATGGCGAAGGCAGGTCTC			660
Query	660	TGGGCTGTAACTGACGCTGAGGAGCGAAAGCATGGGGAGCGAACAGGATTAGATACCCTG			719
Sbjct	661	TGGGCTGTAACTGACGCTGAGGAGCGAAAGCATGGGGAGCGAACAGGATTAGATACCCTG			720
Query	720	GTAGTCCATGCCGTAAACGTTGGGCACTAGGTGTGGGGACCATTCCACGGTTTCCGCGCC			779
Sbjct	721	GTAGTCCATGCCGTAAACGTTGGGCACTAGGTGTGGGGACCATTCCACGGTTTCCGCGCC			780
Query	780	GCAGCTAACGCATTAAGTGCCCGCCCTGGGGAGTACGGCCGCAAGGCTAAAACCTCAAAGG			839
Sbjct	781	GCAGCTAACGCATTAAGTGCCCGCCCTGGGGAGTACGGCCGCAAGGCTAAAACCTCAAAGG			840
Query	840	AATTGACGGGGGGCCCGCACAAAGCGGCGGAGCATGCGGATTAATTCGATGCAACGCGAAGA			899
Sbjct	841	AATTGACGGGGGGCCCGCACAAAGCGGCGGAGCATGCGGATTAATTCGATGCAACGCGAAGA			900
Query	900	ACCTTACCAAGGCTTGACATGTTCTCGATCGCCGTAGAGATACGGTTTCCCCTTTGGGGC			959
Sbjct	901	ACCTTACCAAGGCTTGACATGTTCTCGATCGCCGTAGAGATACGGTTTCCCCTTTGGGGC			960
Query	960	GGGTTTCACAGGTGGTGCATGGTTGTCTGTCAGCTCGTGTCTGAGATGTTGGGTTAAGTCC			1019
Sbjct	961	GGGTTTCACAGGTGGTGCATGGTTGTCTGTCAGCTCGTGTCTGAGATGTTGGGTTAAGTCC			1020
Query	1020	CGCAACGAGCGCAACCCTCGTTCCATGTTGCCAGCAGCTAATGGTGGGGACTCATGGGAG			1079
Sbjct	1021	CGCAACGAGCGCAACCCTCGTTCCATGTTGCCAGCAGCTAATGGTGGGGACTCATGGGAG			1080
Query	1080	ACTGCCGGGGTCAACTCGGAGGAAGGTGAGGACGACGTCAAATCATCATGCCCCCTTATGT			1139
Sbjct	1081	ACTGCCGGGGTCAACTCGGAGGAAGGTGAGGACGACGTCAAATCATCATGCCCCCTTATGT			1140
Query	1140	CTTGGGCTTCACGCATGCTACAATGGCCGGTACAATGGGTTGCGATACTGTGAGGTGGAG			1199
Sbjct	1141	CTTGGGCTTCACGCATGCTACAATGGCCGGTACAATGGGTTGCGATACTGTGAGGTGGAG			1200
Query	1200	CTAATCCCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTC			1259
Sbjct	1201	CTAATCCCAAAAAGCCGGTCTCAGTTCGGATTGGGGTCTGCAACTCGACCCCATGAAGTC			1260
Query	1260	GGAGTCGCTAGTAATCGCAGATCAGCAACGCTGCGGTGAATACGTTCCCGGGCCTTGTAC			1319
Sbjct	1261	GGAGTCGCTAGTAATCGCAGATCAGCAACGCTGCGGTGAATACGTTCCCGGGCCTTGTAC			1320
Query	1320	ACACCGCCCGTCAAGTCACGAAAGTCGGTAACACCCGAAGCCGGTGGCCTAACCCCTTGTG			1379
Sbjct	1321	ACACCGCCCGTCAAGTCACGAAAGTCGGTAACACCCGAAGCCGGTGGCCTAACCCCTTGTG			1380
Query	1380	GGGGAGCCGTCGA	1392		
Sbjct	1381	GGGGAGCCGTCGA	1393		



Staphylococcus epidermidis

Staphylococcus epidermidis partial 16S rRNA gene, strain BGHMC5

Sequence ID: [FR797804.1](#) Length: 815 Number of Matches: 1

Range 1: 1 to 815		GenBank	Graphics	▼ Next Match ▲ Previous Match	
Score		Expect	Identities	Gaps	Strand
1506 bits(815)		0.0	815/815(100%)	0/815(0%)	Plus/Plus
Query	1	AGAAAGTGGAAATCCCATGTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAGTGG			60
Sbjct	1	AGAAAGTGGAAATCCCATGTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAGTGG			60
Query	61	CGAAGGCGACTTTCTGGTCTGTAACTGACGCTGATGTGCGAAAGCGTGGGGATCAAACAG			120
Sbjct	61	CGAAGGCGACTTTCTGGTCTGTAACTGACGCTGATGTGCGAAAGCGTGGGGATCAAACAG			120
Query	121	GATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCC			180
Sbjct	121	GATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCC			180
Query	181	GCCCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGACCGCAAGGT			240
Sbjct	181	GCCCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGACCGCAAGGT			240
Query	241	TGAAACTCAAAGGAATTGACGGGGACCCGACAAAGCGGTGGAGCATGTGGTTTAATTCGA			300
Sbjct	241	TGAAACTCAAAGGAATTGACGGGGACCCGACAAAGCGGTGGAGCATGTGGTTTAATTCGA			300
Query	301	AGCAACGCGAAGAACCTTACCAAATCTTGACATCCTCTGACCCCTCTAGAGATAGAGTTT			360
Sbjct	301	AGCAACGCGAAGAACCTTACCAAATCTTGACATCCTCTGACCCCTCTAGAGATAGAGTTT			360
Query	361	TCCCCCTTCGGGGGACAGAGTGACAGGTGGTGCATGGTTGTCTCGTCAGCTCGTGTCTGAGA			420
Sbjct	361	TCCCCCTTCGGGGGACAGAGTGACAGGTGGTGCATGGTTGTCTCGTCAGCTCGTGTCTGAGA			420
Query	421	TGTTGGGTTAAGTCCCGCAACGAGCGCAACCCCTTAAGCTTAGTTGCCATCATTAAAGTTGG			480
Sbjct	421	TGTTGGGTTAAGTCCCGCAACGAGCGCAACCCCTTAAGCTTAGTTGCCATCATTAAAGTTGG			480
Query	481	GCACTCTAAGTTGACTGCCGGTGACAAACCGGAGGAAGGTGGGGATGACGTCAAATCATC			540
Sbjct	481	GCACTCTAAGTTGACTGCCGGTGACAAACCGGAGGAAGGTGGGGATGACGTCAAATCATC			540
Query	541	ATGCCCCCTTATGATTTGGGCTACACACGTGCTACAATGGACAATACAAAGGGCAGCGAAA			600
Sbjct	541	ATGCCCCCTTATGATTTGGGCTACACACGTGCTACAATGGACAATACAAAGGGCAGCGAAA			600
Query	601	CCGCGAGGTCAAGCAAATCCCATAAAGTTGTTCTCAGTTCGGATTGTAGTCTGCAACTCG			660
Sbjct	601	CCGCGAGGTCAAGCAAATCCCATAAAGTTGTTCTCAGTTCGGATTGTAGTCTGCAACTCG			660
Query	661	ACTATATGAAGCTGGAATCGCTAGTAATCGTAGATCAGCATGCTACGGTGAATACGTTCC			720
Sbjct	661	ACTATATGAAGCTGGAATCGCTAGTAATCGTAGATCAGCATGCTACGGTGAATACGTTCC			720
Query	721	CGGGTCTTGTACACACCGCCCGTCACACCACGAGAGTTTGTAACACCCGAAGCCGGTGG			780
Sbjct	721	CGGGTCTTGTACACACCGCCCGTCACACCACGAGAGTTTGTAACACCCGAAGCCGGTGG			780
Query	781	GTAACCATTGGAGCTAGCCGTTGAAGGGGGACAAA	815		
Sbjct	781	GTAACCATTGGAGCTAGCCGTTGAAGGGGGACAAA	815		



6 - The phylogenetic analysis of bacteria isolated from vacuum cleaner dust samples

Bacillus thuringiensis

FJ174596.1

Bacillus thuringiensis strain 104XG46 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|FJ174596.1](#) Length: 1049 Number of Matches: 1

Range 1: 59 to 1018 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1651 bits(1830)	0.0	944/960(98%)	3/960(0%)	Plus/Plus
Query 1	AGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCATAAGACTGGGATAACTCCGGG	60		
Sbjct 59	AGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCATAAGACTGGGATAACTCCGGG	118		
Query 61	AAACCGGGGCTAATACCGGATAATATTTTGAAGTGCATGGTTCGAAATTGAAAGGCGGCT	120		
Sbjct 119	AAACCGGGGCTAATACCGGATAACATTTTGAAGTGCATGGTTCGAAATTGAAAGGCGGCT	178		
Query 121	TCGGCTGTCACCTTATGGATGGACCCGCGTCGCATTAGCTAGTTGGTGGAGTAACGGCTCA	180		
Sbjct 179	TCGGCTGTCACCTTATGGATGGACCCGCGTCGCATTAGCTAGTTGGTGGAGTAACGGCTCA	238		
Query 181	CCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACAC	240		
Sbjct 239	CCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACAC	298		
Query 241	GGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCGCAATGGACGAAAGTCTGAC	300		
Sbjct 299	GGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCGCAATGGACGAAAGTCTGAC	358		
Query 301	GGAGCAACGCCGCGTGAGTGATGAAGGCTTTTCGGGTCGTAAACTCTGTTGTTAGGGAAG	360		
Sbjct 359	GGAGCAACGCCGCGTGAGTGATGAAGGCTTTTCGGGTCGTAAACTCTGTTGTTAGGGAAG	418		
Query 361	AACAAGTGCTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACCAGAAAGCCACGGCTA	420		
Sbjct 419	AACAAGTGCTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACCAGAAAGCCACGGCTA	478		
Query 421	ACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGC	480		
Sbjct 479	ACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGC	538		
Query 481	GTAAAGCGCGCGCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGG	540		
Sbjct 539	GTAAAGCGCGCGCAGGTGGTTTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGG	598		
Query 541	GGGTCATTGGAACTGGGAGACTTGAGTGCAGAAGAGGAAAGTGAATTCCATGTGTAGC	600		
Sbjct 599	GGGTCATTGGAACTGGGAGACTTGAGTGCAGAAGAGGAAAGTGAATTCCATGTGTAGC	658		
Query 601	GGTGAAATGCGTAGAGATATGGAGGAACACCAAGTGGCGAAGGCGACTTTCTGGTCTGTAA	660		
Sbjct 659	GGTGAAATGCGTAGAGATATGGAGGAACACCAAGTGGCGAAGGCGACTTTCTGGTCTGTAA	718		
Query 661	CTGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACG	720		
Sbjct 719	CTGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACG	778		
Query 721	CCGTAAACGATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGC	780		
Sbjct 779	CCGTAAACGATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGC	838		
Query 781	ATTAAGCACTCCGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAGGAATTGACGGGGG	840		
Sbjct 839	ATTAAGCACTCCGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAGGAATTGACGGGGG	898		
Query 841	CCCGCACAANCGG-GGAGCATGGGGTTTAATTCAAAGC-ACGCGAAAACCTTACCAGGTC	898		
Sbjct 899	CCCGCACAAGCGGTGGAGCATGTGGTTTAATTCAAAGCAACGCGAGAACCTTACCAGGTC	958		
Query 899	TTGACTTCTCTGAAACCTANAAATAGGGCTTC-CCTTCGGGAACAAAAGGACAggggg	957		
Sbjct 959	TTGACATCCTCTGAAACCTAGAGATAGGGCTTCCTTCGGGAGCAGAGTGACAGGTGG	1018		



Bacillus mycoides KR088435.1

Bacillus mycoides strain St02 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KR088435.1](#) Length: 1398 Number of Matches: 1

Range 1: 396 to 1367 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1703 bits(1888)	0.0	964/973(99%)	3/973(0%)	Plus/Minus	
Query 1	TCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGGGCATGCTG				60
Sbjct 1367	TCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGGGCATGCTG				1308
Query 61	ATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACAATCCGAAC				120
Sbjct 1307	ATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACAATCCGAAC				1248
Query 121	GAGAACGGTTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGTACCGTCCAT				180
Sbjct 1247	GAGAACGGTTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGTACCGTCCAT				1188
Query 181	TGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTT				240
Sbjct 1187	TGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTT				1128
Query 241	CCTCCGGTTTGTACCCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGCAACTAAGAT				300
Sbjct 1127	CCTCCGGTTTGTACCCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGCAACTAAGAT				1068
Query 301	CAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAAC				360
Sbjct 1067	CAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAAC				1008
Query 361	CATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTTGTGAGAGG				420
Sbjct 1007	CATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTTGTGAGAGG				948
Query 421	ATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCT				480
Sbjct 947	ATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCT				888
Query 481	TGTGCGGGCCCCGTCAATTCCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCAGGCGGA				540
Sbjct 887	TGTGCGGGCCCCGTCAATTCCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCAGGCGGA				828
Query 541	GTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCCTCTAACACTTAGCACTCAT				600
Sbjct 827	GTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCCTCTAACACTTAGCACTCAT				768
Query 601	CGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGCGCCT				660
Sbjct 767	CGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGCGCCT				708
Query 661	CAGTGTGAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCATATCTCTACG				720
Sbjct 707	CAGTGTGAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTCCTCCATATCTCTACG				648
Query 721	CATTTACCGCTACACATGGAATTCACCTTTCTCTTCTGCACTCAAGTCTCCAGTTTC				780
Sbjct 647	CATTTACCGCTACACATGGAATTCACCTTTCTCTTCTGCACTCAAGTCTCCAGTTTC				588
Query 781	CAATGACCTCCACGGTTGAGCCGTGGGCTTTTACATCAAACCTTAAAAAACCACCTGCGC				840
Sbjct 587	CAATGACCTCCACGGTTGAGCCGTGGGCTTTTACATCAGACTTAAGAAACCACCTGCGC				528
Query 841	GCGCTTTACGCCCAATAATTCCGGA-AACGCTTGCCACCTACGTATTACCGCGGCTGCTG				899
Sbjct 527	GCGCTTTACGCCCAATAATTCCGGAATAACGCTTGCCACCTACGTATTACCGCGGCTGCTG				468
Query 900	GCACGTAATTAGCCG-GGCTTTCTGGGTTAGGTACCGTCAAGGGGCCAGCTTATTCAACT				958
Sbjct 467	GCACGTAGTTAGCCGTGGCTTTCT-GGTAGGTACCGTCAAGGTGCCAGCTTATTCAACT				409
Query 959	AACCCTTGTTCTT	971			
Sbjct 408	AGCACTTGTTCTT	396			



Bacillus licheniformis DQ071560.1

Bacillus licheniformis strain MKU 1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|DQ071560.1](#) Length: 1430 Number of Matches: 1

Range 1: 425 to 1393 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1694 bits(1878)	0.0	957/969(99%)	0/969(0%)	Plus/Minus
Query 1	TTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCCGCG	60		
Sbjct 1393	TTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCCGCG	1334		
Query 61	GCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGA	120		
Sbjct 1333	GCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGA	1274		
Query 121	TCCGAACTGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTT	180		
Sbjct 1273	TCCGAACTGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTT	1214		
Query 181	CTGCCCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATC	240		
Sbjct 1213	CTGCCCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATC	1154		
Query 241	CCCACCTTCCCTCCGGTTTGTACCCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCA	300		
Sbjct 1153	CCCACCTTCCCTCCGGTTTGTACCCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCA	1094		
Query 301	ACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTG	360		
Sbjct 1093	ACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTG	1034		
Query 361	ACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTT	420		
Sbjct 1033	ACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTT	974		
Query 421	GTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCT	480		
Sbjct 973	GTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCT	914		
Query 481	CCACCGCTTGTGCGGGCCCCCGTCAATTCTTTGAGTTTCAGTCTTGCGACCGTACTCCC	540		
Sbjct 913	CCACCGCTTGTGCGGGCCCCCGTCAATTCTTTGAGTTTCAGTCTTGCGACCGTACTCCC	854		
Query 541	CAGGCGGAGTGCTTAATGCGTTTGCTGCAGCACTAAAGGGCGGAAACCCTCTAACACTTA	600		
Sbjct 853	CAGGCGGAGTGCTTAATGCGTTTGCTGCAGCACTAAAGGGCGGAAACCCTCTAACACTTA	794		
Query 601	GCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTCGCTCCCCACGCTT	660		
Sbjct 793	GCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTCGCTCCCCACGCTT	734		
Query 661	TCGCGCCTCAGCGTCAGTTACAGACCAGAGAGTCGCTTCGCCACTGGTGTTCCTCCACA	720		
Sbjct 733	TCGCGCCTCAGCGTCAGTTACAGACCAGAGAGTCGCTTCGCCACTGGTGTTCCTCCACA	674		
Query 721	TCTCTACGCATTTACCGCTACACGTGGAATCCACTCTCCTCTTCTGCACTCAAGTTCC	780		
Sbjct 673	TCTCTACGCATTTACCGCTACACGTGGAATCCACTCTCCTCTTCTGCACTCAAGTTCC	614		
Query 781	CCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTAAAAAACC	840		
Sbjct 613	CCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTAAAAAACC	554		
Query 841	GCCTGCGCGCGCTTTACCCCAATAATTCCCGGAAACCCTTGCCACCTACGTATTACCGC	900		
Sbjct 553	GCCTGCGCGCGCTTTACCCCAATAATTCCCGGAAACCCTTGCCACCTACGTATTACCGC	494		
Query 901	GGCTGCTGGGACGTAATTAGCCGGGGCTTTCTGGTTAGGTACCGTCAAGGAACCCCCCTA	960		
Sbjct 493	GGCTGCTGGGACGTAATTAGCCGGGGCTTTCTGGTTAGGTACCGTCAAGGTACCGCCCTA	434		
Query 961	TTCAAAAGG	969		
Sbjct 433	TTCGAACGG	425		



Bacillus subtilis KF220577.1

Bacillus subtilis strain YLB-P1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KF220577.1|](#) Length: 1050 Number of Matches: 1

Range 1: 51 to 1025 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score		Expect	Identities	Gaps	Strand	
1653 bits(1832)		0.0	955/975(98%)	5/975(0%)	Plus/Plus	
Query	1	CTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGATA				60
Sbjct	51	CTGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGATA				110
Query	61	ACTCCGGGAAACCGGGGCTAATACCGGATGGTTGTTTGAACCGCATGGTTCAAACATAAA				120
Sbjct	111	ACTCCGGGAAACCGGGGCTAATACCGGATGGTTGTTTGAACCGCATGGTTCAAACATAAA				170
Query	121	AGGTGGCTTYYGGCTACCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGTA				180
Sbjct	171	AGGTGGCTTCGGCTACCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGTA				230
Query	181	ACGGCTCACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGAC				240
Sbjct	231	ACGGCTCACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGAC				290
Query	241	TGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAA				300
Sbjct	291	TGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAA				350
Query	301	AGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTGT				360
Sbjct	351	AGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTGT				410
Query	361	TAGGGAAGAACAAGTACCGTTTCAATAGGGCGGTACCTTGACGGTACCTAACCAGAAAGC				420
Sbjct	411	TAGGGAAGAACAAGTACCGTTTCAATAGGGCGGTACCTTGACGGTACCTAACCAGAAAGC				470
Query	421	CACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAAT				480
Sbjct	471	CACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAAT				530
Query	481	TATTGGGCGTAAAGGGCTCGCAGGCGGTTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCA				540
Sbjct	531	TATTGGGCGTAAAGGGCTCGCAGGCGGTTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCA				590
Query	541	ACCGGGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCCA				600
Sbjct	591	ACCGGGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCCA				650
Query	601	CGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCTG				660
Sbjct	651	CGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCTG				710
Query	661	GTCTGTAACCTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT				720
Sbjct	711	GTCTGTAACCTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT				770
Query	721	AGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTTCCGCCCTTAGTGCTGCA				780
Sbjct	771	AGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTTCCGCCCTTAGTGCTGCA				830
Query	781	GCTAACGCATTAAGCACTCCGCCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAAT				840
Sbjct	831	GCTAACGCATTAAGCACTCCGCCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAAT				890
Query	841	TCACGGGGGGCCCGCACAAAGCGGGGAGCATGTGATTT-ATTCTGAAGC-ACGCGAAAACCT				898
Sbjct	891	TGACGGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCTGAAGCAACGCGAAGACCT				950
Query	899	TACCA-GTCTTGACTTCCTCTGAGATCCTaaaaaaaGGACGTCCCCCTTC-GGGCAAAAGG				956
Sbjct	951	TACCAGGTCTTGACATCCTCTGACATCCTAGAGATAGGACGTCCCCCTTCGGGGCCAAGTG				1010
Query	957	ACAG-GGGGGATGGT	970			
Sbjct	1011	ACAGTGGTGCATGGT	1025			



7-The phylogenetic analysis of bacteria isolated from sole of shoes samples

Echireschia coli LN558643.1

Escherichia coli partial 16S rRNA gene, isolate RS1

Sequence ID: [emb|LN558643.1](#) Length: 1484 Number of Matches: 1

Range 1: 87 to 1069 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1710 bits(1896)	0.0	972/983(99%)	5/983(0%)	Plus/Plus
Query 1	AACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGTG	60		
Sbjct 87				
Query 61	CATTCTGATCCACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGACTCCAAT	120		
Sbjct 147				
Query 121	CCGGACTACGACGCACTTTATGAGGTCCGCTTGCTCTCGCGAGGTCGCTTCTCTTTGTAT	180		
Sbjct 207				
Query 181	GCGCCATTGTAGCACGTGTGTAGCCCTGGTCGTAAGGGCCATGATGACTTGACGTCATCC	240		
Sbjct 267				
Query 241	CCACCTTCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGGCCGGACCGCTGGCA	300		
Sbjct 327				
Query 301	ACAAAGGATAAAGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTACAACACGAGCTG	360		
Sbjct 387				
Query 361	ACGACAGCCATGCAGCACCTGTCTCACAGTTCCCGAAGGCACCAATCCATCTCTGGAAAG	420		
Sbjct 447				
Query 421	TTCTGTGGATGTCAAGACCAGGTAAGGTTCTTCGCGTTGCATCGAATTAAACCACATGCT	480		
Sbjct 507				
Query 481	CCACCGCTTGTGCGGGCCCCCGTCAATTCAATTGAGTTTTAACCTTGCGGGCGTACTCCC	540		
Sbjct 567				
Query 541	CAGGCGGTGCGACTTAACGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACCTCCAAG	600		
Sbjct 627				
Query 601	TCGACATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACGCTTT	660		
Sbjct 687				
Query 661	CGCACCTGAGCGTCAGTCTTCGTCCAGGGGGCCGCCTTCGCCACCGGTATTCTCCAGAT	720		
Sbjct 747				
Query 721	CTCTACGCATTTACCGCTACACCTGGAATTCTACCCCCCTCTACGAGACTCAAGCTTGC	780		
Sbjct 807				
Query 781	CAGTATCAGATGCAGTTCCAGGTTGAGCCCGGGATTTACATCTGACTTAACAAACCG	840		
Sbjct 867				
Query 841	CCTGCGTGCCTTTACGCCAGTAATTCCGATTACGCTTGC-CCCTCCGTATTACCGCG	899		
Sbjct 927				
Query 900	GCTGCTGGCACGGAATTAGCCGGGCTTC-TCTGGGGTAACGTCAATGAAC-AAGGTAT	957		
Sbjct 987				
Query 958	TA--CTTACTCCCTTcccccccc 978			
Sbjct 1047				



Brevibacillus borstelensis KT239000.1

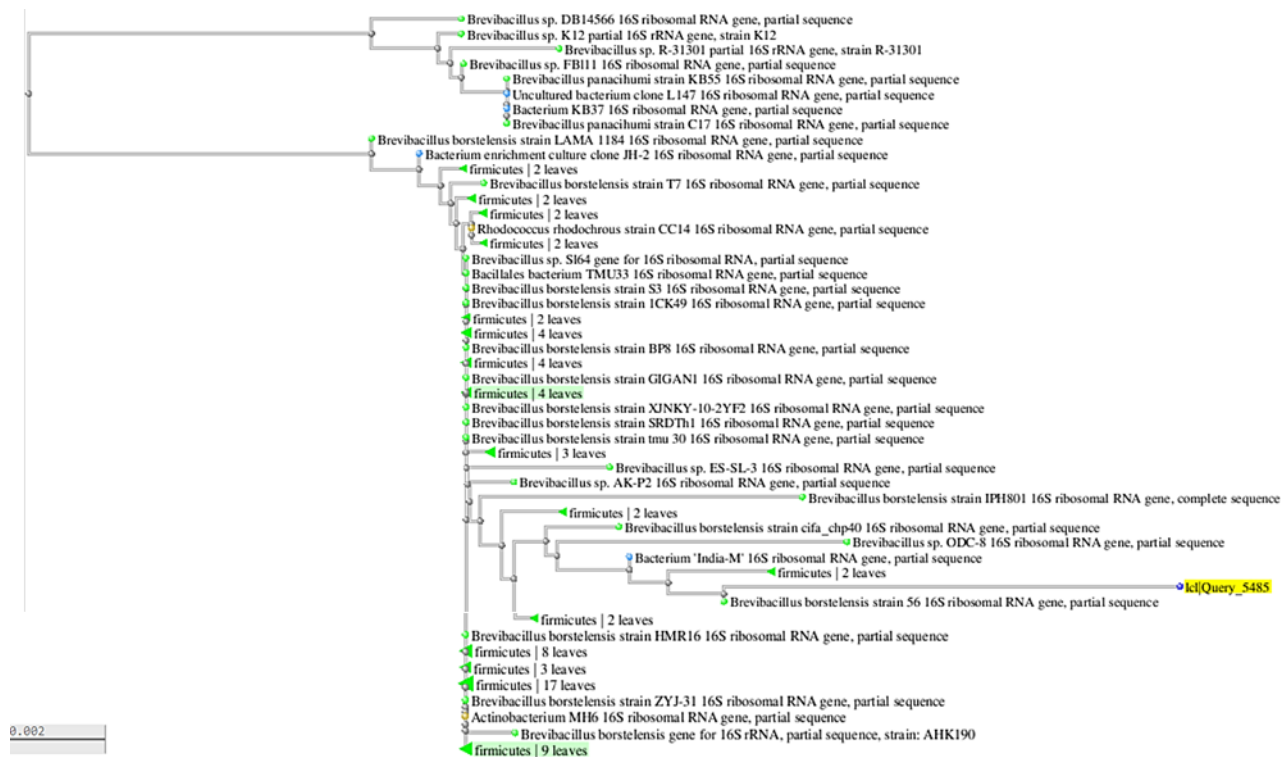
Brevibacillus borstelensis strain HMR16 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT239000.1](#) Length: 1408 Number of Matches: 1

Range 1: 46 to 1030 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1676 bits(1858)	0.0	966/985(98%)	4/985(0%)	Plus/Plus
Query 1	GAGTAACACGTAGGCAACCTGCCCGTAAGCTCGGGATAACATGGGGAAACTCATGCTAAT	60		
Sbjct 46	GAGTAACACGTAGGCAACCTGCCCGTAAGCTCGGGATAACATGGGGAAACTCATGCTAAT	105		
Query 61	ACCGGATAGGGTCTTCTCTCGCATGAGAGGAGACGGAAAGGTGGCGCAAGCTACCACTTA	120		
Sbjct 106	ACCGGATAGGGTCTTCTCTCGCATGAGAGGAGACGGAAAGGTGGCGCAAGCTACCACTTA	165		
Query 121	CGGATGGGCCTGCGGCGCATTAGCTAGTTGGTGGGGTAACGGCCTACCAAGGCGACGATG	180		
Sbjct 166	CGGATGGGCCTGCGGCGCATTAGCTAGTTGGTGGGGTAACGGCCTACCAAGGCGACGATG	225		
Query 181	CGTAGCCGACCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTA	240		
Sbjct 226	CGTAGCCGACCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTA	285		
Query 241	CGGGAGGCAGCAGTAGGGAATTTCCACAATGGACGAAAGTCTGATGGAGCAACGCCGCG	300		
Sbjct 286	CGGGAGGCAGCAGTAGGGAATTTCCACAATGGACGAAAGTCTGATGGAGCAACGCCGCG	345		
Query 301	TGAACGATGAAGGTCTTCGGATTGTAAAGTTCTGTTGTCAGAGACGAACAAGTACCGTTC	360		
Sbjct 346	TGAACGATGAAGGTCTTCGGATTGTAAAGTTCTGTTGTCAGAGACGAACAAGTACCGTTC	405		
Query 361	GAACAGGGCGGTACCTTGACGGTACCTGACGAGAAAGCCACGGCTAACTACGTGCCAGCA	420		
Sbjct 406	GAACAGGGCGGTACCTTGACGGTACCTGACGAGAAAGCCACGGCTAACTACGTGCCAGCA	465		
Query 421	GCCGCGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCGCA	480		
Sbjct 466	GCCGCGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCGCA	525		
Query 481	GGCGGCTATGTAAGTCTGGTGTAAAGCCCGGGGCTCAACCCCGGTTTCGCATCGGAAACT	540		
Sbjct 526	GGCGGCTATGTAAGTCTGGTGTAAAGCCCGGGGCTCAACCCCGGTTTCGCATCGGAAACT	585		
Query 541	GTGTAGCTTGAGTGCGAGAAGAGGAAAGCGGTATTCCACGTGTAGCGGTGAAATGCGTAGA	600		
Sbjct 586	GTGTAGCTTGAGTGCGAGAAGAGGAAAGCGGTATTCCACGTGTAGCGGTGAAATGCGTAGA	645		
Query 601	GATGTGGAGGAACACCAAGTGGCGAAGGCGGCTTTCTGGTCTGTAAGTACGCTGAGGCGC	660		
Sbjct 646	GATGTGGAGGAACACCAAGTGGCGAAGGCGGCTTTCTGGTCTGTAAGTACGCTGAGGCGC	705		
Query 661	GAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGT	720		
Sbjct 706	GAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGT	765		
Query 721	GCTAGGTGTTGGGGGTTTCAATACCCTCAGTGCCGCAGCTAACGCAATAAGCACTCCGCC	780		
Sbjct 766	GCTAGGTGTTGGGGGTTTCAATACCCTCAGTGCCGCAGCTAACGCAATAAGCACTCCGCC	825		
Query 781	TGGGGAGTACGCTCGCAAGAGTGAAACTCAAAGGATTGACGGGGGCCCGCACAAGCGG-	839		
Sbjct 826	TGGGGAGTACGCTCGCAAGAGTGAAACTCAAAGGATTGACGGGGGCCCGCACAAGCGGT	885		
Query 840	GGAGCATGTGGTTTAAATTCAGCAACGCGAAAAACCTTACCA-GTCTTGACATCCCGCT	898		
Sbjct 886	GGAGCATGTGGTTTAAATTCAGCAACGCGAAAAACCTTACCAAGTCTTGACATCCCGCT	945		
Query 899	GACCGTCCTAAAAAAGGGCTTCCTTTC-GGGCAGCGG-GACggggggggCATGGTTGTC	956		
Sbjct 946	GACCGTCCTAGAGATAGGGCTTCCTTTCGGGGCAGCGGTGACAGGTGGTGCATGGTTGTC	1005		
Query 957	GTCCCTCGGGTCGGGAAATGTGGG	981		
Sbjct 1006	GTCAGCTCGTGTCTGAGATGTTGG	1030		



Bacillus licheniformis KP772335.1

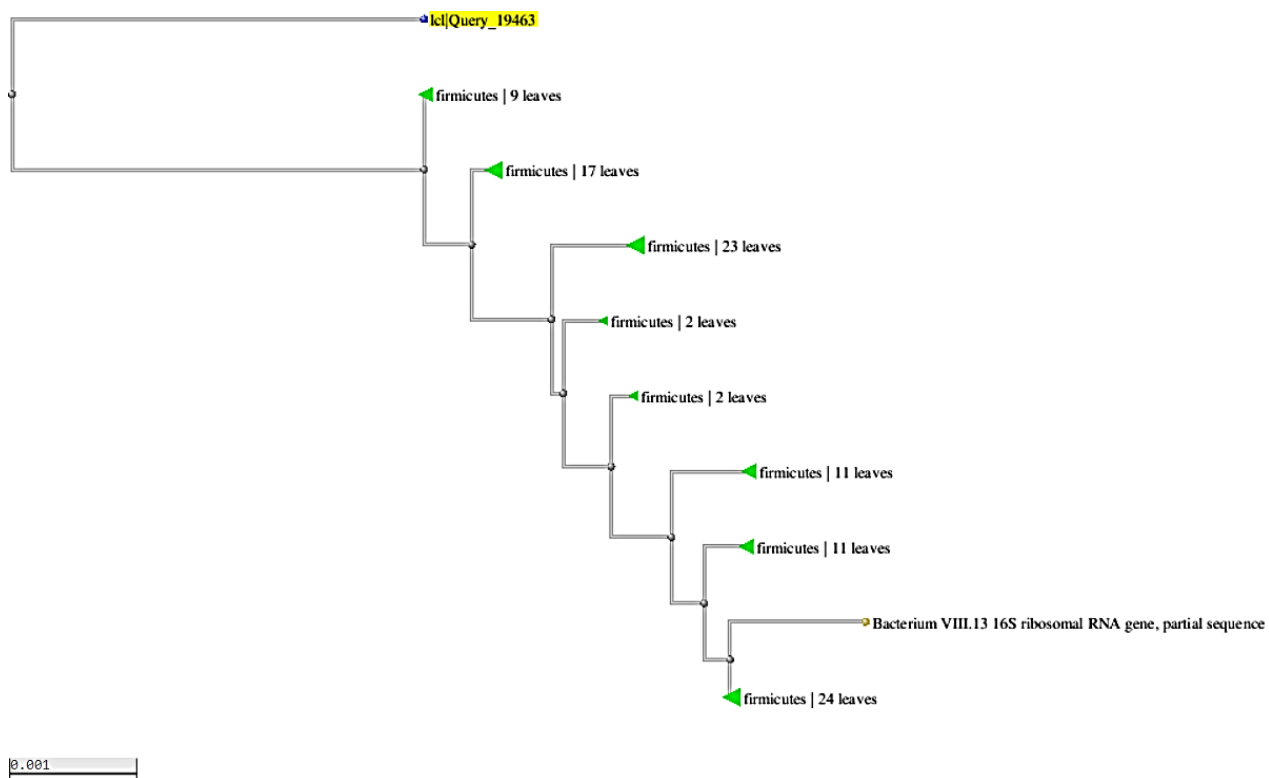
Bacillus licheniformis strain OALB2 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP772335.1|](#) Length: 1445 Number of Matches: 1

Range 1: 45 to 989 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1645 bits(1824)	0.0	935/945(99%)	4/945(0%)	Plus/Plus
Query 1	GGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGATAACTCCGGGAAA	60		
Sbjct 45	GGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGATAACTCCGGGAAA	104		
Query 61	CCGGGGCTAATACCGGATGCTTGATTGAACCGCATGGTTCAATTATAAAAGGTGGCTTTC	120		
Sbjct 105	CCGGGGCTAATACCGGATGCTTGATTGAACCGCATGGTTCAATTATAAAAGGTGGCTTTC	164		
Query 121	AGCTACCACTTGACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGTAACGGCTCACC	180		
Sbjct 165	AGCTACCACTTGACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGTAACGGCTCACC	224		
Query 181	AAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGG	240		
Sbjct 225	AAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGG	284		
Query 241	CCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGG	300		
Sbjct 285	CCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGG	344		
Query 301	AGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAACTCTGTTGTTAGGGAAGAA	360		
Sbjct 345	AGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAACTCTGTTGTTAGGGAAGAA	404		
Query 361	CAAGTACCGTTTCGAATAGGGCGGTACCTTGACGGTACCTAACCAGAAAGCCACGGCTAAC	420		
Sbjct 405	CAAGTACCGTTTCGAATAGGGCGGTACCTTGACGGTACCTAACCAGAAAGCCACGGCTAAC	464		
Query 421	TACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGT	480		
Sbjct 465	TACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGT	524		
Query 481	AAAGCGCGCGCAGGCGGTTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCAACCGGGGAGG	540		
Sbjct 525	AAAGCGCGCGCAGGCGGTTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCAACCGGGGAGG	584		
Query 541	GTCATTGGAAACTGGGGAACCTTGAGTGCAGAAGAGGAGAGTGGAAATCCACGTGTAGCGG	600		
Sbjct 585	GTCATTGGAAACTGGGGAACCTTGAGTGCAGAAGAGGAGAGTGGAAATCCACGTGTAGCGG	644		
Query 601	TGAAATGCGTAGAGATGTGGAGGAACACCAAGTGGCGAAGGCGACTCTCTGGTCTGTAACT	660		
Sbjct 645	TGAAATGCGTAGAGATGTGGAGGAACACCAAGTGGCGAAGGCGACTCTCTGGTCTGTAACT	704		
Query 661	GACGCTGAGGCGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCC	720		
Sbjct 705	GACGCTGAGGCGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGTAGTCCACGCC	764		
Query 721	GTAACGATGAGTGCTAAGTGTTAAAGGGTTTCCGCCCTTTAGTGCTGCAGCAAACGCAT	780		
Sbjct 765	GTAACGATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGCAGCAAACGCAT	824		
Query 781	TAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTC-AAGGAATTGACGGGGGC	839		
Sbjct 825	TAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGC	884		
Query 840	CCGCCAAGCGGGGAGCATGTGGTTTAATTGCAAGCAACGCGAA-AACCTTACCA-GTC	897		
Sbjct 885	CCGCACAAGCGGTGGAGCATGTGGTTTAATTGCAAGCAACGCGAAGAACCTTACCAGGTC	944		
Query 898	TTGACTTCCTCTGAC-ACCCTAAAAATAGGGCTTCCCCTTCGGGG	941		
Sbjct 945	TTGACATCCTCTGACAACCCTAGAGATAGGGCTTCCCCTTCGGGG	989		



Enterococcus mundtii KR085796.1

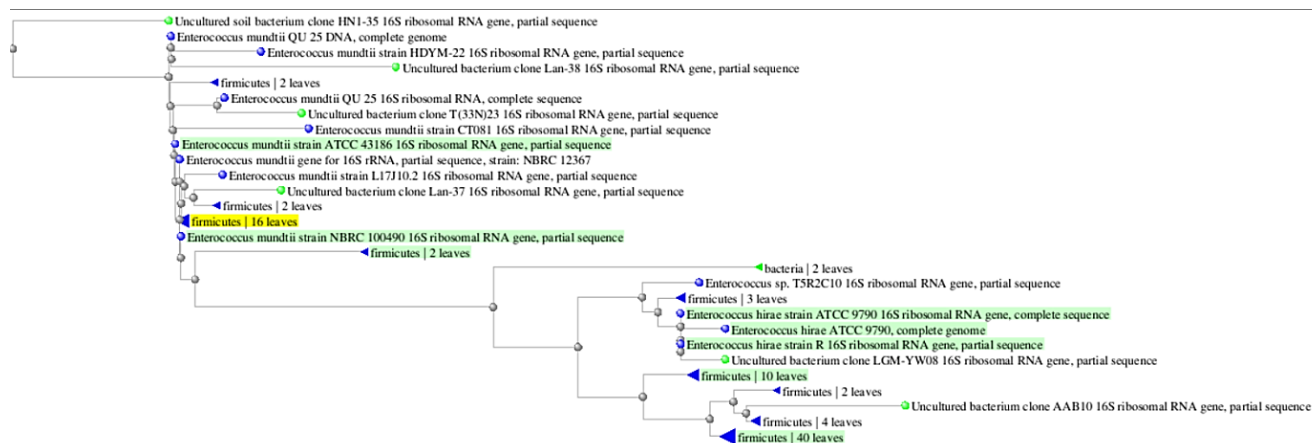
Enterococcus mundtii strain IHBB 9250 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KR085796.1](#) Length: 1518 Number of Matches: 1

Range 1: 81 to 1033 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1674 bits(1856)	0.0	944/953(99%)	1/953(0%)	Plus/Plus
Query 1	CCGGGAAAAGAGGAGTGGCGAACGGGTGAGTAACACGTGGGTAACTGCCCATCAGAAGG	60		
Sbjct 81		140		
Query 61	GGATAACACTTTGGAACAGGTGCTAATACCGTATAACAATCGAAACCGCATGGTTTCGTT	120		
Sbjct 141		200		
Query 121	TTGAAAGGCGCTTTACGGTGCCGCTGATGGATGGACCCGCGGTGCATTAGCTAGTTGGTG	180		
Sbjct 201		260		
Query 181	AGGTAACGGCTCACCAAGGCCACGATGCATAGCCGACCTGAGAGGGTGATCGGCCACATT	240		
Sbjct 261		320		
Query 241	GGGACTGAGACACGGCCCAAACCTCTACGGGAGGCAGCAGTAGGGAATCTTCGGCAATGG	300		
Sbjct 321		380		
Query 301	ACGAAAGTCTGACCGAGCAACGCCGCGTGAGTGAAGAAGGTTTTTCGGATCGTAAACTCT	360		
Sbjct 381		440		
Query 361	GTTGTTAGAGAAGAACAAGGGTGAGAGTAAGTGTTCACCCCTTGACGGTATCTAACCAGA	420		
Sbjct 441		500		
Query 421	AAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCG	480		
Sbjct 501		560		
Query 481	GATTTATTGGGCGTAAAGCGAGCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCGG	540		
Sbjct 561		620		
Query 541	CTCAACCGGGGAGGGTCATTGGAACCTGGGAGACTTGAGTGCAGAAGAGGAGAGTGGAAT	600		
Sbjct 621		680		
Query 601	TCCATGTGTAGCGGTGAAATGCGTAGATATATGGAGGAACACCAAGTGGCGAAGGCGGCTC	660		
Sbjct 681		740		
Query 661	TCTGGTCTGTAACTGACGCTGAGGCTCGAAAGCGTGGGGAGCAAACAGGATTAGATACCC	720		
Sbjct 741		800		
Query 721	TGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTGGAGGGTTTCCGCCCTTCAGTGC	780		
Sbjct 801		860		
Query 781	TGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGACCGCAAGGTTGAAACTCAAAG	840		
Sbjct 861		920		
Query 841	GAATTGACGGGGGCGGCACAAGCGGGGAGCATGGGGTTTAATTCTGAAGCAACGCGAAA	900		
Sbjct 921		980		
Query 901	AACCTTACCA-GTCTTGACATCCTTTGACCCCTCTaaaaaaaaaGCTTCCCCT	952		
Sbjct 981		1033		



Enterococcus hirae KT261200.1

Enterococcus hirae strain RCB988 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT261200.1](#) Length: 1437 Number of Matches: 1

Range 1: 59 to 965 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1546 bits(1714)	0.0	892/910(98%)	4/910(0%)	Plus/Plus
Query 1	AGTAACACGTGGGTAACCTGCCCATCAGAAGGGGATAACACTTGGAAACAGGTGCTAATA	60		
Sbjct 59	AGTAACACGTGGGTAACCTGCCCATCAGAAGGGGATAACACTTGGAAACAGGTGCTAATA	118		
Query 61	CCGTATAACAATCGAAACCGCATGGTTTGTATTTGAAAGGCGCTTTCGGGTGTCGCTGAT	120		
Sbjct 119	CCGTATAACAATCGAAACCGCATGGTTTGTATTTGAAAGGCGCTTTCGGGTGTCGCTGAT	178		
Query 121	GGATGGACCCGCGGTGCATTAGCTAGTTGGTGAGGTAACGGCTCACCAAGGCCACGATGC	180		
Sbjct 179	GGATGGACCCGCGGTGCATTAGCTAGTTGGTGAGGTAACGGCTCACCAAGGCCACGATGC	238		
Query 181	ATAGCCGACCTGAGAGGGTGATCGGCCACATTGGGACTGAGACACGGCCAAACTCCTAC	240		
Sbjct 239	ATAGCCGACCTGAGAGGGTGATCGGCCACATTGGGACTGAGACACGGCCAAACTCCTAC	298		
Query 241	GGGAGGCAGCAGTAGGGAATCTTCGGCAATGGACGAAAGTCTGACCGAGCAACGCCGCGT	300		
Sbjct 299	GGGAGGCAGCAGTAGGGAATCTTCGGCAATGGACGAAAGTCTGACCGAGCAACGCCGCGT	358		
Query 301	GAGTGAAGAAGGTTTTTCGGATCGTAAACTCTGTTGTTAGAGAAGAACAAGGATGAGAGT	360		
Sbjct 359	GAGTGAAGAAGGTTTTTCGGATCGTAAACTCTGTTGTTAGAGAAGAACAAGGATGAGAGT	418		
Query 361	AACTGTTTCATCCCTTGACGGTATCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGC	420		
Sbjct 419	AACTGTTTCATCCCTTGACGGTATCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGC	478		
Query 421	CGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGATTTATTGGGCGTAAAGCGAGCGCAGG	480		
Sbjct 479	CGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGATTTATTGGGCGTAAAGCGAGCGCAGG	538		
Query 481	CGGTTTCTTAAGTCTGATGTGAAAGCCCCCGGCTCAACCGGGGAGGGTCATTGGAAACTG	540		
Sbjct 539	CGGTTTCTTAAGTCTGATGTGAAAGCCCCCGGCTCAACCGGGGAGGGTCATTGGAAACTG	598		
Query 541	GGAGACTTGAGTGCAGAAGAGGAGAGTGGAATTCCATGTGTAGCGGTGAAATGCGTAGAT	600		
Sbjct 599	GGAGACTTGAGTGCAGAAGAGGAGAGTGGAATTCCATGTGTAGCGGTGAAATGCGTAGAT	658		
Query 601	ATATGGAGGAACACCAGTGGCGAAGGCGGCTCTCTGGTCTGTAACGTACGCTGAGGCTCG	660		
Sbjct 659	ATATGGAGGAACACCAGTGGCGAAGGCGGCTCTCTGGTCTGTAACGTACGCTGAGGCTCG	718		
Query 661	AAAGCGTGGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGT	720		
Sbjct 719	AAAGCGTGGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGT	778		
Query 721	GCTAAGTGTTGGAGGGTTTCCGCCCTTCAGTGCTGCAGCTAACGCATTAAACACTCCGCC	780		
Sbjct 779	GCTAAGTGTTGGAGGGTTTCCGCCCTTCAGTGCTGCAGCTAACGCATTAAAGCACTCCGCC	838		
Query 781	TGGGGAGTACAACCGCAAGGTTGAAACTCAAAGGAATTGACGGGGGGCCGCACAAGCGGT	840		
Sbjct 839	TGGGGAGTACGACCGCAAGGTTGAAACTCAAAGGAATTGACGGGGGGCCGCACAAGCGGT	898		
Query 841	GGGAACCTGTGGTTTAAATTTCAAACCAACCCaaaaaaCCTTACCAGG-CTTGAATTCCT	899		
Sbjct 899	-GGAGCATGTGGTTTAA-TTCGAAGCAACGC-GAAGAACCTTACCAGGTCTTGACATCCT	955		
Query 900	TTGACCCCTC 909			
Sbjct 956	TTGACCACTC 965			



Bacillus licheniformis DQ071560.1

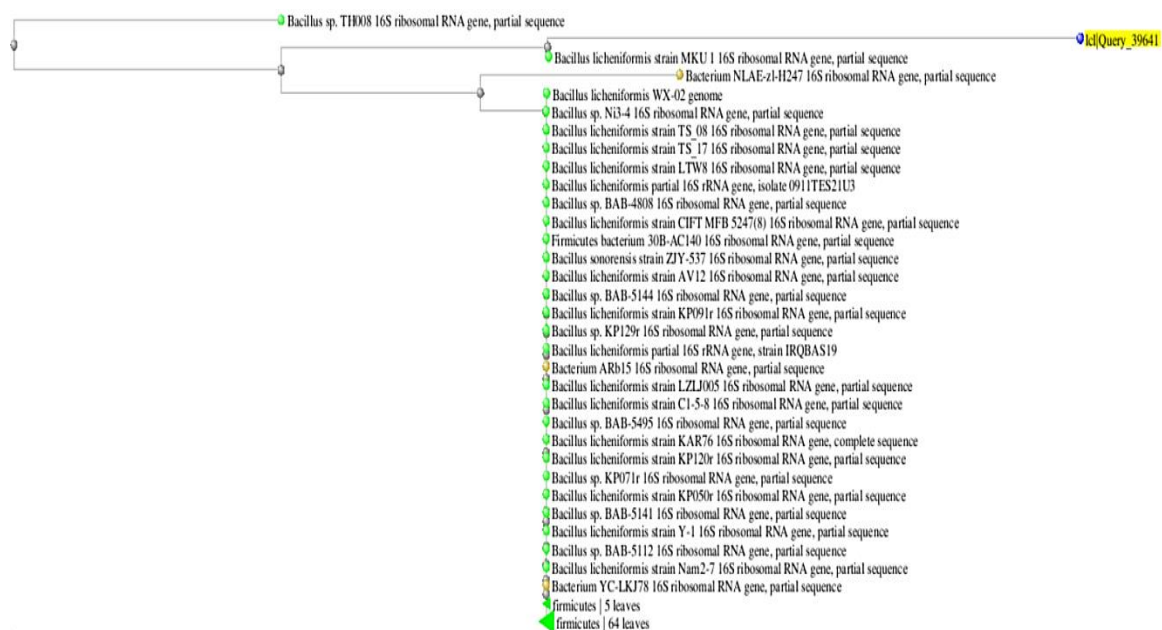
Bacillus licheniformis strain MKU 1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|DQ071560.1|](#) Length: 1430 Number of Matches: 1

Range 1: 431 to 1393 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1703 bits(1888)	0.0	958/963(99%)	3/963(0%)	Plus/Minus
Query 1	TTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCG	60		
Sbjct 1393	TTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCG	1334		
Query 61	GCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGA	120		
Sbjct 1333	GCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGA	1274		
Query 121	TCCGAAC TGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTT	180		
Sbjct 1273	TCCGAAC TGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTT	1214		
Query 181	CTGCCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATC	240		
Sbjct 1213	CTGCCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATC	1154		
Query 241	CCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCA	300		
Sbjct 1153	CCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCA	1094		
Query 301	ACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTG	360		
Sbjct 1093	ACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTG	1034		
Query 361	ACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTT	420		
Sbjct 1033	ACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTT	974		
Query 421	GTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCT	480		
Sbjct 973	GTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCT	914		
Query 481	CCACCGCTTGTGCGGGCCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCC	540		
Sbjct 913	CCACCGCTTGTGCGGGCCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCC	854		
Query 541	CAGGCGGAGTGCTTAATGCGTTTGTCTGCAGCACTAAAGGGCGGAAACCCCTAACACTTA	600		
Sbjct 853	CAGGCGGAGTGCTTAATGCGTTTGTCTGCAGCACTAAAGGGCGGAAACCCCTAACACTTA	794		
Query 601	GCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACGCTT	660		
Sbjct 793	GCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACGCTT	734		
Query 661	TCGCGCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACA	720		
Sbjct 733	TCGCGCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACA	674		
Query 721	TCTCTACGCATTTACCGCTACACGTGGAATTCCACTCTCCTCTTCTGCACTCAAGTTCC	780		
Sbjct 673	TCTCTACGCATTTACCGCTACACGTGGAATTCCACTCTCCTCTTCTGCACTCAAGTTCC	614		
Query 781	CCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTAAAAAACC	840		
Sbjct 613	CCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTAAAAAACC	554		
Query 841	GCCTGCGCGCGCTTTACGCCCAATAATTCCGGA-AACGCTTGCCCCCTACGTATTACCGC	899		
Sbjct 553	GCCTGCGCGCGCTTTACGCCCAATAATTCCGGA-AACGCTTGCCCCCTACGTATTACCGC	494		
Query 900	GGCTGCTGGCACGTAGTTAGCCGGGCTTTCTGGTTAGGTACCGTC-AGGTACC-CCCTA	957		
Sbjct 493	GGCTGCTGGCACGTAGTTAGCCGGGCTTTCTGGTTAGGTACCGTCAAGGTACCGCCCTA	434		
Query 958	TTC 960			
Sbjct 433	TTC 431			



Lysinibacillus fusiformis KP872952.1

Lysinibacillus fusiformis strain XKS50.1 16S ribosomal RNA gene, partial sequence

Sequence ID: [KP872952.1](#) Length: 1450 Number of Matches: 1

Range 1: 1 to 1450 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
2678 bits(1450)	0.0	1450/1450(100%)	0/1450(0%)	Plus/Plus
Query 1	CCCCGGGGGCTCTATACATGCAAGTCGAGCGAACAGAAAAGGAGCTTGCTCCTTTGACGT			60
Sbjct 1	CCCCGGGGGCTCTATACATGCAAGTCGAGCGAACAGAAAAGGAGCTTGCTCCTTTGACGT			60
Query 61	TAGCGGCGGACGGGTGAGTAACACGTGGGCAACCTACCCTATAGTTGGGATAACTCCGG			120
Sbjct 61	TAGCGGCGGACGGGTGAGTAACACGTGGGCAACCTACCCTATAGTTGGGATAACTCCGG			120
Query 121	GAAACCGGGGCTAATACCGAATAATCTCTTTTGCTTCATGGTGAAAGACTGAAAGACGGT			180
Sbjct 121	GAAACCGGGGCTAATACCGAATAATCTCTTTTGCTTCATGGTGAAAGACTGAAAGACGGT			180
Query 181	TTCGGCTGTCTGCTATAGGATGGGCGCGCGGCGCATTAGCTAGTTGGTGAGGTAACGGCTC			240
Sbjct 181	TTCGGCTGTCTGCTATAGGATGGGCGCGCGGCGCATTAGCTAGTTGGTGAGGTAACGGCTC			240
Query 241	ACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACA			300
Sbjct 241	ACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACA			300
Query 301	CGGCCACAGACTCCTACGGGAGGCAGCAGTAGGGAATCTCCACAATGGGCGAAAGCCTGA			360
Sbjct 301	CGGCCACAGACTCCTACGGGAGGCAGCAGTAGGGAATCTCCACAATGGGCGAAAGCCTGA			360
Query 361	TGGAGCAACGCGCGTGTAGTGAAGAAGGTTTTCGGATCGTAAAACTCTGTTGTAAGGGAA			420
Sbjct 361	TGGAGCAACGCGCGTGTAGTGAAGAAGGTTTTCGGATCGTAAAACTCTGTTGTAAGGGAA			420
Query 421	GAACAAGTACAGTAGTAAGTGGCTGTACCTTGACGGTACCTTATTAGAAAGCCACGGCTA			480
Sbjct 421	GAACAAGTACAGTAGTAAGTGGCTGTACCTTGACGGTACCTTATTAGAAAGCCACGGCTA			480
Query 481	ACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGC			540
Sbjct 481	ACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGC			540
Query 541	GTAAAGCGCGCGCAGGCGGTCCTTTAAGTCTGATGTGAAAGCCCACGGCTCAACCGTGGA			600
Sbjct 541	GTAAAGCGCGCGCAGGCGGTCCTTTAAGTCTGATGTGAAAGCCCACGGCTCAACCGTGGA			600
Query 601	GGGTATTGGAAACTGGGGGACTTGAGTGCAGAAGAGGAAAGTGGAAATCCAAAGTGTAGC			660
Sbjct 601	GGGTATTGGAAACTGGGGGACTTGAGTGCAGAAGAGGAAAGTGGAAATCCAAAGTGTAGC			660
Query 661	GGTGAAATGCGTAGAGATTGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTCTGTAA			720
Sbjct 661	GGTGAAATGCGTAGAGATTGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTCTGTAA			720
Query 721	CTGACGCTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACG			780
Sbjct 721	CTGACGCTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACG			780
Query 781	CCGTAAACGATGAGTGCTAAGTGTAGGGGGTTTCCGCCCTTAGTGCTGCAGCTAACGC			840
Sbjct 781	CCGTAAACGATGAGTGCTAAGTGTAGGGGGTTTCCGCCCTTAGTGCTGCAGCTAACGC			840
Query 841	ATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGG			900
Sbjct 841	ATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGG			900
Query 901	GCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACCTTACCAGG			960
Sbjct 901	GCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACCTTACCAGG			960
Query 961	TCTTGACATCCCGTTGACCACTGTAGAGATATAGTTTCCCTTCGGGGGCAACGGTGACA			1020
Sbjct 961	TCTTGACATCCCGTTGACCACTGTAGAGATATAGTTTCCCTTCGGGGGCAACGGTGACA			1020
Query 1021	GGTGGTGATGGTTGTCTGCTCAGCTCGTGTCTGAGATGTTGGGTTAAGTCCCGCAACGAG			1080
Sbjct 1021	GGTGGTGATGGTTGTCTGCTCAGCTCGTGTCTGAGATGTTGGGTTAAGTCCCGCAACGAG			1080
Query 1081	CGCAACCCTTGATCTTAGTTGCCATCATTTAGTTGGGCACTCTAAGGTGACTGCCGGTGA			1140
Sbjct 1081	CGCAACCCTTGATCTTAGTTGCCATCATTTAGTTGGGCACTCTAAGGTGACTGCCGGTGA			1140
Query 1141	CAAACCGGAGGAAGGTGGGGATGACGTCAAATCATCATGCCCCCTTATGACCTGGGCTACA			1200
Sbjct 1141	CAAACCGGAGGAAGGTGGGGATGACGTCAAATCATCATGCCCCCTTATGACCTGGGCTACA			1200
Query 1201	CACGTGCTACAATGGACGATACAAACGGTTGCCAACTCGCGAGAGGGAGCTAATCCGATA			1260
Sbjct 1201	CACGTGCTACAATGGACGATACAAACGGTTGCCAACTCGCGAGAGGGAGCTAATCCGATA			1260
Query 1261	AAGTCGTTCTCAGTTCGGATTGTAGGCTGCAACTCGCCTACATGAAGCCGGAATCGCTAG			1320
Sbjct 1261	AAGTCGTTCTCAGTTCGGATTGTAGGCTGCAACTCGCCTACATGAAGCCGGAATCGCTAG			1320
Query 1321	TAATCGCGGATCAGCATGCCGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTC			1380
Sbjct 1321	TAATCGCGGATCAGCATGCCGCGGTGAATACGTTCCCGGGCCTTGTACACACCGCCCGTC			1380
Query 1381	ACACCACGAGAGTTTGTAAACACCCGAAGTCGGTGAGGTAACTTTGGAGCCAGCCGCCGA			1440
Sbjct 1381	ACACCACGAGAGTTTGTAAACACCCGAAGTCGGTGAGGTAACTTTGGAGCCAGCCGCCGA			1440
Query 1441	AGGTGATGAT 1450			
Sbjct 1441	AGGTGATGAT 1450			

❖ kti/Query_107117

❖ Bacterium BCL13 16S ribosomal RNA gene, partial sequence

0.0001

❖ Bacillaceae bacterium SE17 16S ribosomal RNA gene, partial sequence
❖ Bacillaceae bacterium SE41 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain GT18 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sphaericus strain BL1 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain BL2 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sphaericus strain BL7 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. PZ_7 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. enrichment culture clone MJJ-11 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain R3 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. F8 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain TL 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain Ulm26 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain N169 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain N139 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain LE06 16S ribosomal RNA gene, partial sequence
❖ Bacillales bacterium Cul_0304 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. JN09 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. SSI.22 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain 44A 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain L13 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain L106 16S ribosomal RNA gene, partial sequence
❖ Geobacillus stearothermophilus strain KS141 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus borenitolerans clone C-B-L6 16S ribosomal RNA gene, partial sequence
❖ Firmicutes bacterium K17 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. Y20 16S ribosomal RNA gene, complete sequence
❖ Lysinibacillus macroides strain fwzyl 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus macroides strain fwzyl82 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis partial 16S rRNA gene, strain S5
❖ Lysinibacillus macroides partial 16S rRNA gene, isolate AVSI
❖ Proteus mirabilis strain BCr 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. W1.10-228 16S ribosomal RNA gene, complete sequence
❖ Lysinibacillus sp. S-2 16S ribosomal RNA gene, partial sequence
❖ Uncultured Bacillus sp. clone XT58 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain 164B (BPI) 16S ribosomal RNA gene, partial sequence
❖ Uncultured bacterium gene for 16S ribosomal RNA, partial sequence, clone: 20JB1
❖ Lysinibacillus fusiformis strain S-1 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. XJJC-134-IRF1 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. BAB-2942 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. BAB-637 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain PMM3 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sphaericus strain BI-CDA 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain BD3 16S ribosomal RNA gene, partial sequence
❖ Bacterium BCL19 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. hb47 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. hb56 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. hb60 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. LS-065 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus macroides isolate Y59 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain 3 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain 4 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus macroides strain M067 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain D 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. 6BK6Y10 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sphaericus strain RNB6 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. CZGRY2 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. CZGRY3 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. CZGRY5 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. CZGRY11 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. CZGRY13 16S ribosomal RNA gene, partial sequence
❖ Enterobacter sp. CZGRY7 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. CZGRY8 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus macroides strain W526 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain 14AQ21 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain 37POZ32 16S ribosomal RNA gene, partial sequence
❖ Bacillus sp. TGS 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus partial 16S rRNA gene, isolate KWW 55
❖ Lysinibacillus xylanilyticus partial 16S rRNA gene, isolate KWW 56
❖ Lysinibacillus xylanilyticus partial 16S rRNA gene, isolate KWW 64
❖ Lysinibacillus xylanilyticus partial 16S rRNA gene, isolate KWW 151
❖ Lysinibacillus xylanilyticus partial 16S rRNA gene, isolate KSW 19
❖ Lysinibacillus xylanilyticus partial 16S rRNA gene, isolate KSW 23
❖ Lysinibacillus sphaericus strain Xyn-1 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sphaericus strain DZS1 16S ribosomal RNA gene, partial sequence
❖ Bacterium BCL11 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain D27 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus macroides strain Y-4B 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. BAB-4376 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sphaericus strain YH4 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. 3HIX 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain BCH540 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus xylanilyticus strain AS 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus sp. DB14380 16S ribosomal RNA gene, partial sequence
❖ Uncultured Bacillus sp. gene for 16S rRNA, partial sequence, clone: CLZX151
❖ Lysinibacillus sp. Vx42 partial 16S rRNA gene, strain Vx42
❖ Lysinibacillus fusiformis strain KAR73 16S ribosomal RNA gene, complete sequence
❖ Lysinibacillus sp. FIAT-22044 16S ribosomal RNA gene, partial sequence
❖ Lysinibacillus fusiformis strain SBB30 16S ribosomal RNA gene, partial sequence
❖ Uncultured bacterium clone LXI-B74 16S ribosomal RNA gene, partial sequence
❖ Firmicutes | 8 leaves

Aneurinibacillus migulanus (Bacillus brevis) NR_113764.1

Aneurinibacillus migulanus strain NBRC 15520 16S ribosomal RNA gene, partial sequence

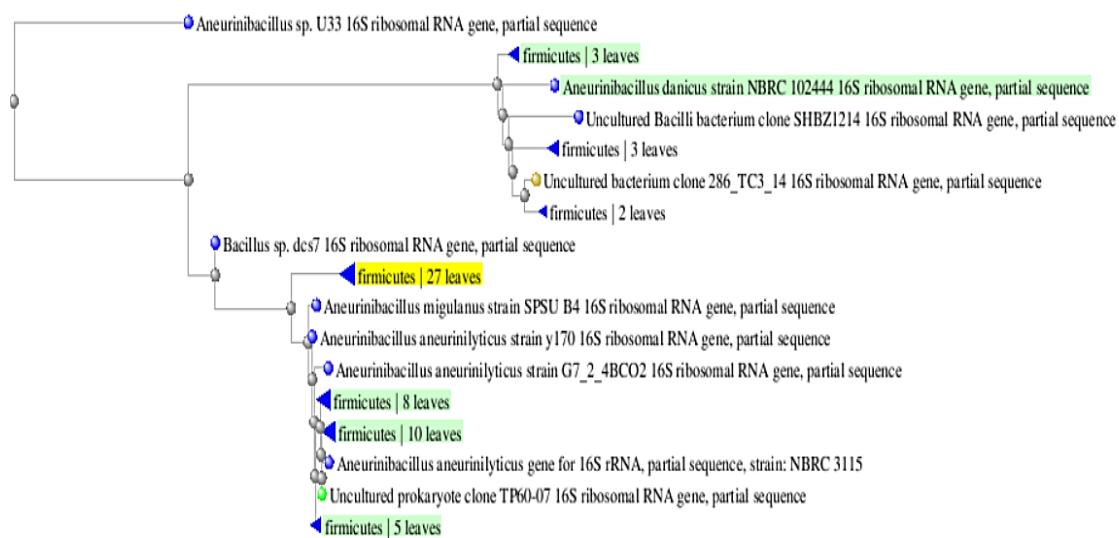
Sequence ID: [ref|NR_113764.1|](#) Length: 1464 Number of Matches: 1

[▶ See 1 more title\(s\)](#)

Range 1: 28 to 1441 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
2531 bits(2806)	0.0	1411/1414(99%)	2/1414(0%)	Plus/Plus	
Query 18		ATGCA-GTCGAGCGGACCAATGAAGAGCTTGCTCTTCGGCGGTTAGCGGCGGACGGGTGA			76
Sbjct 28		ATGCAAGTCGAGCGGACCAATGAAGAGCTTGCTCTTCGGCGGTTAGCGGCGGACGGGTGA			87
Query 77		GTAACACGTAGGCAACCTGCCTGTACGACTGGGATAACTCCGGGAAACCGGAGCTAATAC			136
Sbjct 88		GTAACACGTAGGCAACCTGCCTGTACGACTGGGATAACTCCGGGAAACCGGAGCTAATAC			147
Query 137		CGGATACTTCTTTTCAGACCGCATGGTCTGAAAGGGAAAGACCTTTGGTCACGTACAGATG			196
Sbjct 148		CGGATACTTCTTTTCAGACCGCATGGTCTGAAAGGGAAAGACNTTTGGTCACGTACAGATG			207
Query 197		GGCCTGCGGCGCATTAGCTAGTTGGTGGGGTAACGGCCTACCAAGCGGACGATGCGTAGC			256
Sbjct 208		GGCCTGCGGCGCATTAGCTAGTTGGTGGGGTAACGGCCTACCAAGCGGACGATGCGTAGC			267
Query 257		CGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAG			316
Sbjct 268		CGACCTGAGAGGGTGATCGGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAG			327
Query 317		GCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGGAGCAACGCCGCGTGAACG			376
Sbjct 328		GCAGCAGTAGGGAATCTTCCGCAATGGACGAAAGTCTGACGGAGCAACGCCGCGTGAACG			387
Query 377		ATGAAGGTTTTTCGGATCGTAAAGTTCTGTTGTTAGGGAAGAACC GCCGGGATGACCTCCC			436
Sbjct 388		ATGAAGGTTTTTCGGATCGTAAAGTTCTGTTGTTAGGGAAGAACC GCCGGGATGACCTCCC			447
Query 437		GGTCTGACGGTACCTAACGAGAAAGCCCCGGCTAACTACGTGCCAGCAGCCGCGGTAATA			496
Sbjct 448		GGTCTGACGGTACCTAACGAGAAAGCCCCGGCTAACTACGTGCCAGCAGCCGCGGTAATA			507
Query 497		CGTAGGGGGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCGCAGGCGGCTTCTTA			556
Sbjct 508		CGTAGGGGGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGCGCGCGCAGGCGGCTTCTTA			567
Query 557		AGTCAGGTGTGAAAGCCACGGCTCAACCGTGGAGGGCCACTTGAAACTGGGAAGCTTGA			616
Sbjct 568		AGTCAGGTGTGAAAGCCACGGCTCAACCGTGGAGGGCCACTTGAAACTGGGAAGCTTGA			627
Query 617		GTGAGGAGAGGAGAGCGGAATTCCACGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGA			676
Sbjct 628		GTGAGGAGAGGAGAGCGGAATTCCACGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGA			687
Query 677		ACACCCGTGGCGAAGGCGGCTCTCTGGCCTGTAAGTACGCTGAGGCGCGAAAGCGTGGG			736
Sbjct 688		ACACCCGTGGCGAAGGCGGCTCTCTGGCCTGTAAGTACGCTGAGGCGCGAAAGCGTGGG			747
Query 737		GAGCGAACAGGATTAGATACCTGGTAGTCCACGCCGTAAACGTTGAGTGCTAGGTGTTG			796
Sbjct 748		GAGCGAACAGGATTAGATACCTGGTAGTCCACGCCGTAAACGTTGAGTGCTAGGTGTTG			807
Query 797		GGGACTCCAATCCTCAGTGCCGCGAGCTAACGCAATAAGCACTCCGCCTGGGGAGTACGGC			856
Sbjct 808		GGGACTCCAATCCTCAGTGCCGCGAGCTAACGCAATAAGCACTCCGCCTGGGGAGTACGGC			867
Query 857		CGCAAGGCTGAAACTCAAAGGAATTGACGGGGACCCGCACAAGCGGTGGAGCATGTGGTT			916
Sbjct 868		CGCAAGGCTGAAACTCAAAGGAATTGACGGGGACCCGCACAAGCGGTGGAGCATGTGGTT			927
Query 917		TAATTCGAAGCAACGCGAAGAACCTTACCAGGGCTTGACATCCCGCTGACCCCTCCTAGAG			976
Sbjct 928		TAATTCGAAGCAACGCGAAGAACCTTACCAGGGCTTGACATCCCGCTGACCCCTCCTAGAG			987
Query 977		ATAGGAGCTCTCTTCGGAGCAGCGGTGACAGGTGGTGCATGGTTGTCGTACGCTCGTGTG			1036
Sbjct 988		ATAGGAGCTCTCTTCGGAGCAGCGGTGACAGGTGGTGCATGGTTGTCGTACGCTCGTGTG			1047
Query 1037		GTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCCTTGTCCTTAGTTGCCAGCATTT			1096
Sbjct 1048		GTGAGATGTTGGGTTAAGTCCCGCAACGAGCGCAACCCCTTGTCCTTAGTTGCCAGCATTT			1107
Query 1097		AGTTGGGCACTCTAGGGAGACTGCCGTGACAAAGACGGAGGAAGGTGGGGATGACGTCAA			1156
Sbjct 1108		AGTTGGGCACTCTAGGGAGACTGCCGTGACAAAGACGGAGGAAGGTGGGGATGACGTCAA			1167
Query 1157		ATCATCATGCCCCCTTATGTCTGGGCTACACACGTGCTACAATGGATGGAACAACGGGCA			1216
Sbjct 1168		ATCATCATGCCCCCTTATGTCTGGGCTACACACGTGCTACAATGGATGGAACAACGGGCA			1227
Query 1217		GCCAACTCGCGAGAGTGCGCGAATCCCTTAAACCATTTCTCAGTTCGGATTGCAGGCTGC			1276
Sbjct 1228		GCCAACTCGCGAGAGTGCGCGAATCCCTTAAACCATTTCTCAGTTCGGATTGCAGGCTGC			1287
Query 1277		AACTCGCCTGCATGAAGCCGGAATCGCTAGTAATCGCGGATCAGCATGCCGCGGTGAATA			1336
Sbjct 1288		AACTCGCCTGCATGAAGCCGGAATCGCTAGTAATCGCGGATCAGCATGCCGCGGTGAATA			1347
Query 1337		CGTTCGCCGGTCTTGTACACACCGCCCGTACACACGAGAGTTTGCAACACCCGAAGTC			1396
Sbjct 1348		CGTTCGCCGGTCTTGTACACACCGCCCGTACACACGAGAGTTTGCAACACCCGAAGTC			1407
Query 1397		GGTGAGGTAACCGCAA-GAGCCAGCCGCCGAAGG	1429		
Sbjct 1408		GGTGAGGTAACCGCAAAGGAGCCAGCCGCCGAAGG	1441		



Bacillus subtilis EF488088.1

Bacillus subtilis strain QD9 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|EF488088.1](#) Length: 1412 Number of Matches: 1

Range 1: 366 to 1319 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1700 bits(1884)	0.0	950/954(99%)	1/954(0%)	Plus/Minus
Query 1	AAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCAT	60		
Sbjct 1319	AAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCAT	1260		
Query 61	GCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCG	120		
Sbjct 1259	GCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCG	1200		
Query 121	AACTGAGAACAGATTTGTGGGATTGGCTTAACCTCGCGGTTTCGCTGCCCTTTGTTCTGT	180		
Sbjct 1199	AACTGAGAACAGATTTGTGGGATTGGCTTAACCTCGCGGTTTCGCTGCCCTTTGTTCTGT	1140		
Query 181	CCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCA	240		
Sbjct 1139	CCATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCA	1080		
Query 241	CCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTA	300		
Sbjct 1079	CCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTA	1020		
Query 301	AGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGA	360		
Sbjct 1019	AGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGA	960		
Query 361	CAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGACGTCCTATCTCTAGGATTGTCA	420		
Sbjct 959	CAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGACGTCCTATCTCTAGGATTGTCA	900		
Query 421	GAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCAC	480		
Sbjct 899	GAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCAC	840		
Query 481	CGCTTGTGCGGGCCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGG	540		
Sbjct 839	CGCTTGTGCGGGCCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGG	780		
Query 541	CGGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCAC	600		
Sbjct 779	CGGAGTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCAC	720		
Query 601	TCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTCGCTCCCCACGCTTTTCGC	660		
Sbjct 719	TCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTCGCTCCCCACGCTTTTCGC	660		
Query 661	TCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACATCTC	720		
Sbjct 659	TCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACATCTC	600		
Query 721	TACGCATTTCACCGCTACACGTGGAATTCACACTCTCCTCTTCTGCACTCAAGTTCCCCAG	780		
Sbjct 599	TACGCATTTCACCGCTACACGTGGAATTCACACTCTCCTCTTCTGCACTCAAGTTCCCCAG	540		
Query 781	TTTCCAATGACCCCTCCCCGGTTGAGCCGGGGGCTTTACATCAAACCTAAAAAACCGCCT	840		
Sbjct 539	TTTCCAATGACCCCTCCCCGGTTGAGCCGGGGGCTTTACATCAGACTTAAGAAACCGCCT	480		
Query 841	GCGAGCCCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGGCT	900		
Sbjct 479	GCGAGCCCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGGCT	420		
Query 901	GCTGGCACGTAGTTAGCCGGGCTTTCTGGTTAGGTACCGTC-AGGTACCGCCCT	953		
Sbjct 419	GCTGGCACGTAGTTAGCCGGTCTTTCTGGTTAGGTACCGTCAAGGTACCGCCCT	366		

	<ul style="list-style-type: none"> Query_213911 Bacillus subtilis strain SG6, complete genome Bacillus subtilis strain TO-A, JTC, complete genome Bacillus amyloqueliciens strain SN-23 16S ribosomal RNA gene, partial sequence Bacillus amyloqueliciens strain SN-15 16S ribosomal RNA gene, partial sequence Bacillus sp. cua7 16S ribosomal RNA gene, partial sequence Bacillus sp. TC2 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain KPBa 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain MU S1 16S ribosomal RNA gene, partial sequence Bacillus subtilis gene for 16S ribosomal RNA, partial sequence, strain: KB2 Bacillus sp. KSRH14 partial 16S rRNA gene, strain KSRH14 Bacillus sp. KSRH11 partial 16S rRNA gene, strain KSRH11 Bacillus sp. KSRH13 partial 16S rRNA gene, strain KSRH13 Bacillus subtilis strain D1 16S ribosomal RNA gene, partial sequence Bacillus vallismortis strain DDM7 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain DDI 56 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain NJ1 16S ribosomal RNA gene, partial sequence Bacterium enrichment culture clone J11-16 16S ribosomal RNA gene, partial sequence Bacillus sp. BAB-4896 16S ribosomal RNA gene, partial sequence Bacillus sp. BAB-4886 16S ribosomal RNA gene, partial sequence Bacillus sp. BAB-4886 16S ribosomal RNA gene, partial sequence Bacillus sp. BAB-4886 16S ribosomal RNA gene, partial sequence Bacillus sp. KM-1 gene for 16S ribosomal DNA, partial sequence Bacillus subtilis strain VXR02 16S ribosomal RNA gene, partial sequence Bacillus sp. S113 16S ribosomal RNA gene, partial sequence Bacillus sp. T229 16S ribosomal RNA gene, partial sequence Bacillus sp. 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NXUSASBL06 16S ribosomal RNA gene, partial sequence Bacterium YC-LK-LKJ121 16S ribosomal RNA gene, partial sequence Bacterium YC-LK-LKJ44 16S ribosomal RNA gene, partial sequence Bacterium YC-LK-LKJ45 16S ribosomal RNA gene, partial sequence Bacterium YC-LK-LKJ43 16S ribosomal RNA gene, partial sequence Bacillus sp. BAB-4880 16S ribosomal RNA gene, partial sequence Bacterium Y2 16S ribosomal RNA gene, partial sequence Bacillus sp. ZH-42 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain Natrel 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain RW-401 16S ribosomal RNA gene, partial sequence Bacillus sp. ZLXH4 16S ribosomal RNA gene, partial sequence Bacillus sp. ZLXH3 16S ribosomal RNA gene, partial sequence Bacillus sp. ZLXH3 16S ribosomal RNA gene, partial sequence Bacillus sp. ZLXH2 16S ribosomal RNA gene, partial sequence Bacillus sp. JBS-8 16S ribosomal RNA gene, partial sequence Bacillus sp. JBP-21 16S ribosomal RNA gene, partial sequence Bacillus tequilensis strain BRJ6 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain MSEB 24 16S ribosomal RNA gene, complete sequence Bacillus subtilis strain MSEB 32 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain MSEB 71 16S ribosomal RNA gene, partial sequence Bacillus sonorensis strain HIB B 1252 16S ribosomal RNA gene, partial sequence Bacillus subtilis subsp. inaquosorum strain HIB B 7075 16S ribosomal RNA gene, partial sequence Bacillus subtilis strain Bp-1 16S ribosomal RNA, partial sequence Bacillus tequilensis strain MN33 16S ribosomal RNA gene, partial sequence Bacillus mojavensis strain MN47 16S ribosomal RNA gene, partial sequence Bacillus mojavensis strain MN43 16S ribosomal RNA gene, partial sequence Bacillus mojavensis strain MN42 16S ribosomal RNA gene, partial sequence Bacillus mojavensis strain MN29 16S ribosomal RNA gene, partial sequence Bacillus mojavensis strain MN26 16S ribosomal RNA gene, partial sequence Bacillus sp. 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SSKSD8 16S ribosomal RNA gene, partial sequence
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0.0001

8- The phylogenetic analysis of bacteria isolated from books and shelves in libraries and archive storerooms samples.

Pseudomonas jessenii

LN774645.1

Pseudomonas jessenii partial 16S rRNA gene, isolate 1111MAR14N4

Sequence ID: [emb|LN774645.1|](#) Length: 1191 Number of Matches: 1

Range 1: 1 to 1028 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

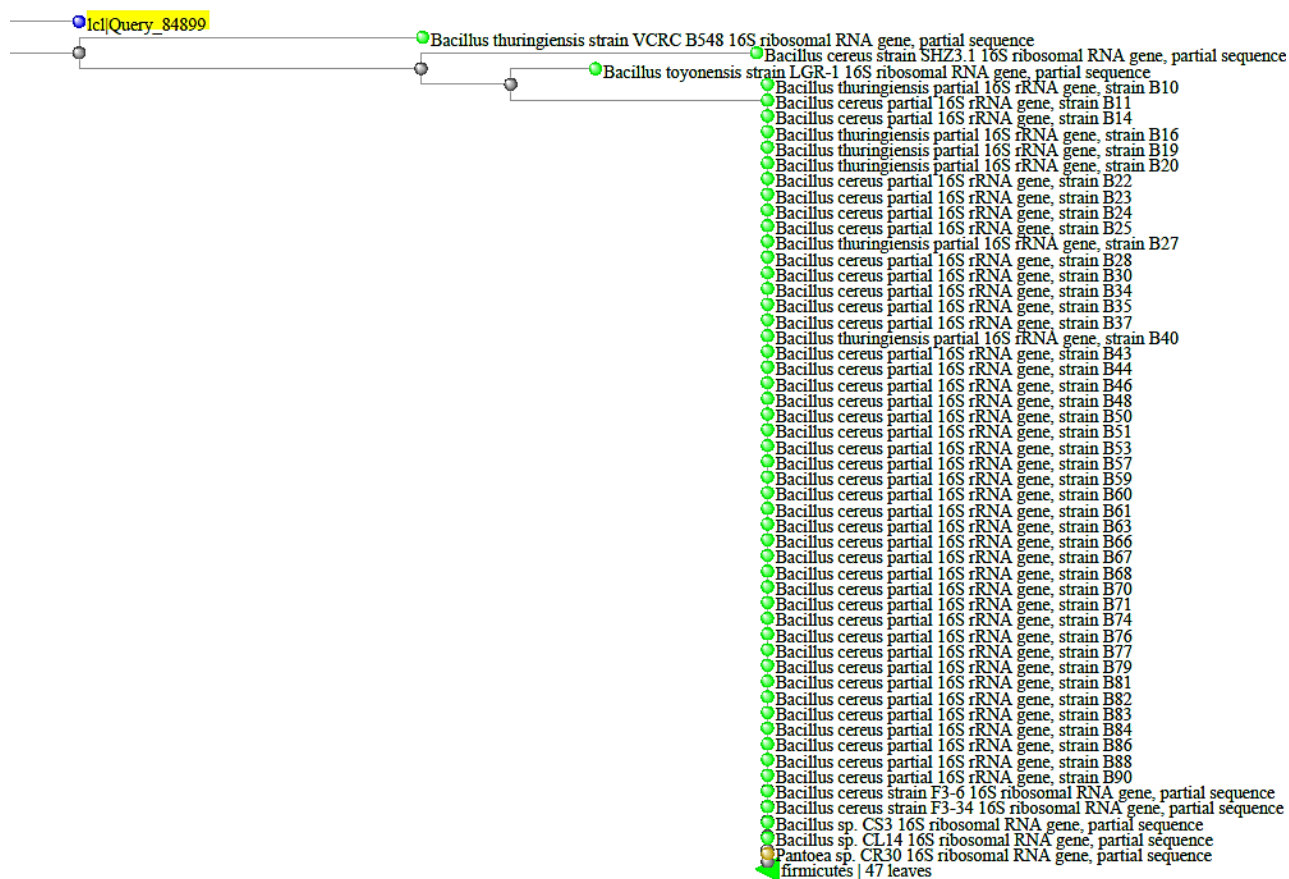
Score	Expect	Identities	Gaps	Strand	
1775 bits(1968)	0.0	1013/1028(99%)	3/1028(0%)	Plus/Plus	
Query	16	AATGCCTAGGAATCTGCCTATTAGTGGGGGACAACGTTTCGAAAGGAACGCTAATACCGC			75
Sbjct	1	AATGCCTAGGAATCTGCCTATTAGTGGGGGACAACGTTTCGAAAGGAACGCTAATACCGC			60
Query	76	ATACGTCCTACGGGAGAAAGCAGGGGACCTTCGGGCCTTGCCTAATAGATGAGCCTAGG			135
Sbjct	61	ATACGTCCTACGGGAGAAAGCAGGGGACCTTCGGGCCTTGCCTAATAGATGAGCCTAGG			120
Query	136	TCGGATTAGCTAGTTGGTGAGGTAATGGCTCACCAGGCGACGATCCGTAAGTGGTCTGA			195
Sbjct	121	TCGGATTAGCTAGTTGGTGAGGTAATGGCTCACCAGGCGACGATCCGTAAGTGGTCTGA			180
Query	196	GAGGATGATCAGTCACACTGGAAGTGAAGACGCGTCCAGACTCCTACGGGAGGCAGCAGT			255
Sbjct	181	GAGGATGATCAGTCACACTGGAAGTGAAGACGCGTCCAGACTCCTACGGGAGGCAGCAGT			240
Query	256	GGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCCATGCCGCGTGTGTGAAGAAGGT			315
Sbjct	241	GGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCCATGCCGCGTGTGTGAAGAAGGT			300
Query	316	CTTCGGATTGTAAAGCACTTTAAGTTGGGAGGAAGGGTCGTTGCCTAATACGTGACGGTT			375
Sbjct	301	CTTCGGATTGTAAAGCACTTTAAGTTGGGAGGAAGGGTCGTTGCCTAATACGTGACGGCT			360
Query	376	TTGACGTTACCGACAGAATAAGCACCGGCTAACTCTGTGCCAGCAGCCGCGGTAATACAG			435
Sbjct	361	TTGACGTTACCGACAGAATAAGCACCGGCTAACTCTGTGCCAGCAGCCGCGGTAATACAG			420
Query	436	AGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGCGCGCGTAGGTGGTTTGTAAAGT			495
Sbjct	421	AGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGCGCGCGTAGGTGGTTTGTAAAGT			480
Query	496	TGGATGTGAAATCCCCGGGCTCAACCTGGGAAGTGCATCCAAAAGTGGCAAGCTAGAGTA			555
Sbjct	481	TGGATGTGAAATCCCCGGGCTCAACCTGGGAAGTGCATCCAAAAGTGGCAAGCTAGAGTA			540
Query	556	CGGTAGAGGGTGGTGGGAATTTCTGTGTAGCGGTGAAATGCGTAGATATAGGAAGGAACA			615
Sbjct	541	CGGTAGAGGGTGGTGGGAATTTCTGTGTAGCGGTGAAATGCGTAGATATAGGAAGGAACA			600
Query	616	TCAGTGGCGAAGGCGACACCTGGACTGATACTGACACTGAGGTGCGAAAGCGTGGGGAG			675
Sbjct	601	TCAGTGGCGAAGGCGACACCTGGACTGATACTGACACTGAGGTGCGAAAGCGTGGGGAG			660
Query	676	C-AACAGGATTAGATACCCTGGTAGTCCACGCCGT-AACGATGTCAACTAGCCGTTGGGA			733
Sbjct	661	CAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGATGTCAACTAGCCGTTGGGA			720
Query	734	TCCTTGAGATCTTAGTGGCGCAGCTAACGCATTAAGTTGACCGCCTGGGGAGTACGGCCG			793
Sbjct	721	TCCTTGAGATCTTAGTGGCGCAGCTAACGCATTAAGTTGACCGCCTGGGGAGTACGGCCG			780
Query	794	CAAGGTTAAAAGTCAAATGAATTGACGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTA			853
Sbjct	781	CAAGGTTAAAAGTCAAATGAATTGACGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTA			840
Query	854	ATTCGAAGCAACGCGAAAAACCTTACCAGGCCTTGACATCCAGTGAACCTTCCAGAAATG			913
Sbjct	841	ATTCGAAGCAACGCGAAGAACCTTACCAGGCCTTGACATCCAGTGAACCTTCCAGAGATG			900
Query	914	GATTGGTGCCTTCGGGAACACTGAAACAGGGGCTGCATGGCTGTCGTGAGCTCGGGTCGG			973
Sbjct	901	GATTGGTGCCTTCGGGAACACTGAGACAGGTGCTGCATGGCTGTCGTGAGCTCGTGTCTGT			960
Query	974	GAAATGTTGGGTTAAGTCCCGTAACGAGCGCAACCCCTTGTCCTTAATTA-CAGCCCGTTA			1032
Sbjct	961	GAGATGTTGGGTTAAGTCCCGTAACGAGCGCAACCCCTTGTCCTTAGTTACCAGCACGTAA			1020
Query	1033	AGGGGGGC 1040			
Sbjct	1021	AGGTGGGC 1028			

Bacillus cereus KP192930.1

Bacillus cereus strain SHZ3.1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP192930.1](#) Length: 1419 Number of Matches: 1

Range 1: 334 to 1385		GenBank	Graphics			▼ Next Match ▲ Previous Match
Score	Expect	Identities	Gaps	Strand		
1824 bits(2022)	0.0	1039/1052(99%)	4/1052(0%)	Plus/Minus		
Query 1		CGGGTGTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGAACGTATTC				60
Sbjct 1385		CGGGTGTGTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGAACGTATTC				1326
Query 61		ACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGC				120
Sbjct 1325		ACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGC				1266
Query 121		CTACAATCCGAACGTGAGAACGGTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTC				180
Sbjct 1265		CTACAATCCGAACGTGAGAACGGTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTC				1206
Query 181		TTTGTACCGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTGAC				240
Sbjct 1205		TTTGTACCGTCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTGAC				1146
Query 241		GTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATG				300
Sbjct 1145		GTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATG				1086
Query 301		ATGGCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCAACATCTCACGACAC				360
Sbjct 1085		ATGGCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCAACATCTCACGACAC				1026
Query 361		GAGCTGACGACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCT				420
Sbjct 1025		GAGCTGACGACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCT				966
Query 421		AGGGTTTTTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCA				480
Sbjct 965		AGGGTTTTTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCA				906
Query 481		CATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGCCTTGC GGCCGT				540
Sbjct 905		CATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGCCTTGC GGCCGT				846
Query 541		ACTCCCCAGGCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGCGGAAACCCCTCTAA				600
Sbjct 845		ACTCCCCAGGCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGCGGAAACCCCTCTAA				786
Query 601		CACTTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCC				660
Sbjct 785		CACTTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCC				726
Query 661		ACGCTTTTCGCGCCTCAGTGTGAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTC				720
Sbjct 725		ACGCTTTTCGCGCCTCAGTGTGAGTTACAGACCAGAAAGTCGCCTTCGCCACTGGTGTTC				666
Query 721		TCCATATCTCTACGCATTTACCGCTACACATGGAATTCACCTTTCTCTTCTGCACTCA				780
Sbjct 665		TCCATATCTCTACGCATTTACCGCTACACATGGAATTCACCTTTCTCTTCTGCACTCA				606
Query 781		AGTCTCCAGTTTCCAATGACCCCTCCACGGTTGAGCCGTGGGCTTTCACATCAAACCTAA				840
Sbjct 605		AGTCTCCAGTTTCCAATGACCCCTCCACGGTTGAGCCGTGGGCTTTCACATCAGACTTAA				546
Query 841		AAAACCACCTGCGCGCGCTTTACGCCCAATAATCCGGATAACGCTTGCCACCTACGTAT				900
Sbjct 545		GAAACCACCTGCGCGCGCTTTACGCCCAATAATCCGGATAACGCTTGCCACCTACGTAT				486
Query 901		TACCGCGGCTGCTGGCACGTAGTTAGCCG-GGCTTTCTGGTTAGGTACCGTCAAGGGGCC				959
Sbjct 485		TACCGCGGCTGCTGGCACGTAGTTAGCCGTTGGCTTTCTGGTTAGGTACCGTCAAGGTGCC				426
Query 960		AGCTTATTCAACTAGCACTTGTCTTCCCTAAC-ACAAAATTTTACAACCCG-AAGCCTT				1017
Sbjct 425		AGCTTATTCAACTAGCACTTGTCTTCCCTAACAAACAAAATTTTACGACCCGAAAGCCTT				366
Query 1018		CCTCACTCCCCCGGCG-TGCTCCGTCCAAATT	1048			
Sbjct 365		CATCACTACGCGGCGTTGCTCCGTCAGAATT	334			



Bacillus altitudinis KT758615.1

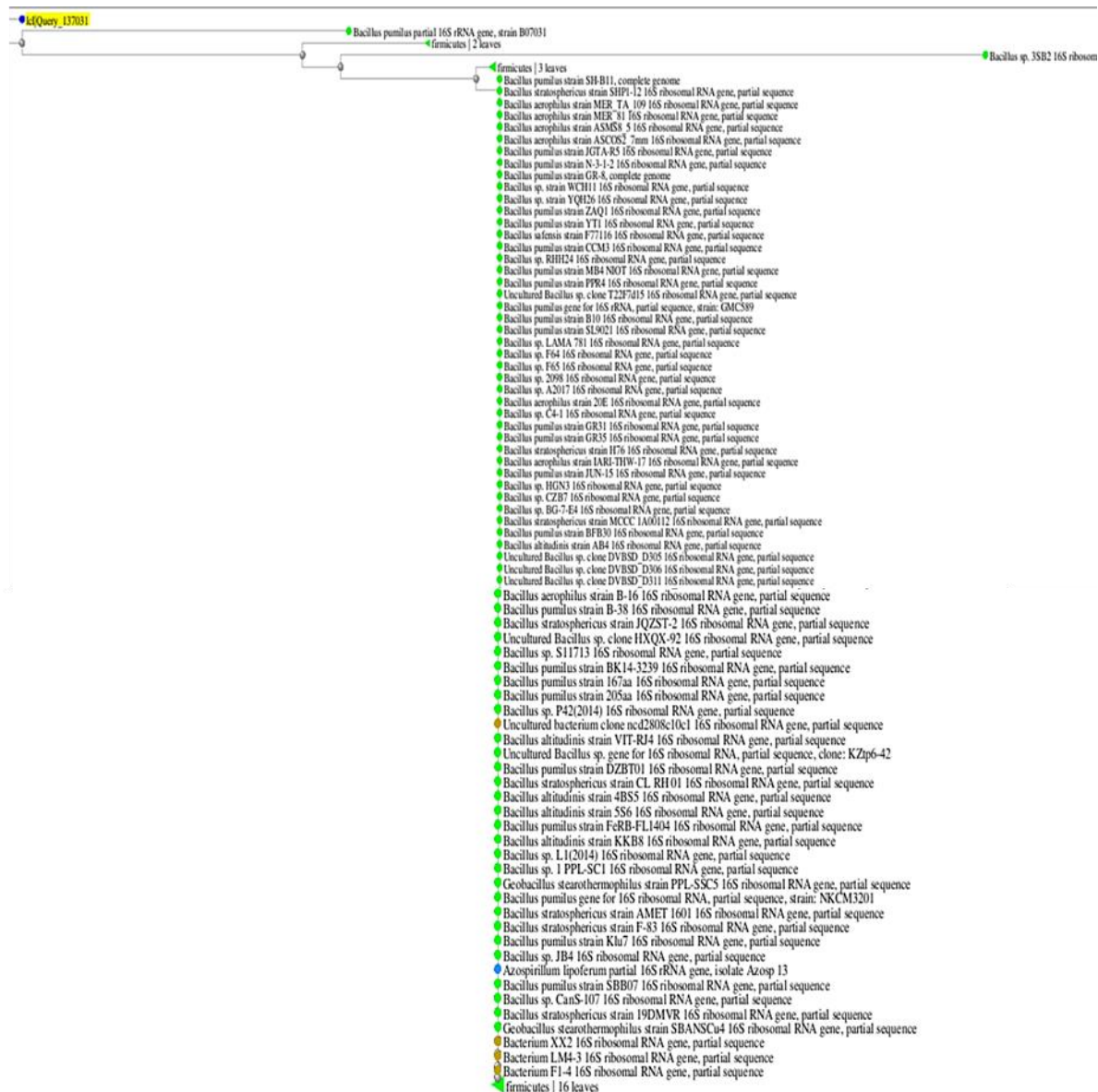
Bacillus altitudinis strain HQB822 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT758615.1](#) Length: 1111 Number of Matches: 1

Range 1: 27 to 1077 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1817 bits(2014)	0.0	1036/1052(98%)	3/1052(0%)	Plus/Plus
Query 1	CCGGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGAT	60		
Sbjct 27	CCGGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGAT	86		
Query 61	AACTCCGGGAAACCGGAGCTAATACCGGATAGTTCCTTGAACCGCATGGTTCAAGGATGA	120		
Sbjct 87	AACTCCGGGAAACCGGAGCTAATACCGGATAGTTCCTTGAACCGCATGGTTCAAGGATGA	146		
Query 121	AAGACGGTTTCGGCTGTCACTTAYAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGT	180		
Sbjct 147	AAGACGGTTTCGGCTGTCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGAGGT	206		
Query 181	AACGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGA	240		
Sbjct 207	AACGGCTCACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGGA	266		
Query 241	CTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGA	300		
Sbjct 267	CTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACGA	326		
Query 301	AAGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTG	360		
Sbjct 327	AAGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTG	386		
Query 361	TTAGGGAAGAACAAAGTGCAAGAGTAAGTGCCTTGACGGTACCTAACAGAAAGC	420		
Sbjct 387	TTAGGGAAGAACAAAGTGCAAGAGTAAGTGCCTTGACGGTACCTAACAGAAAGC	446		
Query 421	CACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAAT	480		
Sbjct 447	CACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAAT	506		
Query 481	TATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCA	540		
Sbjct 507	TATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCGGCTCA	566		
Query 541	ACCGGGGAGGGTCATTGGAAGTGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCCA	600		
Sbjct 567	ACCGGGGAGGGTCATTGGAAGTGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCCA	626		
Query 601	CGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAAGTGGCGAAGGCGACTCTCTG	660		
Sbjct 627	CGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAAGTGGCGAAGGCGACTCTCTG	686		
Query 661	GTCTGTAAGTACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT	720		
Sbjct 687	GTCTGTAAGTACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT	746		
Query 721	AGTCCACGCCGTAAACGATGAGTGCTAAGTGTAGGGGGTTTCCGCCCTTAGTGCTGCA	780		
Sbjct 747	AGTCCACGCCGTAAACGATGAGTGCTAAGTGTAGGGGGTTTCCGCCCTTAGTGCTGCA	806		
Query 781	GCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAAT	840		
Sbjct 807	GCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAAT	866		
Query 841	TGACGGGGGGCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAAAACC	900		
Sbjct 867	TGACGGGGGGCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACC	926		
Query 901	TTACCAGGTCTTGACATCCTCTGACACCCTAAAGATAGGGCTTTCCCTTC-GGGAAAGAG	959		
Sbjct 927	TTACCAGGTC-TGACATCCTCTGACACCCTAGAGATAGGGCTTTCCCTTCGGGGACAGAG	985		
Query 960	TGACAGGGGGGCATGGTTGTCTGCTGAGCTCGGGTCGGGAAATGTGGGTTAAGTCCCGCAC	1019		
Sbjct 986	TGACAGTGGTGATGGTTGTCTGCTGAGCTCTGGTCGTGAGATGTGGGTTAAGTCCCGCAC	1045		
Query 1020	GAGCGCACCCCTTGATCTA-TTGCCAGCATTTA	1050		
Sbjct 1046	GAGCGCAACCCTGATCTAGTTGCCAGCATTTA	1077		



Bacillus pumilus KP322017.1

Bacillus pumilus strain MK16 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP322017.1|](#) Length: 1469 Number of Matches: 1

Range 1: 64 to 1120 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1799 bits(1994)	0.0	1038/1057(98%)	6/1057(0%)	Plus/Plus
Query 1	CCCCGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGA	60		
Sbjct 64	CCCCGATGTTAGCGGCGGACGGGTGAGTAACACGTGGGTAACCTGCCTGTAAGACTGGGA	123		
Query 61	TAACTCCGGGAAACCGGAGCTAATACCGGATAGTTCCTTGAACCGCATGGTTCAAGGATG	120		
Sbjct 124	TAACTCCGGGAAACCGGAGCTAATACCGGATAGTTCCTTGAACCGCATGGTTCAAGGATG	183		
Query 121	AAAGACGGTTTTCGGCTGTCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGGGG	180		
Sbjct 184	AAAGACGGTTTTCGGCTGTCACTTACAGATGGACCCGCGGCGCATTAGCTAGTTGGTGGGG	243		
Query 181	TAATGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGG	240		
Sbjct 244	TAATGGCTCACCAAGGCGACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACTGGG	303		
Query 241	ACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACG	300		
Sbjct 304	ACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGGACG	363		
Query 301	AAAGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTT	360		
Sbjct 364	AAAGTCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTT	423		
Query 361	GTTAGGGAAGAACAAGTGCAGAGTAAGTCTCGCACCTTGACGGTACCTAACAGAAAAG	420		
Sbjct 424	GTTAGGGAAGAACAAGTGCAGAGTAAGTCTCGCACCTTGACGGTACCTAACAGAAAAG	483		
Query 421	CCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAA	480		
Sbjct 484	CCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAA	543		
Query 481	TTATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCCGGCTC	540		
Sbjct 544	TTATTGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCCCCGGCTC	603		
Query 541	AACCGGGGAGGGTCATTGGAACTGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCC	600		
Sbjct 604	AACCGGGGAGGGTCATTGGAACTGGGAACTTGAGTGCAGAAGAGGAGAGTGGAATTCC	663		
Query 601	ACGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCT	660		
Sbjct 664	ACGTGTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCT	723		
Query 661	GGTCTGTAACGTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCTTGG	720		
Sbjct 724	GGTCTGTAACGTGACGCTGAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCTTGG	783		
Query 721	TAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCTTAGTGCTGC	780		
Sbjct 784	TAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCTTAGTGCTGC	843		
Query 781	AGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAA	840		
Sbjct 844	AGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAA	903		
Query 841	TTGACGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTGCAAGCAACGCGAAAAAC	900		
Sbjct 904	TTGACGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTGCAAGCAACGCGAAGAAC	963		
Query 901	CTTACCA-GTCTTGACATCCTCTGAC-ACCCTAAAAATAGGGCTTTCCCTTC-GGGACAG	957		
Sbjct 964	CTTACCAGGTCTTGACATCCTCTGACAACCCTAGAGATAGGGCTTTCCCTTCGGGGACAG	1023		
Query 958	AGTGACAgggggggCATGGTTGTCGTCGCTCGGGTCGGGAAATGTTGGGTAA-TCCCC	1016		
Sbjct 1024	AGTGACAGTGGTGATGGTTGTCGTCAGCTCGTGTCTGAGATGTTGGGTAAAGTCCGC	1083		
Query 1017	CACGAGCGC-ACCCTTGATC-TAATTGCCAGCATCCA	1051		
Sbjct 1084	AACGAGCGCAACCCTTGATCTTAGTTGCCAGCATTCA	1120		



Bacillus stratosphericus KJ672335.1

Bacillus stratosphericus strain MUGA150 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ672335.1](#) Length: 1404 Number of Matches: 1

Range 1: 326 to 1371 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1781 bits(1974)	0.0	1026/1046(98%)	7/1046(0%)	Plus/Minus
Query 1	TCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTG	60		
Sbjct 1371	TCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTG	1312		
Query 61	ATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACT	120		
Sbjct 1311	ATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACT	1252		
Query 121	GAGAACAGATTTGTGGGATTGGCTAAACCTTGCGGTCTCGCAGCCCTTTGTTCTGTCCAT	180		
Sbjct 1251	GAGAACAGATTTGTGGGATTGGCTAAACCTTGCGGTCTCGCAGCCCTTTGTTCTGTCCAT	1192		
Query 181	TGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTT	240		
Sbjct 1191	TGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTT	1132		
Query 241	CCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGAT	300		
Sbjct 1131	CCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGAT	1072		
Query 301	CAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAAC	360		
Sbjct 1071	CAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAAC	1012		
Query 361	CATGCACCACCTGTCACTCTGTCCCGAAGGGAAAGTCTATCTCTAGGGTTGTGAGAGG	420		
Sbjct 1011	CATGCACCACCTGTCACTCTGTCCCGAAGGGAAAGCCCTATCTCTAGGGTTGTGAGAGG	952		
Query 421	ATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCT	480		
Sbjct 951	ATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCT	892		
Query 481	TGTGCGGGCCCCCGTCAATTCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGCGGA	540		
Sbjct 891	TGTGCGGGCCCCCGTCAATTCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGCGGA	832		
Query 541	GTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCAT	600		
Sbjct 831	GTGCTTAATGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCAT	772		
Query 601	CGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCGCTCCCCACGCTTTGCTCCT	660		
Sbjct 771	CGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCGCTCCCCACGCTTTGCTCCT	712		
Query 661	CAGCGTCAGTTACAGACCAGAGAGTCGCCCTTCGCCACTGGTGTTCCTCCACATCTCTACG	720		
Sbjct 711	CAGCGTCAGTTACAGACCAGAGAGTCGCCCTTCGCCACTGGTGTTCCTCCACATCTCTACG	652		
Query 721	CATTTACCGCTACACGTGGAATTCACCTCTCCTCTTCTGCACTCAAGTTTCCAGTTTC	780		
Sbjct 651	CATTTACCGCTACACGTGGAATTCACCTCTCCTCTTCTGCACTCAAGTTTCCAGTTTC	592		
Query 781	CAATGACCCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTAAAAAACCGCTGCGA	840		
Sbjct 591	CAATGACCCCTCCCCGGTTGAGCCGGGGGCTTTCACATCAGACTTAAGAAACCGCTGCGA	532		
Query 841	GCCCTTTACGCCCCGATAATTCCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCT	900		
Sbjct 531	GCCCTTTACGCCCCAATAATTCCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCT	472		
Query 901	GGCACGTATTTAGCCGTGGCTTTCTGGTTAGGTACCGTCAAGGGGCAAGCAGTTACTCTT	960		
Sbjct 471	GGCACGTAGTTAGCCGTGGCTTTCTGGTTAGGTACCGTCAAGGTGCAAGCAGTTACTCTT	412		
Query 961	GCACTTGTCTTCCCTA--CACAAAACCTTTACAATCC--AAAACCTTCATCCCTC-CGCGG	1016		
Sbjct 411	GCACTTGTCTTCCCTAACAACAGAGCTTTACGATCCGAAAACCTTCATCACTCAGCGG	352		
Query 1017	CGTTGCT-CGTCAAA--TTCCTCCAT	1039		
Sbjct 351	CGTTGCTCCGTCAGACTTTCGTCCAT	326		



Bacillus weithenstephanensis KC527665.1

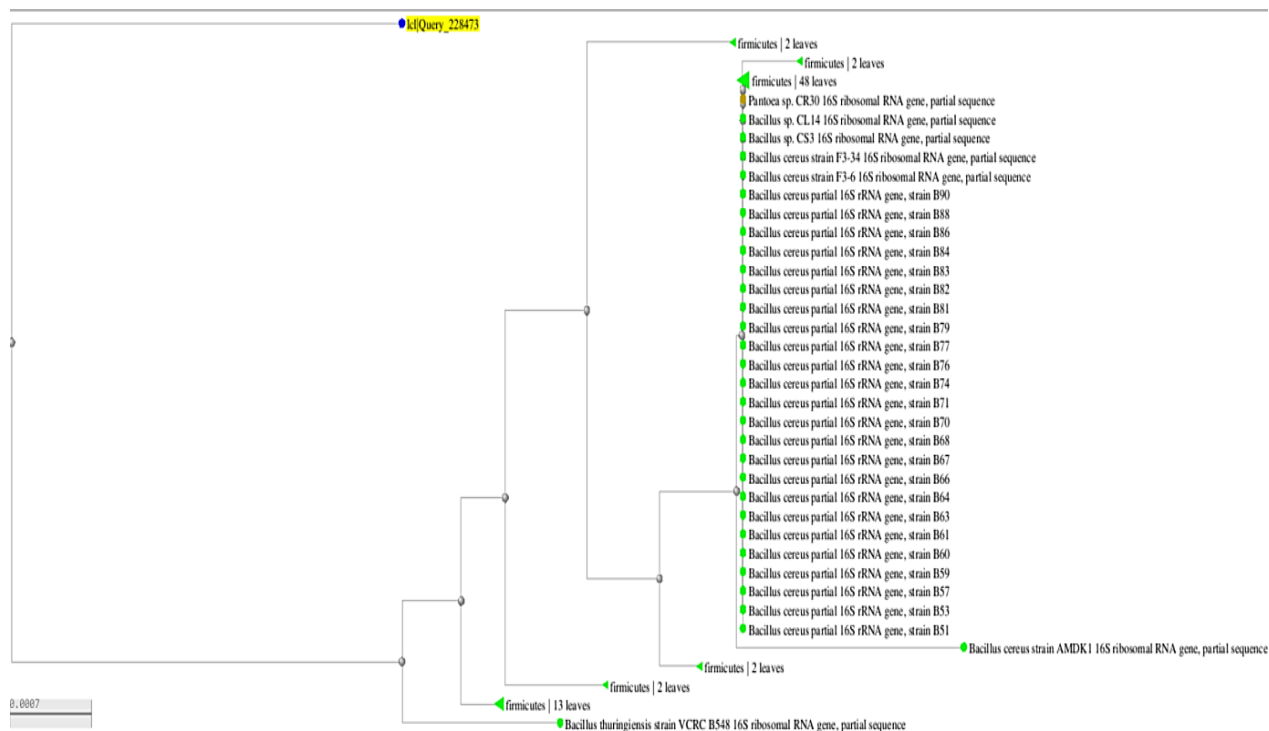
Bacillus weihenstephanensis strain A2-25c-6b 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KC527665.1|](#) Length: 1252 Number of Matches: 1

Range 1: 161 to 1197 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1750 bits(1940)	0.0	1016/1038(98%)	7/1038(0%)	Plus/Minus
Query 1	CAAACCTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCA	60		
Sbjct 1197	CAAACCTCTCG-GGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCA	1139		
Query 61	TGCTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACAATCC	120		
Sbjct 1138	TGCTGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACAATCC	1079		
Query 121	GAACCTGAGAACGGTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGTACCG	180		
Sbjct 1078	GAACCTGAGAACGGTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGTACCG	1019		
Query 181	TCCATTGTAGCACGTGTGTAGCCAGGTCTAAGGGGCATGATGATTTGACGTCATCCCC	240		
Sbjct 1018	TCCATTGTAGCACGTGTGTAGCCAGGTCTAAGGGGCATGATGATTTGACGTCATCCCC	959		
Query 241	ACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGCAACT	300		
Sbjct 958	ACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGCAACT	899		
Query 301	AAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACG	360		
Sbjct 898	AAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACG	839		
Query 361	ACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCTAGGGTTTTC	420		
Sbjct 838	ACAACCATGCACCACCTGTCACTCTGCTCCCGAAGGAGAAGCTCTATCTCTAGAGTTTTC	779		
Query 421	AGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTGAATTAACACCATGCTCCA	480		
Sbjct 778	AGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTGAATTAACACCATGCTCCA	719		
Query 481	CCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCCAG	540		
Sbjct 718	CCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCCAG	659		
Query 541	GCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCTCTAACACTTAGCA	600		
Sbjct 658	GCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCTCTAACACTTAGCA	599		
Query 601	CTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCCCACGCTTTTCG	660		
Sbjct 598	CTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCCCACGCTTTTCG	539		
Query 661	CGCCTCAGTGTGAGTTACAGACCAGAAAGTCGCTTCGCCACTGGTGTTCCTCCATATCT	720		
Sbjct 538	CGCCTCAGTGTGAGTTACAGACCAGAAAGTCGCTTCGCCACTGGTGTTCCTCCATATCT	479		
Query 721	CTACGCATTTACCGCTACACATGGAATTCACCTTTCCTCTTCTGCACTCAAGTCTCCCA	780		
Sbjct 478	CTACGCATTTACCGCTACACATGGAATTCACCTTTCCTCTTCTGCACTCAAGTCTCCCA	419		
Query 781	GTTTCCAATGACCCCTCCACGGTTGAGCCGTGGGCTTTACATCAAACCTTAA-AAACCACC	839		
Sbjct 418	GTTTCCAATGACCCCTCCACGGTTGAGCCGTGGGCTTTACATCAGACTTAAGAAACCACC	359		
Query 840	TGCGCGCGCTTTACGCCCAATAATTCCGGA-AACGCTTGCCACCTACGTATTACCGCGGC	898		
Sbjct 358	TGCGCGCGCTTTACGCCCAATAATTCCGGAATAACGCTTGCCACCTACGTATTACCGCGGC	299		
Query 899	TGCTGGCACGTAATTAGCCG-GGCTTTCTGGTTAGGTACCGTCAGGGGCCAGNTTATTCA	957		
Sbjct 298	TGCTGGCACGTAGTTAGCCGTGGCTTTCTGGTTAGGTACCGTCAGGTGCCAGCTTATTCA	239		
Query 958	ACTAGCATTTGTTCTTCCT-ACACAAATTTTAC-AACCGAAGCTTCATCACTC-CNCGCG	1014		
Sbjct 238	ACTAGCACTTGTTCCTTAACACAGAGTTTACGACCCGAAGCTTCATCACTCAGCGCGC	179		
Query 1015	TGCCCGTCAAAATTCCTC 1032			
Sbjct 178	TGCTCGTCAGACTTCGTC 161			



Acinetobacter lofwii KT387352.1

Acinetobacter lwoffii strain CL_102 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT387352.1|](#) Length: 1437 Number of Matches: 1

Range 1: 425 to 1356 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1640 bits(1818)	0.0	923/932(99%)	0/932(0%)	Plus/Minus
Query 1	GTACAAGGCCCGGGAACGTATTACCGCGGCATTCTGATCCGCGATTACTAGCGATTCCG	60		
Sbjct 1356	GTACAAGGCCCGGGAACGTATTACCGCGGCATTCTGATCCGCGATTACTAGCGATTCCG	1297		
Query 61	ACTTCATGGAGTCGAGTTGCAGACTCCAATCCGGACTACGATCGGCTTTTTGAGATTAGC	120		
Sbjct 1296	ACTTCATGGAGTCGAGTTGCAGACTCCAATCCGGACTACGATCGGCTTTTTGAGATTAGC	1237		
Query 121	ATCCTCTCGCGAGGTAGCAACCCTTTGTACCGACCATTGTAGCACGTGTGTAGCCCTGGT	180		
Sbjct 1236	ATCCTCTCGCGAGGTAGCAACCCTTTGTACCGACCATTGTAGCACGTGTGTAGCCCTGGT	1177		
Query 181	CGTAAGGGCCATGATGACTTGACGTCGTCCCGCCTTCTCCAGTTTGTCACTGGCAGTA	240		
Sbjct 1176	CGTAAGGGCCATGATGACTTGACGTCGTCCCGCCTTCTCCAGTTTGTCACTGGCAGTA	1117		
Query 241	TCCTTAAAGTTCCCGGCTTAACCCGCTGGCAAATAAGGAAAAGGGTTGCGCTCGTTGCGG	300		
Sbjct 1116	TCCTTAAAGTTCCCGGCTTAACCCGCTGGCAAATAAGGAAAAGGGTTGCGCTCGTTGCGG	1057		
Query 301	GACTTAACCCAACATCTCACGACACGAGCTGACGACAGCCATGCAGCACCTGTATGTAAG	360		
Sbjct 1056	GACTTAACCCAACATCTCACGACACGAGCTGACGACAGCCATGCAGCACCTGTATGTAAG	997		
Query 361	TTCCCGAAGGCACCAATCCATCTCTGGAAAGTTCTTACTATGTCAAGACCAGGTAAGGT	420		
Sbjct 996	TTCCCGAAGGCACCAATCCATCTCTGGAAAGTTCTTACTATGTCAAGACCAGGTAAGGT	937		
Query 421	CTTCGCGTTGCATCGAATTAAACACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTC	480		
Sbjct 936	CTTCGCGTTGCATCGAATTAAACACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTC	877		
Query 481	ATTTGAGTTTTAGTCTTGCGACCGTACTCCCCAGGCGGTCTACTTATCGCGTTAGCTGCG	540		
Sbjct 876	ATTTGAGTTTTAGTCTTGCGACCGTACTCCCCAGGCGGTCTACTTATCGCGTTAGCTGCG	817		
Query 541	CCACTAAAGCCTCAAAGGCCCAACGGCTAGTAGACATCGTTTACGGCATGGACTACCAG	600		
Sbjct 816	CCACTAAAGCCTCAAAGGCCCAACGGCTAGTAGACATCGTTTACGGCATGGACTACCAG	757		
Query 601	GGTATCTAATCCTGTTTGCTCCCCATGCTTTCGCACCTCAGTGTGAGTATTAGGCCAGAT	660		
Sbjct 756	GGTATCTAATCCTGTTTGCTCCCCATGCTTTCGCACCTCAGTGTGAGTATTAGGCCAGAT	697		
Query 661	GGCTGCCTTCGCCATCGGTATTCTCCAGATCTCTACGCATTTACCGCTACACCTGGAA	720		
Sbjct 696	GGCTGCCTTCGCCATCGGTATTCTCCAGATCTCTACGCATTTACCGCTACACCTGGAA	637		
Query 721	TTCTACCATCCTCTCCCATACTCTAGCCAACAGTATCGAATGCAATTCCCAAGTTAAGC	780		
Sbjct 636	TTCTACCATCCTCTCCCATACTCTAGCCAACAGTATCGAATGCAATTCCCAAGTTAAGC	577		
Query 781	TCGGGGATTTACATTTGACTTAATTGGCCACCTACGCGCGCTTTACGCCCAGAAAATCC	840		
Sbjct 576	TCGGGGATTTACATTTGACTTAATTGGCCACCTACGCGCGCTTTACGCCCAGTAAATCC	517		
Query 841	GATTAACGCTTGACACCTCTGTATTACCGCGGCTGCTGGCACAAAATTAACCGGGGCTTA	900		
Sbjct 516	GATTAACGCTTGACACCTCTGTATTACCGCGGCTGCTGGCACAGAGTTAGCCGGTGCTTA	457		
Query 901	TTCTGCAATAACGTCCACTATCCAAAAATAT	932		
Sbjct 456	TTCTGCGAGTAACGTCCACTATCCAAGAGTAT	425		

- Acinetobacter lwoffii strain JCM 6840 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. JUN-14 16S ribosomal RNA gene, partial sequence
- Gamma proteobacterium F3 a5 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium partial 16S rRNA gene, clone J12
- Prolinoborus fasciculus strain MB-11 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MG-3 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MR-13 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MR-81 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain 263ZG5 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WG-70 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WG-93 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WR-168 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WB-203 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain WL-242 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain WTB-120 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AM-72 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain TB-142 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AB-185 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. LHG-BW4 partial 16S rRNA gene, strain LHG-BW4
- Bacillus methylotrophicus strain 262XY6 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain M0209 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MTB-4-1 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MTA-4 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain 263AY1 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain HA 522 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium clone XJDN-640 16S ribosomal RNA gene, partial sequence
- Bacterium N2-3 16S ribosomal RNA gene, partial sequence
- Endophytic bacterium SV900 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain SBB32 16S ribosomal RNA gene, partial sequence
- Bacterium JNKL A9 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WD206 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain LAHP2-7 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium clone 958-19 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. 11K17 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain CL 102 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WD224 16S ribosomal RNA gene, partial sequence
- Uncultured Acinetobacter sp. clone TCCC 11167 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain HIBB 9208 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. 36-19 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium clone BacB100 small subunit ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MTA-33 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MTA-2 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. Tbet-YD4320-5 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. LHG-2BW3 partial 16S rRNA gene, strain LHG-2BW3
- Acinetobacter lwoffii strain AB-207 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AR-135 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain TB-128 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AL-11 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain WR-169 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WL-228 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WL-199 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WM-50 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WG-92 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain 265ZG8 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. Cai-b1 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain ML-21 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MB-14 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MTA-24-1 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MTA-34-1 16S ribosomal RNA gene, partial sequence
- Gamma proteobacterium F3 a10 16S ribosomal RNA gene, partial sequence
- Bacterium CS0(2013) 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain T24 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WD238 16S ribosomal RNA gene, partial sequence
- Bacterium ST8(2015) strain ST8 16S ribosomal RNA gene, partial sequence
- Uncultured bacterium clone 382 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain KAR20 16S ribosomal RNA gene, complete sequence
- Acinetobacter lwoffii strain ZY-778 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain SD11 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii partial 16S rRNA gene, isolate 10
- Acinetobacter lwoffii strain 263AY7 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain WR-139 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MTB-12 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain ML-39 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain 266XY3 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain 272XG8 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AB-204 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AR-180 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain TB-122 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain AL-23 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain WR-156 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain WL-241 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WB-216-2 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WM-65 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain WG-82 16S ribosomal RNA gene, partial sequence
- Acinetobacter sp. 265ZG9 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain MR-18-2 16S ribosomal RNA gene, partial sequence
- Acinetobacter lwoffii strain ML-9 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MB-24 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MM-111 16S ribosomal RNA gene, partial sequence
- Prolinoborus fasciculus strain MG-20 16S ribosomal RNA gene, partial sequence
- e-proteobacteria | 3 leaves
- Prolinoborus fasciculus partial 16S rRNA gene, isolate 56 SP635

● Acinetobacter lwoffii partial 16S rRNA gene, strain Marseille-1786

Bacillus licheniformis DQ071560.1

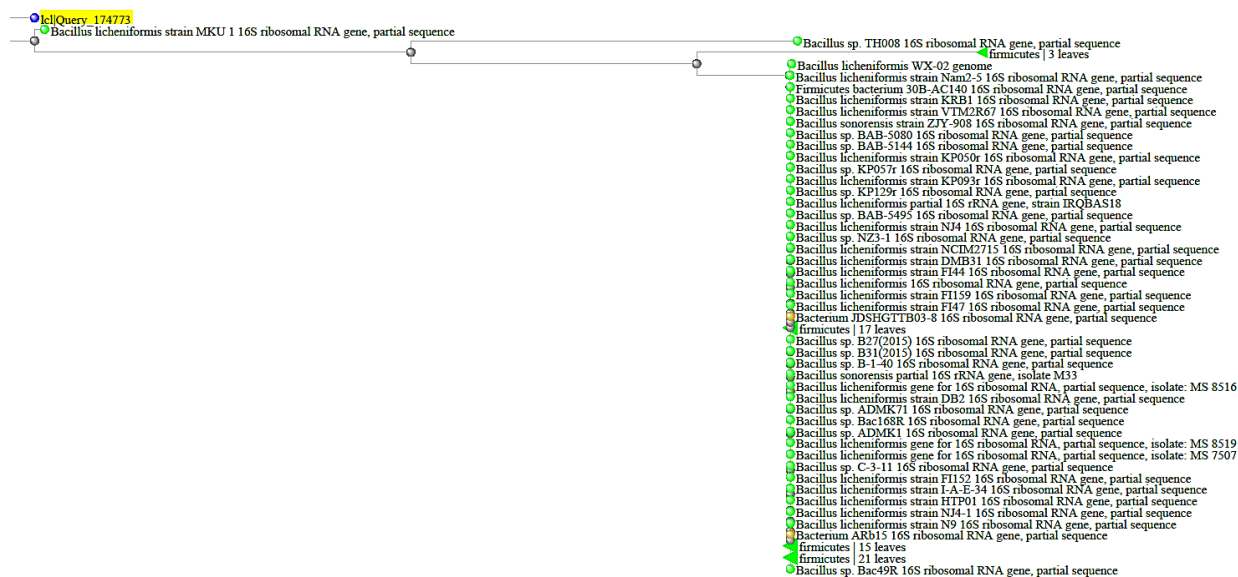
Bacillus licheniformis strain MKU 1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|DQ071560.1|](#) Length: 1430 Number of Matches: 1

Range 1: 437 to 1388 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1701 bits(1886)	0.0	949/952(99%)	1/952(0%)	Plus/Minus
Query 1	AACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGAACGTATTCACGCGGCATG	60		
Sbjct 1388	AACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGAACGTATTCACGCGGCATG	1329		
Query 61	CTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGA	120		
Sbjct 1328	CTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGA	1269		
Query 121	ACTGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTTCTGCC	180		
Sbjct 1268	ACTGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTTCTGCC	1209		
Query 181	CATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCAC	240		
Sbjct 1208	CATTGTAGCACGTGTGTAGCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCAC	1149		
Query 241	CTTCCTCCGGTTTGTACCCGCGAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAA	300		
Sbjct 1148	CTTCCTCCGGTTTGTACCCGCGAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAA	1089		
Query 301	GATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAAACATCTCACGACACGAGCTGACGAC	360		
Sbjct 1088	GATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAAACATCTCACGACACGAGCTGACGAC	1029		
Query 361	AACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTTGTGAG	420		
Sbjct 1028	AACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGCCCTATCTCTAGGGTTGTGAG	969		
Query 421	AGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAACCCACATGCTCCACC	480		
Sbjct 968	AGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAACCCACATGCTCCACC	909		
Query 481	GCTTGTGCGGGCCCCGTCATTCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGC	540		
Sbjct 908	GCTTGTGCGGGCCCCGTCATTCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGC	849		
Query 541	GGAGTGCTTAATGCGTTTGTGTCAGCACTAAAGGGCGGAAACCCCTCTAACACTTAGCACT	600		
Sbjct 848	GGAGTGCTTAATGCGTTTGTGTCAGCACTAAAGGGCGGAAACCCCTCTAACACTTAGCACT	789		
Query 601	CATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACGCTTTCGCG	660		
Sbjct 788	CATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTGCTCCCCACGCTTTCGCG	729		
Query 661	CCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACATCTCT	720		
Sbjct 728	CCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCTCCACATCTCT	669		
Query 721	ACGCATTTACCGCTACACGTGGAATTCACCTCTCCTCTTCTGCACTCAAGTTCCCCAGT	780		
Sbjct 668	ACGCATTTACCGCTACACGTGGAATTCACCTCTCCTCTTCTGCACTCAAGTTCCCCAGT	609		
Query 781	TTCCAATGANCCCTCCCGGTTGAGCCGGGGGCTTTTACATCAAACCTAAAAAACCGCCTG	840		
Sbjct 608	TTCCAATGACCCCTCCCGGTTGAGCCGGGGGCTTTTACATCAAACCTAAAAAACCGCCTG	549		
Query 841	CGCGCGCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTG	900		
Sbjct 548	CGCGCGCTTTACGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTG	489		
Query 901	CTGGCAGCTAATTAGCCG-GGCTTTCTGGTTAGGTACCGTCAAGGTACCGCC	951		
Sbjct 488	CTGGCAGCTAATTAGCCGTTGCTTTCTGGTTAGGTACCGTCAAGGTACCGCC	437		



Bacillus megaterium KU550043.1

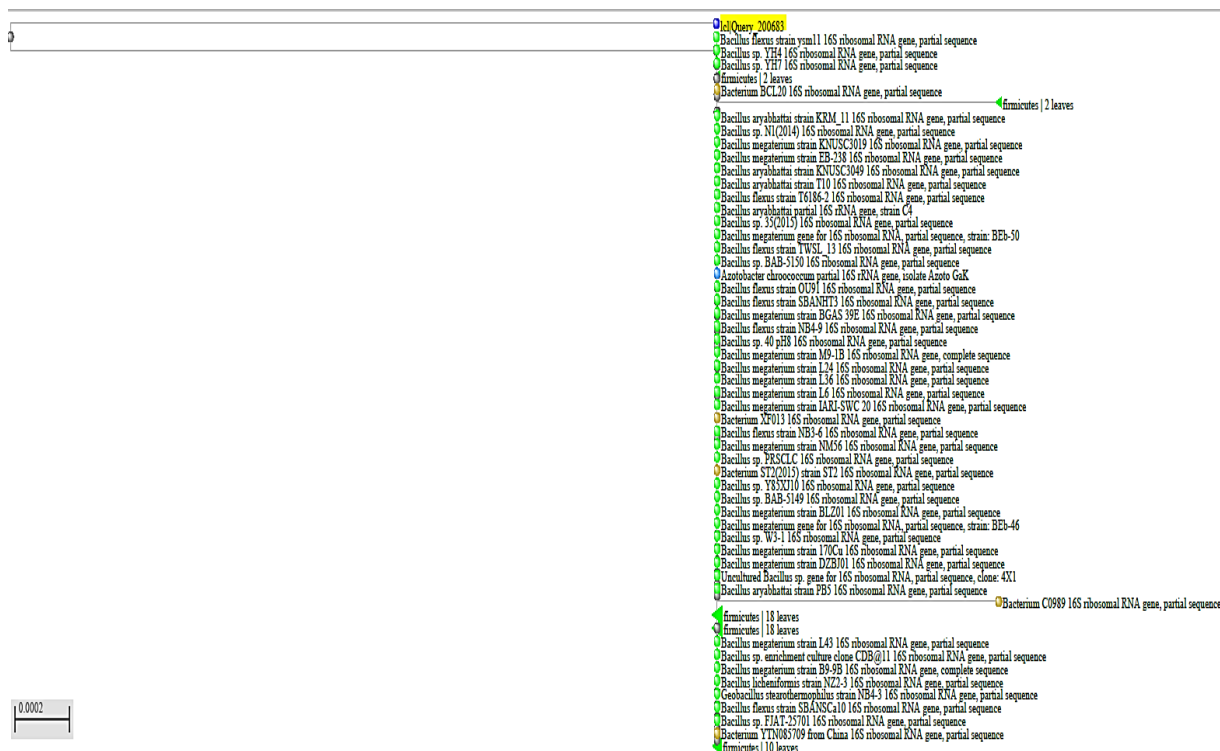
Bacillus megaterium strain EB-238 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KU550043.1](#) Length: 1406 Number of Matches: 1

Range 1: 395 to 1361 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1721 bits(1908)	0.0	962/967(99%)	0/967(0%)	Plus/Minus
Query 1	ACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCATGC	60		
Sbjct 1361	ACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCATGC	1302		
Query 61	TGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACAATCCGAA	120		
Sbjct 1301	TGATCCGCGATTACTAGCGATTCCAGCTTCATGTAGGCGAGTTGCAGCCTACAATCCGAA	1242		
Query 121	CTGAGAATGGTTTTATGGGATTGGCTTGACCTCGCGGTCTTGACGCCCTTTGTACCATCC	180		
Sbjct 1241	CTGAGAATGGTTTTATGGGATTGGCTTGACCTCGCGGTCTTGACGCCCTTTGTACCATCC	1182		
Query 181	ATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACC	240		
Sbjct 1181	ATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCCACC	1122		
Query 241	TTCTCCGGTTTGTACCCGGCAGTCACCTTAGAGTGCCCAACTAAATGCTGGCAACTAAG	300		
Sbjct 1121	TTCTCCGGTTTGTACCCGGCAGTCACCTTAGAGTGCCCAACTAAATGCTGGCAACTAAG	1062		
Query 301	ATCAAGGGTTGCGCTCGTTGCGGGACTTAACCAACATCTCACGACACGAGCTGACGACA	360		
Sbjct 1061	ATCAAGGGTTGCGCTCGTTGCGGGACTTAACCAACATCTCACGACACGAGCTGACGACA	1002		
Query 361	ACCATGCACCACCTGTCACTCTGTCCCCGAAGGGGAACGCTCTATCTCTAGAGTTGTCA	420		
Sbjct 1001	ACCATGCACCACCTGTCACTCTGTCCCCGAAGGGGAACGCTCTATCTCTAGAGTTGTCA	942		
Query 421	GAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCAC	480		
Sbjct 941	GAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCAC	882		
Query 481	CGCTTGTGCGGGCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGG	540		
Sbjct 881	CGCTTGTGCGGGCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGG	822		
Query 541	CGGAGTGCTTAATGCGTTAGCTGCAGCACTAAAGGGCGGAAACCCTCTAACACTTAGCAC	600		
Sbjct 821	CGGAGTGCTTAATGCGTTAGCTGCAGCACTAAAGGGCGGAAACCCTCTAACACTTAGCAC	762		
Query 601	TCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCACGCTTTCGC	660		
Sbjct 761	TCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGTCTCCACGCTTTCGC	702		
Query 661	GCCTCAGCGTCAGTTACAGACCAAAAAGCCGCCTTCGCCACTGGTGTTCTCCACATCTC	720		
Sbjct 701	GCCTCAGCGTCAGTTACAGACCAAAAAGCCGCCTTCGCCACTGGTGTTCTCCACATCTC	642		
Query 721	TACGCATTTACCGCTACACGTGGAATTCGCTTTTCTCTTCTGCACTCAAGTTCCCCAG	780		
Sbjct 641	TACGCATTTACCGCTACACGTGGAATTCGCTTTTCTCTTCTGCACTCAAGTTCCCCAG	582		
Query 781	TTTCCAATGACCTCCACGGTTGAGCCGTGGGCTTTCACATCAAACCTAAAAAACCGCCT	840		
Sbjct 581	TTTCCAATGACCTCCACGGTTGAGCCGTGGGCTTTCACATCAGACTTAAGAAACCGCCT	522		
Query 841	GCGCGCGCTTTACGCCCAATAATTCGCGGAAAACGCTTGCCACCTACGTATTACCGCGGC	900		
Sbjct 521	GCGCGCGCTTTACGCCCAATAATTCGCGGATAACGCTTGCCACCTACGTATTACCGCGGC	462		
Query 901	TGCTGGCACGTATTTAGCCGGGGCTTTCTGGTTAGGTACCGTCAAGGTACAAGCAGTTAC	960		
Sbjct 461	TGCTGGCACGTAGTTAGCCGTGGCTTTCTGGTTAGGTACCGTCAAGGTACAAGCAGTTAC	402		
Query 961	TCTTGTA 967			
Sbjct 401	TCTTGTA 395			



Staphylococcus succinus KJ888125.1

Staphylococcus succinus strain MI-6 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ888125.1](#) Length: 1381 Number of Matches: 1

Range 1: 40 to 988 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1660 bits(1840)	0.0	940/949(99%)	3/949(0%)	Plus/Plus
Query 1	AACACGTGGGTAACCTACCTATAAGACTGGAATAACTTCGGGAAACCGGAGCTAATGCCG	60		
Sbjct 40	AACACGTGGGTAACCTACCTATAAGACTGGAATAACTTCGGGAAACCGGAGCTAATGCCG	99		
Query 61	GATAACATATAGAACCGCATGGTTCTATAGTGAAAGATGGTTTTGCTATCACTTATAGAT	120		
Sbjct 100	GATAACATATAGAACCGCATGGTTCTATAGTGAAAGATGGTTTTGCTATCACTTATAGAT	159		
Query 121	GGACCCGCGCCGTATTAGCTAGTTGGTAAGGTAATGGCTTACCAAGGCGACGATACGTAG	180		
Sbjct 160	GGACCCGCGCCGTATTAGCTAGTTGGTAAGGTAATGGCTTACCAAGGCGACGATACGTAG	219		
Query 181	CCGACCTGAGAGGGTGATCGGCCACACTGGAAGTACGACACGGTCCAGACTCCTACGGGA	240		
Sbjct 220	CCGACCTGAGAGGGTGATCGGCCACACTGGAAGTACGACACGGTCCAGACTCCTACGGGA	279		
Query 241	GGCAGCAGTAGGGAATCTTCCGCAATGGGCGAAAGCCTGACGGAGCAACGCCGCGTGAGT	300		
Sbjct 280	GGCAGCAGTAGGGAATCTTCCGCAATGGGCGAAAGCCTGACGGAGCAACGCCGCGTGAGT	339		
Query 301	GATGAAGGTTTTTCGGATCGTAAACTCTGTTATTAGGGAAGAACAATGCGTAAGTAACT	360		
Sbjct 340	GATGAAGGTTTTTCGGATCGTAAACTCTGTTATTAGGGAAGAACAATGCGTAAGTAACT	399		
Query 361	GTGCGCATCTTGACGGTACCTAATCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCG	420		
Sbjct 400	GTGCGCATCTTGACGGTACCTAATCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCG	459		
Query 421	GTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCGTAAAGCGCGCGTAGGCCGT	480		
Sbjct 460	GTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCGTAAAGCGCGCGTAGGCCGT	519		
Query 481	TTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGAGGGTCATTGGAACTGGGAA	540		
Sbjct 520	TTCTTAAGTCTGATGTGAAAGCCACGGCTCAACCGTGGAGGGTCATTGGAACTGGGAA	579		
Query 541	ACTTGAGTGCAGAAGAGGAAAGTGAATTCATGTGTAGCGGTGAAATGCGCAGAGATAT	600		
Sbjct 580	ACTTGAGTGCAGAAGAGGAAAGTGAATTCATGTGTAGCGGTGAAATGCGCAGAGATAT	639		
Query 601	GGAGGAACACCAGTGGCGAAGGCGACTTCTGGTCTGTAAGTACGCTGATGTGCGAAAG	660		
Sbjct 640	GGAGGAACACCAGTGGCGAAGGCGACTTCTGGTCTGTAAGTACGCTGATGTGCGAAAG	699		
Query 661	CGTGGGGATCAAACAGGATTAGATACCTGGTAGTCCACGCCGTAAACGATGAGTGCTAA	720		
Sbjct 700	CGTGGGGATCAAACAGGATTAGATACCTGGTAGTCCACGCCGTAAACGATGAGTGCTAA	759		
Query 721	GTGTTAGGGGGTTTTCCGCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCCTGGGG	780		
Sbjct 760	GTGTTAGGGGGTTTTCCGCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCCTGGGG	819		
Query 781	AGTACGACCGCAAGGTTGAAACTCAAAGGAATTGACGGGGACCCGCACAAGCGGTGGAGC	840		
Sbjct 820	AGTACGACCGCAAGGTTGAAACTCAAAGGAATTGACGGGGACCCGCACAAGCGGTGGAGC	879		
Query 841	ATGTGGTTTTAATTCGAAGCAACGCGAAAAACCTTACC-AATCTTGACATCCTTTG-AAAC	898		
Sbjct 880	ATGTGGTTTTAATTCGAAGCAACGCGAAGAACCTTACCAAATCTTGACATCCTTTGAAAAAC	939		
Query 899	TCTAAAGATAAAGCCTTCCCTTCGGGGGA-AAAGGGACAggggggggCA	946		
Sbjct 940	TCTAGAGATAGAGCCTTCCCTTCGGGGGACAAAGTGACAGGTGGTGCA	988		



Bacillus pumilus KU230023.1

Bacillus pumilus strain m350 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KU230023.1](#) Length: 1396 Number of Matches: 1

Range 1: 399 to 1340 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1663 bits(1844)	0.0	934/942(99%)	0/942(0%)	Plus/Minus
Query 1	TGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTGATCCGCGA	60		
Sbjct 1340	TGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGCGGCATGCTGATCCGCGA	1281		
Query 61	TTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACTGAGAACAG	120		
Sbjct 1280	TTACTAGCGATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACTGAGAACAG	1221		
Query 121	ATTTATGGGATTGGCTAAACCTTGCGGTCTTGCGGCCCTTGTTCTGTCCATTGTAGCAC	180		
Sbjct 1220	ATTTATGGGATTGGCTAAACCTTGCGGTCTTGCGGCCCTTGTTCTGTCCATTGTAGCAC	1161		
Query 181	GTGTGTAGCCAGGTTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCCGGT	240		
Sbjct 1160	GTGTGTAGCCAGGTTCATAAGGGGCATGATGATTTGACGTCATCCCCACCTTCCTCCGGT	1101		
Query 241	TTGTCACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGATCAAGGGTT	300		
Sbjct 1100	TTGTCACCGGCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGATCAAGGGTT	1041		
Query 301	GCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAACCATGCACC	360		
Sbjct 1040	GCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAACCATGCACC	981		
Query 361	ACCTGTCACTCTGTCCCCGAAGGGAAAGCCCTATCTCTAGGGTTGTCAGAGGATGTCAAG	420		
Sbjct 980	ACCTGTCACTCTGTCCCCGAAGGGAAAGCCCTATCTCTAGGGTTGTCAGAGGATGTCAAG	921		
Query 421	ACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGG	480		
Sbjct 920	ACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGG	861		
Query 481	CCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGCGGAGTGCTTAA	540		
Sbjct 860	CCCCCGTCAATTCCCTTTGAGTTTCAGTCTTGCGACCGTACTCCCCAGGCGGAGTGCTTAA	801		
Query 541	TGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCATCGTTTACG	600		
Sbjct 800	TGCGTTAGCTGCAGCACTAAGGGGCGGAAACCCCTAACACTTAGCACTCATCGTTTACG	741		
Query 601	GCGTGGAATACCAGGGTATCTAATCCTGTTGCGCTCCCCACGCTTTCGCTCCTCAGCGTCA	660		
Sbjct 740	GCGTGGAATACCAGGGTATCTAATCCTGTTGCGCTCCCCACGCTTTCGCTCCTCAGCGTCA	681		
Query 661	GTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCCTCCACATCTCTACGCATTTTAC	720		
Sbjct 680	GTTACAGACCAGAGAGTCGCCTTCGCCACTGGTGTTCCCTCCACATCTCTACGCATTTTAC	621		
Query 721	CGCTACACGTGGAATTCCACTCTCCTCTTCTGCACTCAAGTTTCCAGTTTCCAATGACC	780		
Sbjct 620	CGCTACACGTGGAATTCCACTCTCCTCTTCTGCACTCAAGTTTCCAGTTTCCAATGACC	561		
Query 781	CTCCCCGGTTGAGCCGGGGGCTTTCACATCAAACCTTAAAAAACCGCTGCGAGCCCTTTA	840		
Sbjct 560	CTCCCCGGTTGAGCCGGGGGCTTTCACATCAGACTTAAGAAACCGCTGCGAGCCCTTTA	501		
Query 841	CGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAA	900		
Sbjct 500	CGCCCAATAATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAG	441		
Query 901	TTAGCCGGGGCTTTCTGGGTTAGGTACCGTCAAGGGGCGAGCA	942		
Sbjct 440	TTAGCCGTGGCTTTCTGGTTAGGTACCGTCAAGGTGCGAGCA	399		



Bacillus licheniformis KT200463.1

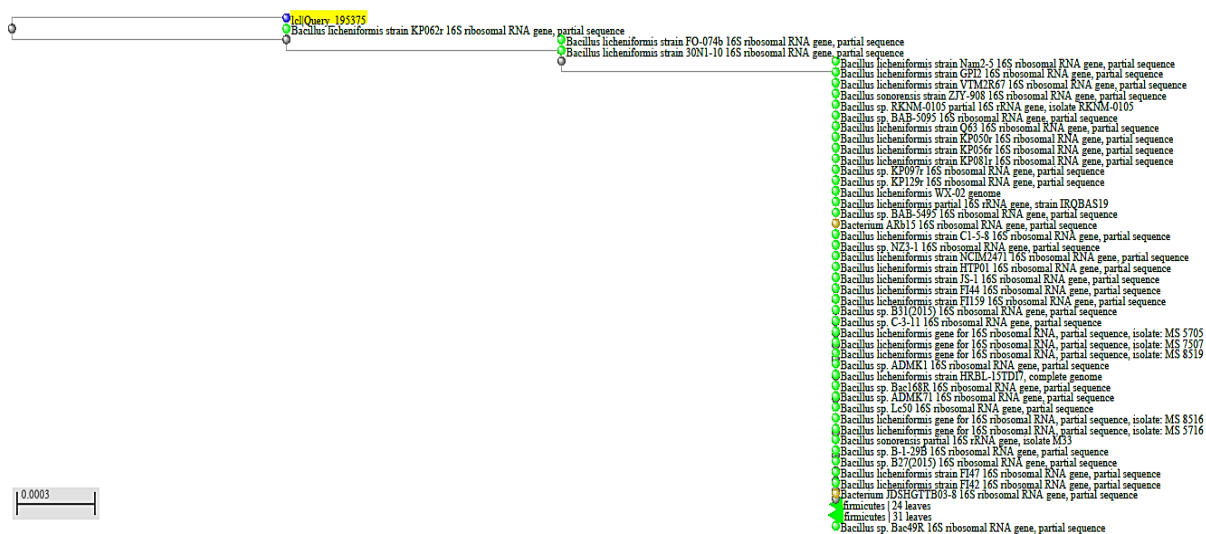
Bacillus licheniformis strain KP062r 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT200463.1](#) Length: 1201 Number of Matches: 1

Range 1: 68 to 1014 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1700 bits(1884)	0.0	945/947(99%)	0/947(0%)	Plus/Plus
Query 1	CGGTGTGTACAAGGCCCGGAACGTATTACCGCGGCATGCTGATCCGCGATTACTAGCG	60		
Sbjct 68	CGGTGTGTACAAGGCCCGGAACGTATTACCGCGGCATGCTGATCCGCGATTACTAGCG	127		
Query 61	ATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACGAGAAACAGATTTGTGGG	120		
Sbjct 128	ATTCCAGCTTCACGCAGTCGAGTTGCAGACTGCGATCCGAACGAGAAACAGATTTGTGGG	187		
Query 121	ATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTTCTGCCCATTTAGTACACGTGTGTAGC	180		
Sbjct 188	ATTGGCTTAGCCTCGCGGCTTCGCTGCCCTTTGTTCTGCCCATTTAGTACACGTGTGTAGC	247		
Query 181	CCAGGTCATAAAGGGGCATGATGATTTGACGTATCCACCTTCCTCCGGTTGTACACCG	240		
Sbjct 248	CCAGGTCATAAAGGGGCATGATGATTTGACGTATCCACCTTCCTCCGGTTGTACACCG	307		
Query 241	GCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGATCAAGGGTTGCGCTCGTT	300		
Sbjct 308	GCAGTCACCTTAGAGTGCCCAACTGAATGCTGGCAACTAAGATCAAGGGTTGCGCTCGTT	367		
Query 301	GCGGGACTTAACCCAACATCTCAGCACAGAGCTGACGACAACCATGCACCACCTGTAC	360		
Sbjct 368	GCGGGACTTAACCCAACATCTCAGCACAGAGCTGACGACAACCATGCACCACCTGTAC	427		
Query 361	TCTGCCCGCGAAGGGGAAGCCCTATCTCTAGGGTTGTGAGAGGATGTCAAGACCTGGTAA	420		
Sbjct 428	TCTGCCCGCGAAGGGGAAGCCCTATCTCTAGGGTTGTGAGAGGATGTCAAGACCTGGTAA	487		
Query 421	GGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGGCCCCGTCA	480		
Sbjct 488	GGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGGCCCCGTCA	547		
Query 481	ATTCTTTGAGTTTCAGTCTTGCAGACCGTACTCCCCAGGCGGAGTGCTTAATGCGTTTGC	540		
Sbjct 548	ATTCTTTGAGTTTCAGTCTTGCAGACCGTACTCCCCAGGCGGAGTGCTTAATGCGTTTGC	607		
Query 541	TGCAGCACTAAAGGGCGGAAACCTCTAACACTTAGCACTCATCGTTTACGGCGTGGACT	600		
Sbjct 608	TGCAGCACTAAAGGGCGGAAACCTCTAACACTTAGCACTCATCGTTTACGGCGTGGACT	667		
Query 601	ACCAGGGTATCTAATCCTGTTGCTGCCACGCTTTCGCGCCTCAGCGTCAGTTACAGAC	660		
Sbjct 668	ACCAGGGTATCTAATCCTGTTGCTGCCACGCTTTCGCGCCTCAGCGTCAGTTACAGAC	727		
Query 661	CAGAGAGTCGCTTCGCCACTGGTGTTCCTCCACATCTCTACGCATTTACCGCTACACG	720		
Sbjct 728	CAGAGAGTCGCTTCGCCACTGGTGTTCCTCCACATCTCTACGCATTTACCGCTACACG	787		
Query 721	TGGAATTCCACTCTCCTCTTCTGCACTCAAGTTCCCAAGTTTCCAATGACCCTCCCCGGT	780		
Sbjct 788	TGGAATTCCACTCTCCTCTTCTGCACTCAAGTTCCCAAGTTTCCAATGACCCTCCCCGGT	847		
Query 781	TGAGCCGGGGGCTTTACATCAAACCTTAAGAAACCGCTGCGCGCGCTTTACGCCCAATA	840		
Sbjct 848	TGAGCCGGGGGCTTTACATCAAACCTTAAGAAACCGCTGCGCGCGCTTTACGCCCAATA	907		
Query 841	ATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAATTAGCCGTG	900		
Sbjct 908	ATTCCGGACAACGCTTGCCACCTACGTATTACCGCGGCTGCTGGCACGTAATTAGCCGTG	967		
Query 901	GCTTTCTGGTTAGGTACCGTCAAGGTACCGCCCTATTCCAACGGGAC	947		
Sbjct 968	GCTTTCTGGTTAGGTACCGTCAAGGTACCGCCCTATTCCAACGGGAC	1014		

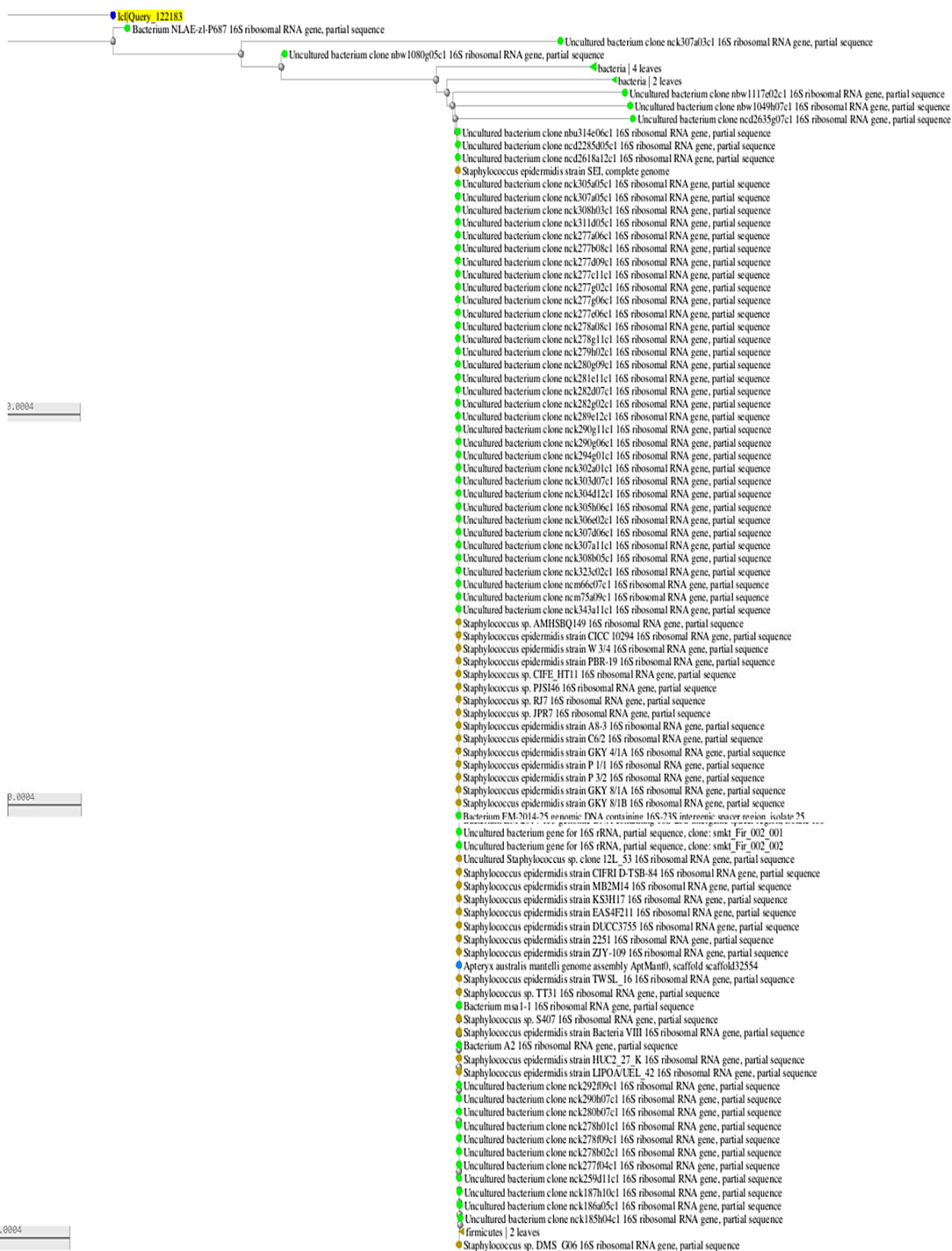


9- The phylogenetic analysis of bacteria isolated from upper surfaces of water taps samples

Staphylococcus epidermidis KT887972.1

Staphylococcus epidermidis strain LIPOA/Uel_42 16S ribosomal RNA gene, partial sequence
Sequence ID: [gb|KT887972.1](#) Length: 1327 Number of Matches: 1

Range 1: 3 to 1051		GenBank	Graphics			Next Match	Previous Match
Score	Expect	Identities		Gaps	Strand		
1810 bits(2006)	0.0	1033/1049(98%)		3/1049(0%)	Plus/Plus		
Query	1	GCTCCTCTGACGTTAGCGGCGGACGGGTGAGTAACACGTTGGATAACCTACCTATAAGACT				60	
Sbjct	3	GCTCCTCTGACGTTAGCGGCGGACGGGTGAGTAACACGTTGGATAACCTACCTATAAGACT				62	
Query	61	GGGATAAATTTCGGGAAACCGGAGCTAATACCGGATAATATATTGAACCGCATGGTTCAAT				120	
Sbjct	63	GGGATAAATTTCGGGAAACCGGAGCTAATACCGGATAATATATTGAACCGCATGGTTCAAT				122	
Query	121	AGTGAAAGACGGTTTGTGCTCACTTATAGATGGATCCGCGCCGCATTAGCTAGTTGGTA				180	
Sbjct	123	AGTGAAAGACGGTTTGTGCTCACTTATAGATGGATCCGCGCCGCATTAGCTAGTTGGTA				182	
Query	181	AGGTAACGGCTTACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACT				240	
Sbjct	183	AGGTAACGGCTTACCAAGGCAACGATGCGTAGCCGACCTGAGAGGGTGATCGGCCACACT				242	
Query	241	GGAAGTGAACACGGTCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGG				300	
Sbjct	243	GGAAGTGAACACGGTCCAGACTCCTACGGGAGGCAGCAGTAGGGAATCTTCCGCAATGG				302	
Query	301	GCGAAAGCCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTCTTCGGATCGTAAACTCT				360	
Sbjct	303	GCGAAAGCCTGACGGAGCAACGCCGCGTGAGTGATGAAGGTCTTCGGATCGTAAACTCT				362	
Query	361	GTTATTAGGGAAGAACAAATGTGTAAAGTAAGTATGCACGTCTTGACGGTACCTAATCAGA				420	
Sbjct	363	GTTATTAGGGAAGAACAAATGTGTAAAGTAAGTATGCACGTCTTGACGGTACCTAATCAGA				422	
Query	421	AAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCG				480	
Sbjct	423	AAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCG				482	
Query	481	GAATTATTGGGCGTAAAGCGCGCTAGGCGGTTTTTTAAGTCTGATGTGAAAGCCCACGG				540	
Sbjct	483	GAATTATTGGGCGTAAAGCGCGCTAGGCGGTTTTTTAAGTCTGATGTGAAAGCCCACGG				542	
Query	541	CTCAACCGTGGAGGGTCATTGGAAACTGGAAACTTGAGTGCAGAGAGGAAAGTGGAAT				600	
Sbjct	543	CTCAACCGTGGAGGGTCATTGGAAACTGGAAACTTGAGTGCAGAGAGGAAAGTGGAAT				602	
Query	601	TCCATGTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAAGTGGCGAAGGCGACTT				660	
Sbjct	603	TCCATGTGTAGCGGTGAAATGCGCAGAGATATGGAGGAACACCAAGTGGCGAAGGCGACTT				662	
Query	661	TCTGGTCTGTAAGTACGCTGATGTGCGAAAGCGTGGGGATCAAACAGGATTAGATACCC				720	
Sbjct	663	TCTGGTCTGTAAGTACGCTGATGTGCGAAAGCGTGGGGATCAAACAGGATTAGATACCC				722	
Query	721	TGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCTTAGTGC				780	
Sbjct	723	TGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGTTTCCGCCCTTAGTGC				782	
Query	781	TGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGACCGCAAGGTTGAAACTCAAAG				840	
Sbjct	783	TGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGTACGACCGCAAGGTTGAAACTCAAAG				842	
Query	841	GAATTGACGGGGACCCGCACAAGCGGTGGAGCATGTGGTTTTAATTCGAAGCAACGCGAAA				900	
Sbjct	843	GAATTGACGGGGACCCGCACAAGCGGTGGAGCATGTGGTTTTAATTCGAAGCAACGCGAAG				902	
Query	901	AACCTTACCAAATCTTGACTTCCTCTGACCCCTCTAAAAATAAAGTTTCCCTTCGGGG				960	
Sbjct	903	AACCTTACCAAATCTTGACTTCCTCTGACCCCTCTAGAGATAGAGTTTCCCTTCGGGG				962	
Query	961	GAAAAAATGACAGG-GGGGCATGG-TGTCGTCAGCTCGGGTGGGAAATGTTGGGTTAAG				1018	
Sbjct	963	GACAGAGTGACAGGTGGTGCATGGTTGTCGTCAGCTCGTGTCTGAGATGTTGGGTTAAG				1022	
Query	1019	TCCC-CCACGAGCGCAACCTTAAGCTTA		1046			
Sbjct	1023	TCCCGCAACGAGCGCAACCTTAAGCTTA		1051			



Rothia amarae NR_029045.1

Rothia amarae strain J18 16S ribosomal RNA gene, partial sequence

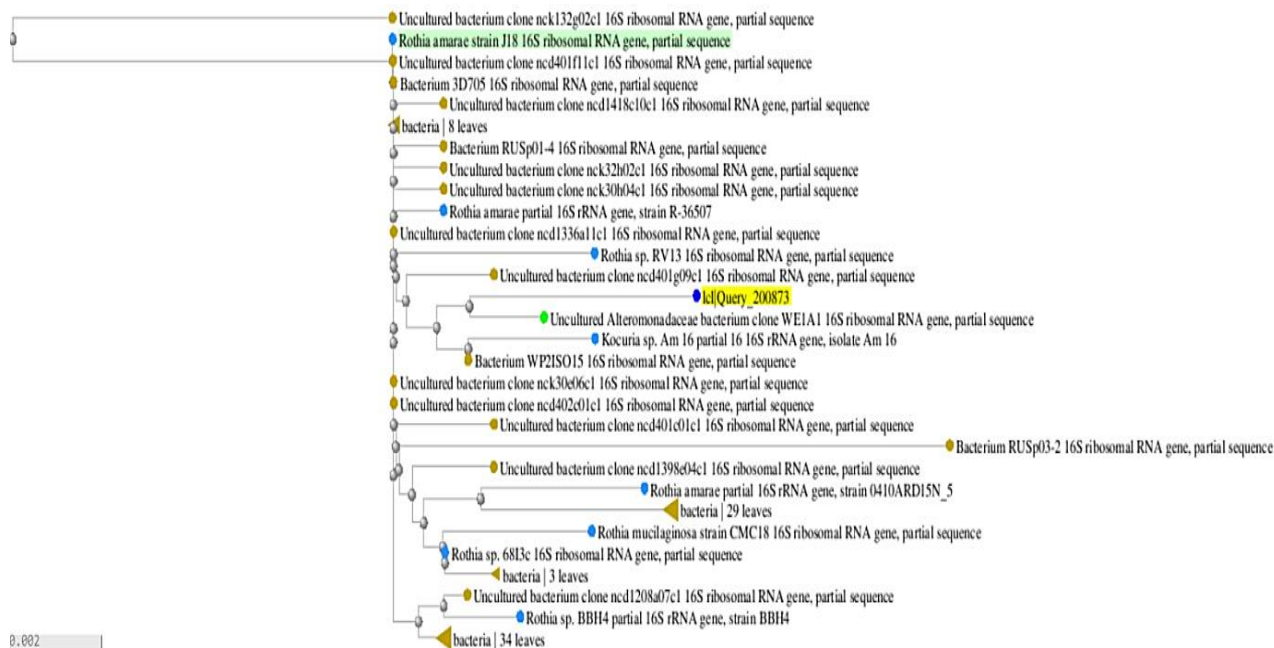
Sequence ID: [ref|NR_029045.1|](#) Length: 1467 Number of Matches: 1

► [See 1 more title\(s\)](#)

Range 1: 98 to 1028 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1644 bits(1822)	0.0	924/931(99%)	1/931(0%)	Plus/Plus
Query 1	GAACGGGTGAGTAATACGTGAGTAACCTGCCTTTGACTCTGGGATAAGCCTGGGAAACTG	60		
Sbjct 98	GAACGGGTGAGTAATACGTGAGTAACCTGCCTTTGACTCTGGGATAAGCCTGGGAAACTG	157		
Query 61	GGTCTAATACCGGATATGACAAGGAACCGCATGGTTTTTTGTGGAAAGGGTTTGTACTGG	120		
Sbjct 158	GGTCTAATACCGGATATGACAAGGAACCGCATGGTTTTTTGTGGAAAGGGTTTGTACTGG	217		
Query 121	TTTTAGATGGGCTCACGGCCTATCAGCTTGTTGGTGGGGTAATGGCTCACCAAGGCGACG	180		
Sbjct 218	TTTTAGATGGGCTCACGGCCTATCAGCTTGTTGGTGGGGTAATGGCTCACCAAGGCGACG	277		
Query 181	ACGGGTAGCCGGCCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCAGACTC	240		
Sbjct 278	ACGGGTAGCCGGCCTGAGAGGGTGACCGGCCACACTGGGACTGAGACACGGCCAGACTC	337		
Query 241	CTACGGGAGGCAGCAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCC	300		
Sbjct 338	CTACGGGAGGCAGCAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCGACGCC	397		
Query 301	GCGTGAGGGATGACGGCCTTCGGGTTGTAAACCTCTTTCAGCAGGGAAGAAGCGAAAGTG	360		
Sbjct 398	GCGTGAGGGATGACGGCCTTCGGGTTGTAAACCTCTTTCAGCAGGGAAGAAGCGAAAGTG	457		
Query 361	ACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGG	420		
Sbjct 458	ACGGTACCTGCAGAAGAAGCGCCGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGG	517		
Query 421	GCGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTTGTAGGCGGTTTGTTCGCTCTG	480		
Sbjct 518	GCGCGAGCGTTGTCCGGAATTATTGGGCGTAAAGAGCTTGTAGGCGGTTTGTTCGCTCTG	577		
Query 481	CTGTGAAAGACCGGGGCTTAACCCCGGTATTGCAGTGGGTACGGGCAGACTAGAGTGCAG	540		
Sbjct 578	CTGTGAAAGACCGGGGCTTAACCCCGGTATTGCAGTGGGTACGGGCAGACTAGAGTGCAG	637		
Query 541	TAGGGGAGACTGGAATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCG	600		
Sbjct 638	TAGGGGAGACTGGAATTCCTGGTGTAGCGGTGAAATGCGCAGATATCAGGAGGAACACCG	697		
Query 601	ATGGCGAAGGCAGGTCTCTGGGCTGTAACGTGACGCTGAGAAGCGAAAGCATGGGGAGCGA	660		
Sbjct 698	ATGGCGAAGGCAGGTCTCTGGGCTGTAACGTGACGCTGAGAAGCGAAAGCATGGGGAGCGA	757		
Query 661	ACAGGATTAGATACCCTGGTAGTCCATGCCGTAAACGTTGGGCACTAGGTGTGGGGGACA	720		
Sbjct 758	ACAGGATTAGATACCCTGGTAGTCCATGCCGTAAACGTTGGGCACTAGGTGTGGGGGACA	817		
Query 721	TTCCACGTTTTCCGCGCCGTAGCTAACGCATTAAGTGCCCCGCCTGGGGAGTACGGCCGC	780		
Sbjct 818	TTCCACGTTTTCCGCGCCGTAGCTAACGCATTAAGTGCCCCGCCTGGGGAGTACGGCCGC	877		
Query 781	AAGGCTAAAACCTCAAAGGAATTGACGGGGGCCCGCACAAAGCGGCGACCATGCGGATTAA	840		
Sbjct 878	AAGGCTAAAACCTCAAAGGAATTGACGGGGGCCCGCACAAAGCGGCGGAGCATGCGGATTAA	937		
Query 841	TTCGATGCAACGCGAAAAACCTTACCAAGGCTTGACATATACTGGACCGCCTCAAAAATG	900		
Sbjct 938	TTCGATGCAACGCGAAGAACCTTACCAAGGCTTGACATATACTGGACCGCCTCAGAGATG	997		
Query 901	GGGTTTCCCTTC-GGGCTGGTATACAGGGGG	930		
Sbjct 998	GGGTTTCCCTTCGGGGCTGGTATACAGGTGG	1028		



0.002

Delftia lacustris KT958881.1

Delftia lacustris strain BMCH-IB-C 142 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT958881.1](#) Length: 1360 Number of Matches: 1

Range 1: 355 to 1340 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1745 bits(1934)	0.0	980/986(99%)	3/986(0%)	Plus/Minus
Query 1	CCTACTTCTGGCGAGACCCGCTCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAA	60		
Sbjct 1340	CCTACTTCTGGCGAGACCCGCTCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAA	1281		
Query 61	CGTATTACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCGACTTCACGCAGTCGAG	120		
Sbjct 1280	CGTATTACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCGACTTCACGCAGTCGAG	1221		
Query 121	TTGCAGACTGCGATCCGGACTACGACTGGTTTTATGGGATTAGCTCCCCCTCGCGGGTTG	180		
Sbjct 1220	TTGCAGACTGCGATCCGGACTACGACTGGTTTTATGGGATTAGCTCCCCCTCGCGGGTTG	1161		
Query 181	GCAACCCCTCTGTACCAGCCATTGTATGACGTGTGTAGCCCCACCTATAAGGGCCATGAGG	240		
Sbjct 1160	GCAACCCCTCTGTACCAGCCATTGTATGACGTGTGTAGCCCCACCTATAAGGGCCATGAGG	1101		
Query 241	ACTTGACGTCATCCCCACCTTCTCCTCCGGTTTGTACCGGCAGTCTCATTAGAGTGCTCAA	300		
Sbjct 1100	ACTTGACGTCATCCCCACCTTCTCCTCCGGTTTGTACCGGCAGTCTCATTAGAGTGCTCAA	1041		
Query 301	CTGAATGTAGCAACTAATGACAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCA	360		
Sbjct 1040	CTGAATGTAGCAACTAATGACAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCA	981		
Query 361	CGACACGAGCTGACGACAGCCATGCAGCACCTGTGTGCAGGTTCTCTTCGAGCACGAAT	420		
Sbjct 980	CGACACGAGCTGACGACAGCCATGCAGCACCTGTGTGCAGGTTCTCTTCGAGCACGAAT	921		
Query 421	CCATCTCTGGAAACTTCCTGCCATGTCAAAGGTGGGTAAGGTTTTTCGCGTTGCATCGAA	480		
Sbjct 920	CCATCTCTGGAAACTTCCTGCCATGTCAAAGGTGGGTAAGGTTTTTCGCGTTGCATCGAA	861		
Query 481	TTAAACCACATCATCCACCGCTTGTGCGGGTCCCGTCAATTTCCTTTGAGTTTCAACCTT	540		
Sbjct 860	TTAAACCACATCATCCACCGCTTGTGCGGGTCCCGTCAATTTCCTTTGAGTTTCAACCTT	801		
Query 541	GCGGCCGTACTCCCCAGGCGGTCAACTTCACGCGTTAGCTTCGTTACTGAGAAAATAAT	600		
Sbjct 800	GCGGCCGTACTCCCCAGGCGGTCAACTTCACGCGTTAGCTTCGTTACTGAGAAAATAAT	741		
Query 601	TCCCAACAACCAAGTTGACATCGTTTAGGGCGTGGACTACCAGGGTATCTAATCCTGTTTG	660		
Sbjct 740	TCCCAACAACCAAGTTGACATCGTTTAGGGCGTGGACTACCAGGGTATCTAATCCTGTTTG	681		
Query 661	CTCCCCACGCTTTTCGTGCATGAGCGTCAGTACAGGTCCAGGGGATTGCCTTCGCCATCGG	720		
Sbjct 680	CTCCCCACGCTTTTCGTGCATGAGCGTCAGTACAGGTCCAGGGGATTGCCTTCGCCATCGG	621		
Query 721	TGTTCTCCTCCGCATATCTACGCATTTCACTGCTACACGCGGAATTCATCCCCCTCTACCG	780		
Sbjct 620	TGTTCTCCTCCGCATATCTACGCATTTCACTGCTACACGCGGAATTCATCCCCCTCTACCG	561		
Query 781	TACTCTAGCCATGCAGTCACAAATGCAGTTCCAGGTTGAGCCCGGGATTTCACATCTG	840		
Sbjct 560	TACTCTAGCCATGCAGTCACAAATGCAGTTCCAGGTTGAGCCCGGGATTTCACATCTG	501		
Query 841	TCTTACATAAACCGCCTGCGCAGCCTTTACGCCAGTAATTCCGATTAAACGCTCGCACCCCT	900		
Sbjct 500	TCTTACATAAACCGCCTGCGCAGCCTTTACGCCAGTAATTCCGATTAAACGCTCGCACCCCT	441		
Query 901	ACGTATTACCGCGGCTGCTGGCAGTAATTAGCCGGGGCTTATTCTTACGGTACCGTCAT	960		
Sbjct 440	ACGTATTACCGCGGCTGCTGGCAGTAGTTAGCCGGTGCTTATTCTTACGGTACCGTCAT	381		
Query 961	GGG-CCCCTGTATT--AAAGAGCTTT	983		
Sbjct 380	GGGCCCCCTGTATTAGAAGGAGCTTT	355		

	klQuery_4095
	Delfia sp. PVRh-YHB-9-1 16S ribosomal RNA gene, partial sequence
	Bacterium RRP2-22 16S rRNA gene
	Delfia sp. AN3 16S ribosomal RNA gene, partial sequence
	Delfia sp. 6.13 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium partial 16S rRNA gene, clone SMQ25
	Delfia sp. BN-HKY2 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain MTQ1 16S ribosomal RNA gene, partial sequence
	Delfia sp. JDC-3 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone 6 16S ribosomal RNA gene, partial sequence
	b-proteobacteria 3 leaves
	Delfia tsunhatensis strain P027 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain D10 16S ribosomal RNA gene, partial sequence
	Delfia sp. enrichment culture clone 12 16S ribosomal RNA gene, partial sequence
	Delfia sp. SS12.34 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain IPPBC R15 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone A5 16S ribosomal RNA gene, partial sequence
	Delfia sp. BGW1 16S ribosomal RNA gene, partial sequence
	Delfia sp. DM101 16S ribosomal RNA gene, partial sequence
	Delfia sp. SFG 16S ribosomal RNA gene, partial sequence
	Bacterium BM0414 16S ribosomal RNA gene, partial sequence
	Delfia lacustris voucher RIFA 1193 16S ribosomal RNA gene, partial sequence
	Delfia sp. X-a12 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain CI-23 16S ribosomal RNA gene, partial sequence
	Bacterium WYLW2-4 16S ribosomal RNA gene, partial sequence
	Uncultured Delfia sp. clone GIS-002-F11 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain A90 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium gene for 16S rRNA, partial sequence, clone: SSmCB08-74
	Delfia sp. PHD-6 16S ribosomal RNA gene, partial sequence
	Delfia sp. AER321 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain P039 16S ribosomal RNA gene, partial sequence
	Uncultured organism clone ELU0127-T314-S-N1_000208 small subunit ribosomal RNA gene, partial sequence
	Delfia lacustris strain BMCH-IB-C 142 16S ribosomal RNA gene, partial sequence
	Bacterium BM0429 16S ribosomal RNA gene, partial sequence
	b-proteobacteria 5 leaves
	b-proteobacteria 7 leaves
	Delfia sp. 183PP-As 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain L7 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain BN-HKY6 16S ribosomal RNA gene, partial sequence
	Delfia tsunhatensis strain SJ109 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone BC_B1_12f 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium gene for 16S rRNA, partial sequence, clone: U-1
	Delfia lacustris strain ABBid12-01 16S ribosomal RNA gene, partial sequence
	Delfia sp. 209Zn-As 16S ribosomal RNA gene, partial sequence
	Delfia lacustris strain IARI-NIAW1-34 16S ribosomal RNA gene, partial sequence
	Bacterium A134(2011) 16S ribosomal RNA gene, partial sequence
	b-proteobacteria 4 leaves
	b-proteobacteria 7 leaves
	b-proteobacteria 7 leaves
	b-proteobacteria 7 leaves
	b-proteobacteria 7 leaves
	Delfia tsunhatensis strain JPR21 16S ribosomal RNA gene, partial sequence
	Bacterium BM0431 16S ribosomal RNA gene, partial sequence
	b-proteobacteria 5 leaves

Pseudomonas aeruginosa KF680991.1

Pseudomonas aeruginosa strain ATHA23 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KF680991.1](#) Length: 1115 Number of Matches: 1

Range 1: 98 to 1040 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1649 bits(1828)	0.0	933/943(99%)	2/943(0%)	Plus/Minus	
Query 1	GTGATTGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGTGACATTCTGATTACGAT	60			
Sbjct 1040	GTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTACCGTGACATTCTGATTACGAT	981			
Query 61	TACTAGCGATTCCGACTTCACGCAGTCGAGTTGCAGACTGCGATCCGGACTACGATCGGT	120			
Sbjct 980	TACTAGCGATTCCGACTTCACGCAGTCGAGTTGCAGACTGCGATCCGGACTACGATCGGT	921			
Query 121	TTTATGGGATTAGCTCCACCTCGCGGCTTGGCAACCCTTTGTACCGACCATTGTAGCACG	180			
Sbjct 920	TTTATGGGATTAGCTCCACCTCGCGGCTTGGCAACCCTTTGTACCGACCATTGTAGCACG	861			
Query 181	TGTGTAGCCCTGGCCGTAAGGGCCATGATGACTTGACGTCATCCCCACCTTCCTCCGGTT	240			
Sbjct 860	TGTGTAGCCCTGGCCGTAAGGGCCATGATGACTTGACGTCATCCCCACCTTCCTCCGGTT	801			
Query 241	TGTCACCGGCAGTCTCCTTAGAGTGCCACCCGAGGTGCTGGTAACTAAGGACAAGGGTT	300			
Sbjct 800	TGTCACCGGCAGTCTCCTTAGAGTGCCACCCGAGGTGCTGGTAACTAAGGACAAGGGTT	741			
Query 301	GCGCTCGTTACGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAGCCATGCAGC	360			
Sbjct 740	GCGCTCGTTACGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAGCCATGCAGC	681			
Query 361	ACCTGTGTCTGAGTTCCCGAAGGCACCAATCCATCTCTGGAAAGTTCTCAGCATGTCAAG	420			
Sbjct 680	ACCTGTGTCTGAGTTCCCGAAGGCACCAATCCATCTCTGGAAAGTTCTCAGCATGTCAAG	621			
Query 421	GCCAGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGG	480			
Sbjct 620	GCCAGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCACCGCTTGTGCGGG	561			
Query 481	CCCCCGTCAATTCAATTTGAGTTTTAACCTTGCGGCCGTACTCCCCAGGCGGTGCACTTAT	540			
Sbjct 560	CCCCCGTCAATTCAATTTGAGTTTTAACCTTGCGGCCGTACTCCCCAGGCGGTGCACTTAT	501			
Query 541	CGCGTTAGCTGCGCCACTAAGATCTCAAGGATCCCAACGGCTAGTCGACATCGTTTACGG	600			
Sbjct 500	CGCGTTAGCTGCGCCACTAAGATCTCAAGGATCCCAACGGCTAGTCGACATCGTTTACGG	441			
Query 601	CGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGCACCTCAGTGTGAG	660			
Sbjct 440	CGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGCACCTCAGTGTGAG	381			
Query 661	TATCAGTCCAGGTGGTCGCCTTCGCCACTGGTGTTCCTTCCTATATCTACGCATTTACC	720			
Sbjct 380	TATCAGTCCAGGTGGTCGCCTTCGCCACTGGTGTTCCTTCCTATATCTACGCATTTACC	321			
Query 721	GCTACACAGGAAATTCACCACCCTCTACCGTACTCTAGCTCAGTAGTTTTGGATGCAAT	780			
Sbjct 320	GCTACACAGGAAATTCACCACCCTCTACCGTACTCTAGCTCAGTAGTTTTGGATGCAAT	261			
Query 781	TCCCAGGTTGAGCCCGGGGATTTACATCCAACCTTGCTGAACCACCTACGCGCGCTTTAC	840			
Sbjct 260	TCCCAGGTTGAGCCCGGGGATTTACATCCAACCTTGCTGAACCACCTACGCGCGCTTTAC	201			
Query 841	GCCCAGTAATTCCGATTAACCCCTTGACCCCTTCGTATTACCGCGGCTGCTGGCCCGAAGT	900			
Sbjct 200	GCCCAGTAATTCCGATTAACGCTTGACCCCTTCGTATTACCGCGGCTGCTGGCCCGAAGT	141			
Query 901	TACCCGG-GCTTATTCTGTTGG-AACGTCAAAAAGCAAGGGAT	941			
Sbjct 140	TAGCCGGTGCTTATTCTGTTGGTAACGTCAAACAGCAAGGTAT	98			



Delftia acidovorans (comamonas acid) JX090199.1

Delftia acidovorans strain Aal-4 16S ribosomal RNA gene, partial sequence
Sequence ID: [gb|JX090199.1](#) Length: 1412 Number of Matches: 1

Range 1: 399 to 1347		GenBank	Graphics			 Next Match	 Previous Match
Score	Expect	Identities		Gaps	Strand		
1692 bits(1876)	0.0	945/949(99%)		1/949(0%)	Plus/Minus		
Query	1	TCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTACCGCGGCATGCTG	60				
Sbjct	1347	TCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTACCGCGGCATGCTG	1288				
Query	61	ATCCGCGATTACTAGCGATTCCGACTTCACGCAGTCGAGTTGCAGACTGCGATCCGGACT	120				
Sbjct	1287	ATCCGCGATTACTAGCGATTCCGACTTCACGCAGTCGAGTTGCAGACTGCGATCCGGACT	1228				
Query	121	ACGACTGGTTTTATGGGATTAGCTCCCCCTCGCGGGTTGGCAACCCTCTGTACCAGCCAT	180				
Sbjct	1227	ACGACTGGTTTTATGGGATTAGCTCCCCCTCGCGGGTTGGCAACCCTCTGTACCAGCCAT	1168				
Query	181	TGTATGACGTGTGTAGCCCCACCTATAAGGGCCATGAGGACTTGACGTCATCCCCACCTT	240				
Sbjct	1167	TGTATGACGTGTGTAGCCCCACCTATAAGGGCCATGAGGACTTGACGTCATCCCCACCTT	1108				
Query	241	CCTCCGGTTTTGTCACCGGCAGTCTCATTAGAGTGCCCCAACTAAATGTAGCAACTAATGAC	300				
Sbjct	1107	CCTCCGGTTTTGTCACCGGCAGTCTCATTAGAGTGCCCCAACTAAATGTAGCAACTAATGAC	1048				
Query	301	AAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAGCC	360				
Sbjct	1047	AAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAGCC	988				
Query	361	ATGCAGCACCTGTGTGCAGGTTCTCTTTCGAGCACGAATCCATCTCTGGAACCTTCCTGC	420				
Sbjct	987	ATGCAGCACCTGTGTGCAGGTTCTCTTTCGAGCACGAATCCATCTCTGGAACCTTCCTGC	928				
Query	421	CATGTCAAAGGTGGGTAAGGTTTTTCGCGTTGCATCGAATTAAACCACATCATCCACCGC	480				
Sbjct	927	CATGTCAAAGGTGGGTAAGGTTTTTCGCGTTGCATCGAATTAAACCACATCATCCACCGC	868				
Query	481	TTGTGCGGGTCCCCGTCAATTCCTTTGAGTTTCAACCTTGCGGCCGTACTCCCCAGGCGG	540				
Sbjct	867	TTGTGCGGGTCCCCGTCAATTCCTTTGAGTTTCAACCTTGCGGCCGTACTCCCCAGGCGG	808				
Query	541	TCAACTTCACGCGTTAGCTTCGTTACTGAGAAAACTAATTCCCAACAACCAAGTTGACATC	600				
Sbjct	807	TCAACTTCACGCGTTAGCTTCGTTACTGAGAAAACTAATTCCCAACAACCAAGTTGACATC	748				
Query	601	GTTTAGGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGTGCATG	660				
Sbjct	747	GTTTAGGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGTGCATG	688				
Query	661	AGCGTCAGTACAGGTCCAGGGGATTGCCTTCGCCATCGGTGTTTCCTCCGCATATCTACGC	720				
Sbjct	687	AGCGTCAGTACAGGTCCAGGGGATTGCCTTCGCCATCGGTGTTTCCTCCGCATATCTACGC	628				
Query	721	ATTTCACTGCTACACGCGGAATTCATCCCCCTCTACCGTACTCTAGCCATGCAGTCACA	780				
Sbjct	627	ATTTCACTGCTACACGCGGAATTCATCCCCCTCTACCGTACTCTAGCCATGCAGTCACA	568				
Query	781	AATGCAGTTCCCA-GTTGAGCCCGGGGATTTACATCTGTCTTACATAACCGCCTGCGCA	839				
Sbjct	567	AATGCAGTTCCCAAGTTGAGCCCGGGGATTTACATCTGTCTTACATAACCGCCTGCGCA	508				
Query	840	CGCTTTACGCCCAGTAATTCGATTAAACGCTCGCACCTACGTATTACCGCGGCTGCTGG	899				
Sbjct	507	CGCTTTACGCCCAGTAATTCGATTAAACGCTCGCACCTACGTATTACCGCGGCTGCTGG	448				
Query	900	CACGTAATTAGCCGGGCTTATTCTTACGGTACCGTCATGGTCCTCTCG	948				
Sbjct	447	CACGTARTTAGCCGGTGCTTATTCTTACGGTACCGTCATGGGCCTCTCG	399				



Arthrobacter sanguinis strain 741 16S ribosomal RNA gene, partial sequence

Sequence ID: [ref|NR_044399.1|](#) Length: 1452 Number of Matches: 1

[► See 1 more title\(s\)](#)

Range 1: 419 to 1390 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1727 bits(1914)	0.0	967/972(99%)	1/972(0%)	Plus/Minus	
Query 1		CGGGTGTTACCAACTTTTCGTGACTTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTC			60
Sbjct 1390		CGGGTGTTACCAACTTTTCGTGACTTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTC			1331
Query 61		ACCGCAGCGTTGCTGATCTGCGATTACTAGCGACTCCGACTTCATGGGGTCGAGTTGCAG			120
Sbjct 1330		ACCGCAGCGTTGCTGATCTGCGATTACTAGCGACTCCGACTTCATGGGGTCGAGTTGCAG			1271
Query 121		ACCCCAATCCGAACTGAGACCGACTTTTTGGGATTAGCTCCACCTCACAGTATCGCAACC			180
Sbjct 1270		ACCCCAATCCGAACTGAGACCGACTTTTTGGGATTAGCTCCACCTCACAGTATCGCAACC			1211
Query 181		CTTTGTATCGGCCATTGTAGCATGCTTGAAGCCCAAGACATAAGGGGCATGATGATTGA			240
Sbjct 1210		CTTTGTATCGGCCATTGTAGCATGCTTGAAGCCCAAGACATAAGGGGCATGATGATTGA			1151
Query 241		CGTCATCCCCACCTTCCTCCGAGTTGACCCCGGCAGTCTCCTATGAGTCCCCACCATCAC			300
Sbjct 1150		CGTCATCCCCACCTTCCTCCGAGTTGACCCCGGCAGTCTCCTATGAGTCCCCACCATCAC			1091
Query 301		GTGCTGGCAACATAGAACGAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGA			360
Sbjct 1090		GTGCTGGCAACATAGAACGAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGA			1031
Query 361		CACGAGCTGACGACAACCATGCACCACCTGTAAACCAGCCCCGAAGGGAAACGCATCTC			420
Sbjct 1030		CACGAGCTGACGACAACCATGCACCACCTGTAAACCAGCCCCGAAGGGAAACGCATCTC			971
Query 421		TGCGGCGGTCGGTTTCATGTCAAGCCTTGTTAAGGTTCTTCGCGTTGCATCGAATTAATC			480
Sbjct 970		TGCGGCGGTCGGTTTCATGTCAAGCCTTGTTAAGGTTCTTCGCGTTGCATCGAATTAATC			911
Query 481		AGCATGCTCCGCCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTTCAGCCTTGCGGCC			540
Sbjct 910		AGCATGCTCCGCCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTTCAGCCTTGCGGCC			851
Query 541		GTACTCCCCAGGCGGGGCACTTAATGCGTTAGCTACGGCGCGGAAAACGTGGAATGCTCC			600
Sbjct 850		GTACTCCCCAGGCGGGGCACTTAATGCGTTAGCTACGGCGCGGAAAACGTGGAATGCTCC			791
Query 601		CCACACCTAGTGCCCAACGTTTACGGCATGGACTACCAGGGTATCTAATCCTGTTTCGCTC			660
Sbjct 790		CCACACCTAGTGCCCAACGTTTACGGCATGGACTACCAGGGTATCTAATCCTGTTTCGCTC			731
Query 661		CCCATGCTTTTCGCTTCTCAGCGTCAGTAAATGCCAGTAACCTGCCTTCGCCATCGGTGT			720
Sbjct 730		CCCATGCTTTTCGCTTCTCAGCGTCAGTAAATGCCAGTAACCTGCCTTCGCCATCGGTGT			671
Query 721		TCTTCCTGATATCTGCGCATTCACCGCTACACCAGGAATTCAGTTACCCCTACATCAC			780
Sbjct 670		TCTTCCTGATATCTGCGCATTCACCGCTACACCAGGAATTCAGTTACCCCTACATCAC			611
Query 781		TCTAGCCTGCCCCGTACCCACTGCAGACCCGGAGTTAAGCCCCGGGCTTTCACAGCAAACG			840
Sbjct 610		TCTAGCCTGCCCCGTACCCACTGCAGACCCGGAGTTAAGCCCCGGGCTTTCACAGCAGACG			551
Query 841		CGACAAACCGCCTACAAGCTCTTTACGCCCAATAATCCGGAAAACGCTCGCACCCCTACG			900
Sbjct 550		CGACAAACCGCCTACAAGCTCTTTACGCCCAATAATCCGGATAACGCTCGCACCCCTACG			491
Query 901		TATTACGCGGGCTGCTGGCACGTAGTTAGCCGGGGCTTCTTCTGCCAGTACCCTCCCCCG			960
Sbjct 490		TATTACGCGGGCTGCTGGCACGTAGTTAGCCGGGTGCTTCTTCTGCCAGTACCCTCACCCG			431
Query 961		AA-GCTTGTTC			971
Sbjct 430		AAGGCTTGTTC			419



Delftia acidovorans (comamonas acid) KJ781879.1

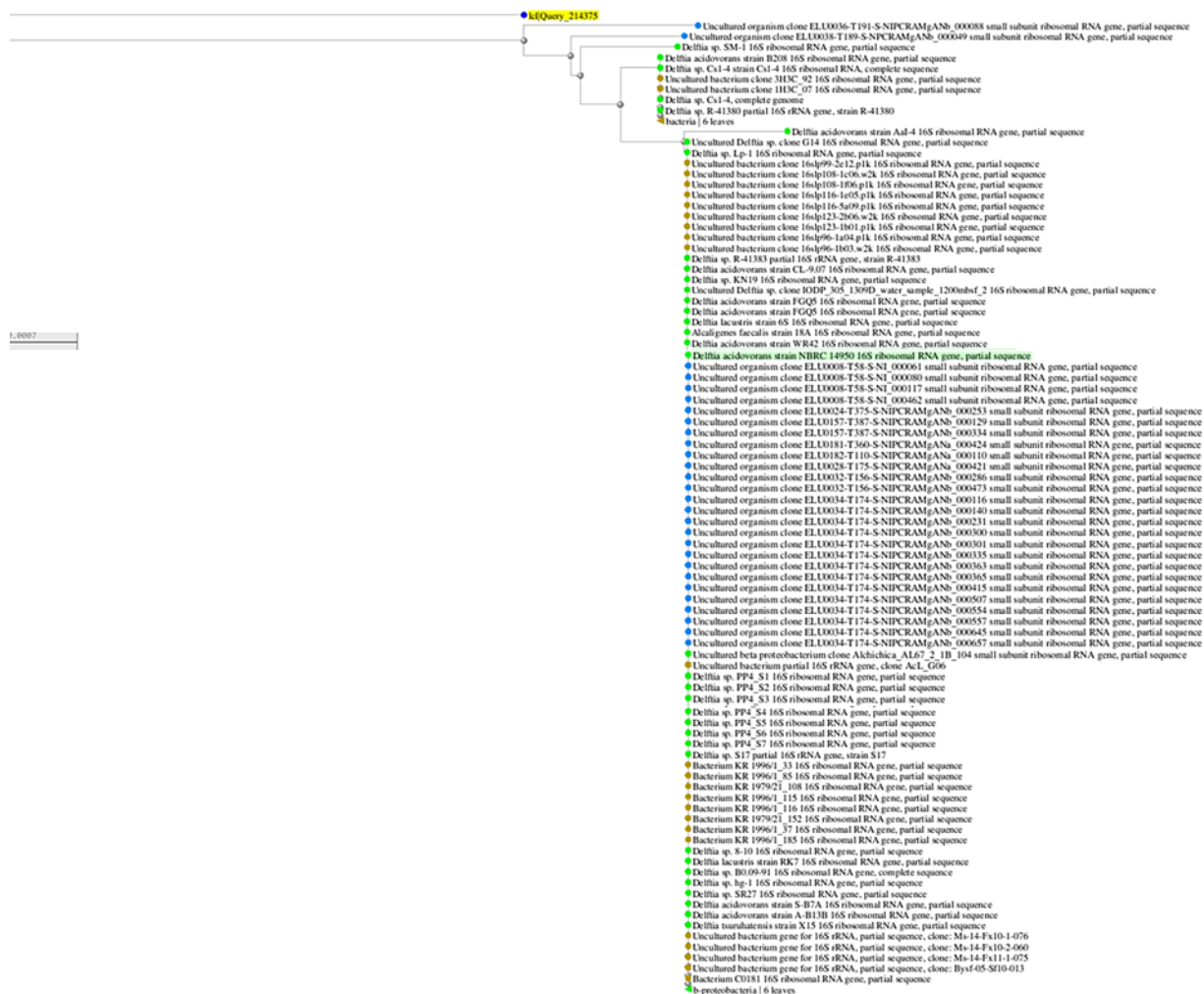
Delftia acidovorans strain B208 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ781879.1](#) Length: 1421 Number of Matches: 1

Range 1: 428 to 1376 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1599 bits(1772)	0.0	927/950(98%)	3/950(0%)	Plus/Minus	
Query 1	GGCGAGACCCGCTCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTAC	60			
Sbjct 1376	GGCGAGACCCGCTCCCATGGTGTGACGGGCGGTGTGTACAAGACCCGGGAACGTATTAC	1317			
Query 61	CGCGGCATGCTGATCCGCGATTACTAGCGATTCCGACTTCACGCAGTCGAGTTGCAGACT	120			
Sbjct 1316	CGCGGCATGCTGATCCGCGATTACTAGCGATTCCGACTTCACGCAGTCGAGTTGCAGACT	1257			
Query 121	GCGATCCGGACTACGACTGGTTTTATGGGATTAGCTCCCCCTCGCGGGATGGCAACCCTC	180			
Sbjct 1256	GCGATCCGGACTACGACTGGTTTTATGGGATTAGCTCCCCCTCGCGGGATGGCAACCCTC	1197			
Query 181	TGTACCAGCCATTGTATGACGTGTGTAGCCCCACCTATAAGGGCCATGAGGACTTGACGT	240			
Sbjct 1196	TGTACCAGCCATTGTATGACGTGTGTAGCCCCACCTATAAGGGCCATGAGGACTTGACGT	1137			
Query 241	CATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCTCATTAGAGTGCCCAACTAAATGTA	300			
Sbjct 1136	CATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCTCATTAGAGTGCCCAACTAAATGTA	1077			
Query 301	GCAACTAATGACAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAG	360			
Sbjct 1076	GCAACTAATGACAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAG	1017			
Query 361	CTGACGACAGCCATGCAGCACCTGTGTGCAGGTTCTCTTTCGAGCACGAATCCATCTCTG	420			
Sbjct 1016	CTGACGACAGCCATGCAGCACCTGTGTGCAGGTTCTCTTTCGAGCACGAATCCATCTCTG	957			
Query 421	GAAACTTCCTGCCATGTCAAAGGTGGGTAAGGTTTTTCGCGTTGCATCGAATTAAACCAC	480			
Sbjct 956	GAAACTTCCTGCCATGTCAAAGGTGGGTAAGGTTTTTCGCGTTGCATCGAATTAAACCAC	897			
Query 481	ATCATCCACCGCTTGTGCGGGTCCCCGTCAATTCTTTGAGTTTCAACCTTGCGGCCGTA	540			
Sbjct 896	ATCATCCACCGCTTGTGCGGGTCCCCGTCAATTCTTTGAGTTTCAACCTTGCGGCCGTA	837			
Query 541	CTCCCCAGGCGGTCAACTTCACGCGTTAGCTTCGTTACTGAGAAAATAATTCCCAACAA	600			
Sbjct 836	CTCCCCAGGCGGTCAACTTCACGCGTTAGCTTCGTTACTGAGAAAATAATTCCCAACAA	777			
Query 601	CCAGTTGACATCGTTTAGGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACG	660			
Sbjct 776	CCAGTTGACATCGTTTAGGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACG	717			
Query 661	CTTTCGTGCATGAGCGTCAGTACAGGTCCAGGGGATTGCCTTCGCCATCGGTGTTCTCTC	720			
Sbjct 716	CTTTCGTGCATGAGCGTCAGTACAGGTCCAGGGGATTGCCTTCGCCATCGGTGTTCTCTC	657			
Query 721	GCATATCTACGCATTTCACTGCTACACGCGGAATTCCATCCCCCTCTACCGTACTCTAAC	780			
Sbjct 656	GCATATCTACGCATTTCACTGCTACACGCGGAATTCCATCCCCCTCTACCGTACTCTAGC	597			
Query 781	CATGCAGTCACAAATGCAGTTCCAGGTTGAACCCGGGATTTCACATCTGTCTTAAATA	840			
Sbjct 596	CATGCAGTCACAAATGCAGTTCCAGGTTGAGCCCGGGATTTCACATCTGTCTTACATA	537			
Query 841	ACCCCTGGGACGCTTTACCCCCAAAAATTCC-AATAAAGCTTGCCCCCTACGTATTAC	899			
Sbjct 536	ACCGCTGCGACGCTTTACGCCAGTAATTCCGATTAACGCTTGACCCCTACGTATTAC	477			
Query 900	CCCGGCTGGTGGGGCGGTATTTAACCGG-GGTTATTCTTACGGAACCGTC	948			
Sbjct 476	CGCGGCTGCT-GGCACGTAGTTAGCCGGTGCTTATTCTTACGGTACCGTC	428			



Bacillus cereus GQ344804.1

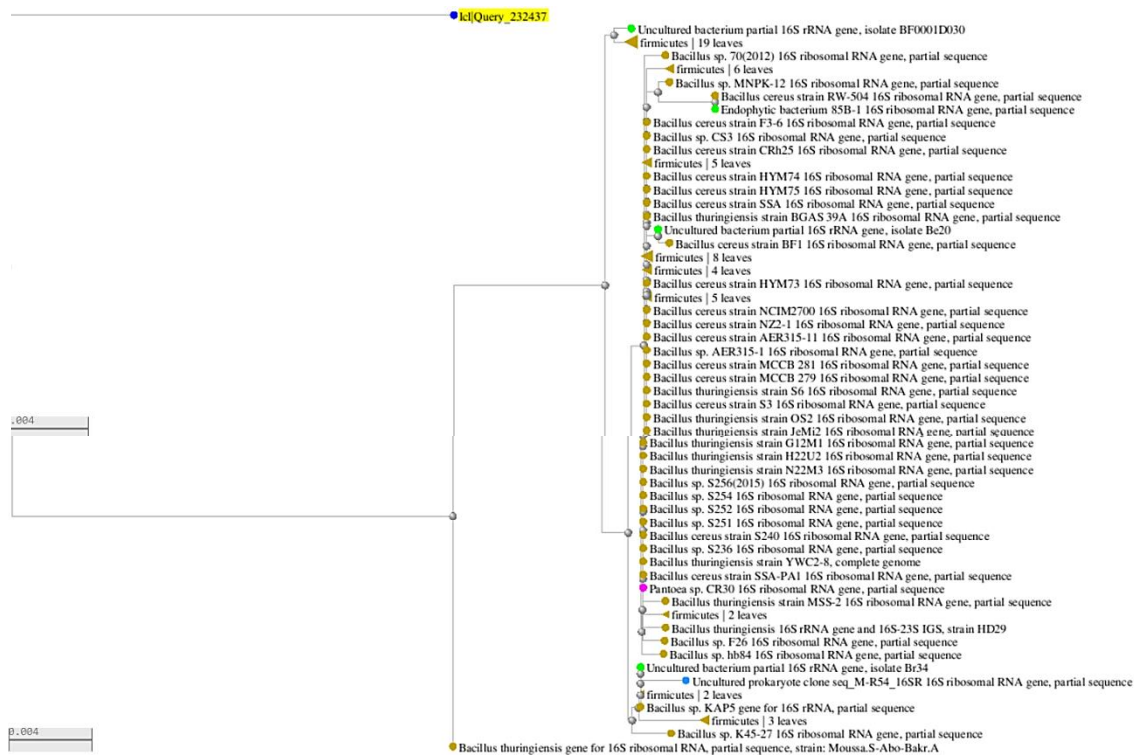
Bacillus cereus strain DC2 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|GQ344804.1|](#) Length: 1424 Number of Matches: 1

Range 1: 442 to 1382 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1397 bits(1548)	0.0	883/941(94%)	11/941(1%)	Plus/Minus
Query 1	CAAACCTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCA	60		
Sbjct 1382	CAAACCTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCACCGCGGCA	1323		
Query 61	TGCTGATCCGCGATTACTAGCGATTCCAACCTTCATGTAGGCGAGTTGCAGCCGACAATCC	120		
Sbjct 1322	TGCTGATCCGCGATTACTAGCGATTCCAACCTTCATGTAGGCGAGTTGCAGCCTACAATCC	1263		
Query 121	GAACTGAGAACGGTTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGTACCG	180		
Sbjct 1262	GAACTGAGAACGGTTTTATGAGATTAGCTCCACCTCGCGGTCTTGCAGCTCTTTGTACCG	1203		
Query 181	TCCATTGTATCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCC	240		
Sbjct 1202	TCCATTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGATGATTTGACGTCATCCCC	1143		
Query 241	ACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGCAACT	300		
Sbjct 1142	ACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCCAACTTAATGATGGCAACT	1083		
Query 301	AGAATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGATCTGAAC	360		
Sbjct 1082	AAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACGAGCTGACG	1023		
Query 361	ACAACCATGCACACCTGTCACTCTGCTCCCGAAGGACAAACCTATCTCTAGGGTTTTC	420		
Sbjct 1022	ACAACCATGCACACCTGTCACTCTGCTCCCGAAGGAGAAGCCCTATCTCTAGGGTTTTC	963		
Query 421	A-AGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCA	479		
Sbjct 962	AGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGAATTAAACCACATGCTCCA	903		
Query 480	CCGCTTGTGCGGGCCCCCGTCAATTCCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCCAG	539		
Sbjct 902	CCGCTTGTGCGGGCCCCCGTCAATTCCCTTTGAGTTTCAGCCTTGCGGCCGTACTCCCCAG	843		
Query 540	GCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCTCTAACACTTATCA	599		
Sbjct 842	GCGGAGTGCTTAATGCGTTAACTTCAGCACTAAAGGGCGGAAACCTCTAACACTTAGCA	783		
Query 600	CTCATCGTTTACGGCGTGGACTACCGGGATATCTAATCGTGTGTTGCTCCCCACTCTTTTCG	659		
Sbjct 782	CTCATCGTTTACGGCGTGGACTACCGGGATATCTAATCCTGTGTTGCTCCCCACGCTTTTCG	723		
Query 660	CGCCTCAGTGTCAATTACGGACCA-AAAGTCCCCTTCCCCACTGGTGTTCCTCCATATCT	718		
Sbjct 722	CGCCTCAGTGTCAATTACAGACCAGAAAGTGCCTTCGCCACTGGTGTTCCTCCATATCT	663		
Query 719	CTACCCCTTTTACCGCTACACATGAAATT-CACTTTCTCTTCTGCACTCAAGTCTCCCA	777		
Sbjct 662	CTACGCATTTACCGCTACACATGGAATTCACCTTTCTCTTCTGCACTCAAGTCTCCCA	603		
Query 778	GTTTCCAATGACCC-CCACGGTTGACCCG-GGGTTTTCACTTCAAACCTAA-AAACC-CC	833		
Sbjct 602	GTTTCCAATGACCCCTCCACGGTTGAGCCGTGGGCTTTCACATCAGACTTAAGAAACCACC	543		
Query 834	GGCCCGCCCTTTAC-CCCAAT-ATTCCAAGAAACCTTGCCCCCTACTTATTACC-Cggg	890		
Sbjct 542	TGCGCGCGCTTTACGCCCAATAATTCCGGATAACGCTTGCCACCTACGTATTACCGCGGC	483		
Query 891	ggggggCCCCGAATTTACCGGGGTTTTTCTGG-TAGGAACC	930		
Sbjct 482	TGCTGGCACGTAGTTAGCCGTGGCTTTTTCTGGTTAGGTACC	442		



10- The phylogenetic analysis of bacteria isolated from used toothbrushes samples

Roseomonas mucosa KF247232.1

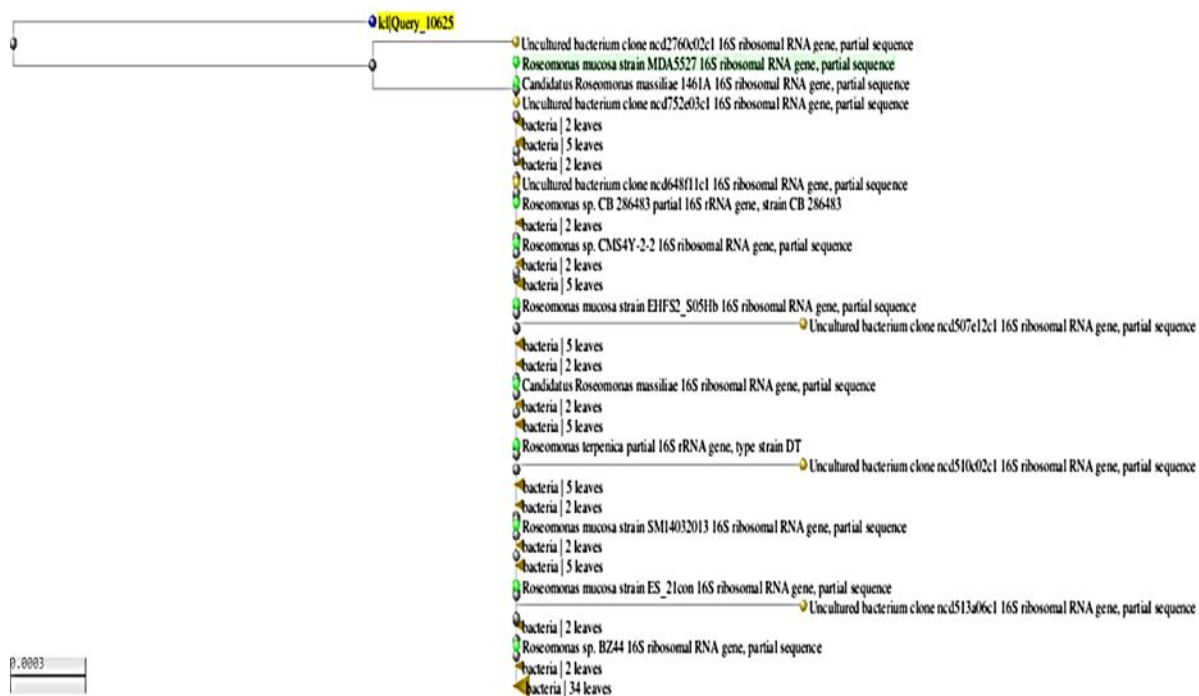
Roseomonas mucosa strain SM14032013 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KF247232.1|](#) Length: 1455 Number of Matches: 1

Range 1: 102 to 983 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1572 bits(1742)	0.0	879/883(99%)	1/883(0%)	Plus/Plus	
Query	1	GGACGGGTGAGTAACGCGTAGGAACGTGTCTGAGGTGGGGGACAACCCCGGGAACTGG			60
Sbjct	102	GGACGGGTGAGTAACGCGTAGGAACGTGTCTGAGGTGGGGGACAACCCCGGGAACTGG			161
Query	61	GGCTAATACCGCATATGGGCTGAGGCCCAAAGCCGAGAGGCGCCTTTGGAGCGGCCTGCG			120
Sbjct	162	GGCTAATACCGCATATGGGCTGAGGCCCAAAGCCGAGAGGCGCCTTTGGAGCGGCCTGCG			221
Query	121	TCCGATTAGGTAGTTGGTGGGGTAAAGGCCTACCAAGCCTGCGATCGGTAGCTGGTCTGA			180
Sbjct	222	TCCGATTAGGTAGTTGGTGGGGTAAAGGCCTACCAAGCCTGCGATCGGTAGCTGGTCTGA			281
Query	181	GAGGACGACCAGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGT			240
Sbjct	282	GAGGACGACCAGCCACACTGGGACTGAGACACGGCCCAGACTCCTACGGGAGGCAGCAGT			341
Query	241	GGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCAATGCCGCGTGGGTGAAGAAGGT			300
Sbjct	342	GGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCAATGCCGCGTGGGTGAAGAAGGT			401
Query	301	CTTCGGATCGTAAAGCCCTTTTCGACGGGGACGATGATGACGGTACCCGTAGAAGAAGCCC			360
Sbjct	402	CTTCGGATCGTAAAGCCCTTTTCGACGGGGACGATGATGACGGTACCCGTAGAAGAAGCCC			461
Query	361	CGGCTAACTTCGTGCCAGCAGCCGCGGTAATACGAAGGGGGCTAGCGTTGCTCGGAATTA			420
Sbjct	462	CGGCTAACTTCGTGCCAGCAGCCGCGGTAATACGAAGGGGGCTAGCGTTGCTCGGAATTA			521
Query	421	CTGGGCGTAAAGGGCGCGTAGGCGGGCGGCCAAGTCAGGCGTGAAATTCCTGGGCTCAAC			480
Sbjct	522	CTGGGCGTAAAGGGCGCGTAGGCGGGCGGCCAAGTCAGGCGTGAAATTCCTGGGCTCAAC			581
Query	481	CTGGGGACTGCGCTTGATACTGGGTTGCTTGAGGATGGAAGAGGCTCGTGGAATTCACAG			540
Sbjct	582	CTGGGGACTGCGCTTGATACTGGGTTGCTTGAGGATGGAAGAGGCTCGTGGAATTCACAG			641
Query	541	TGTAGAGGTGAAATTCGTAGATATTGGGAAGAACACCGGTGGCGAAGGCGGCGAGCTGGT			600
Sbjct	642	TGTAGAGGTGAAATTCGTAGATATTGGGAAGAACACCGGTGGCGAAGGCGGCGAGCTGGT			701
Query	601	CCATTACTGACGCTGAGGCGCGACAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAG			660
Sbjct	702	CCATTACTGACGCTGAGGCGCGACAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAG			761
Query	661	TCCACGCCGTAAACGATGTGCGCTGGATGTTGGGGCCCATAGGGTCTCAGTGTCGTAGCC			720
Sbjct	762	TCCACGCCGTAAACGATGTGCGCTGGATGTTGGGGCCCATAGGGTCTCAGTGTCGTAGCC			821
Query	721	AACGCGGTAAGCGCACCGCCTGGGGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGA			780
Sbjct	822	AACGCGGTAAGCGCACCGCCTGGGGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGA			881
Query	781	CGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGCAAAACCTT			840
Sbjct	882	CGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGCAGAA-CCTT			940
Query	841	ACCAGCCCTTGACATGGTCACGACCGGTCCAAAAATGGACTTT		883	
Sbjct	941	ACCAGCCCTTGACATGGTCACGACCGGTCCAGAGATGGACTTT		983	



Stenotrophomonas maltophilia LN890169.1

Stenotrophomonas maltophilia partial 16S rRNA gene, strain M83

Sequence ID: [emb|LN890169.1](#) Length: 1688 Number of Matches: 1

Range 1: 190 to 1127 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1665 bits(1846)	0.0	933/938(99%)	1/938(0%)	Plus/Plus
Query 1	TGGGTGGCGAGTGGCGGACGGGTGAGGAATACATCGGAATCTACTCTGTCGTGGGGGATA	60		
Sbjct 190	TGGGTGGCGAGTGGCGGACGGGTGAGGAATACATCGGAATCTACTCTGTCGTGGGGGATA	249		
Query 61	ACGTAGGGAAACTTACGCTAATACCGCATACGACCTACGGGTGAAAGCAGGGGATCTTCG	120		
Sbjct 250	ACGTAGGGAAACTTACGCTAATACCGCATACGACCTACGGGTGAAAGCAGGGGATCTTCG	309		
Query 121	GACCTTGC GCGATTGAATGAGCCGATGTCGGATTAGCTAGTTGGCGGGGTAAAGGCCAC	180		
Sbjct 310	GACCTTGC GCGATTGAATGAGCCGATGTCGGATTAGCTAGTTGGCGGGGTAAAGGCCAC	369		
Query 181	CAAGGCGACGATCCGTAGCTGGTCTGAGAGGATGATCAGCCACACTGGAAGTGAACACG	240		
Sbjct 370	CAAGGCGACGATCCGTAGCTGGTCTGAGAGGATGATCAGCCACACTGGAAGTGAACACG	429		
Query 241	GTCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGCAAGCCTGATC	300		
Sbjct 430	GTCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGCAAGCCTGATC	489		
Query 301	CAGCCATACCGCGTGGGTGAAGAAGGCCTTCGGGTTGTAAAGCCCTTTTGTGGGAAAGA	360		
Sbjct 490	CAGCCATACCGCGTGGGTGAAGAAGGCCTTCGGGTTGTAAAGCCCTTTTGTGGGAAAGA	549		
Query 361	AATCCAGCTGGCTAATACCCGGTTGGGATGACGGTACCCAAAGAATAAGCACCGGCTAAC	420		
Sbjct 550	AATCCAGCTGGCTAATACCCGGTTGGGATGACGGTACCCAAAGAATAAGCACCGGCTAAC	609		
Query 421	TTCGTGCCAGCAGCCGCGGTAATACGAAGGGTGCAAGCGTTACTCGGAATTACTGGGCGT	480		
Sbjct 610	TTCGTGCCAGCAGCCGCGGTAATACGAAGGGTGCAAGCGTTACTCGGAATTACTGGGCGT	669		
Query 481	AAAGCGTGCGTAGGTGGTTCGTTTAAAGTCCGTTGTGAAAGCCCTGGGCTCAACCTGGGAAC	540		
Sbjct 670	AAAGCGTGCGTAGGTGGTTCGTTTAAAGTCCGTTGTGAAAGCCCTGGGCTCAACCTGGGAAC	729		
Query 541	TGCAGTGGATACTGGGCGACTAGAGTGTGGTAGAGGGTAGCGGAATTCCTGGTGTAGCAG	600		
Sbjct 730	TGCAGTGGATACTGGGCGACTAGAGTGTGGTAGAGGGTAGCGGAATTCCTGGTGTAGCAG	789		
Query 601	TGAAATGCGTAGAGATCAGGAGGAACATCCATGGCGAAGGCAGCTACCTGGACCAACACT	660		
Sbjct 790	TGAAATGCGTAGAGATCAGGAGGAACATCCATGGCGAAGGCAGCTACCTGGACCAACACT	849		
Query 661	GACACTGAGGCACGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCC	720		
Sbjct 850	GACACTGAGGCACGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCC	909		
Query 721	CTAAACGATGCGAACTGGATGTTGGGTGCAATTTGGCACGCAGTATCGAAGCTAACGCGT	780		
Sbjct 910	CTAAACGATGCGAACTGGATGTTGGGTGCAATTTGGCACGCAGTATCGAAGCTAACGCGT	969		
Query 781	TAAATTTCGCCGCTGGGGAGTACGGTCGCAAGACTGAAACTC-AAGGAATTGACGGGGGC	839		
Sbjct 970	TAAATTTCGCCGCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGC	1029		
Query 840	CCGCACAAGCGGTGGAGTATGTGGTTTAATTCGATGCAACGCGAAAAACCTTACCTGGCC	899		
Sbjct 1030	CCGCACAAGCGGTGGAGTATGTGGTTTAATTCGATGCAACGCGAAGAACCTTACCTGGCC	1089		
Query 900	TTGACATGTCGAGAACTTTCCAAAAATGGATGGGGGCC	937		
Sbjct 1090	TTGACATGTCGAGAACTTTCCAGAGATGGATGGGTGCC	1127		



Pseudomonas aeruginosa KR815846.1

Pseudomonas aeruginosa strain Pse12 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KR815846.1](#) Length: 1444 Number of Matches: 1

Range 1: 34 to 999 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1685 bits(1868)	0.0	955/966(99%)	5/966(0%)	Plus/Plus
Query 1	TGGATTTCNGCGGCGGACGGGTGAGTAATGCCTAGGAATCTGCCTGGTAGTGGGGGATAAC	60		
Sbjct 34	TGGATTTCAGCGGCGGACGGGTGAGTAATGCCTAGGAATCTGCCTGGTAGTGGGGGATAAC	93		
Query 61	GTCCGGAACGGGCGCTAATACCGCATACTGCTGAGGGAGAAAAGTGGGGGATCTTCGGA	120		
Sbjct 94	GTCCGGAACGGGCGCTAATACCGCATACTGCTGAGGGAGAAAAGTGGGGGATCTTCGGA	153		
Query 121	CCTCAGCTATCAGATGAGCCTAGGTCGGATTAGCTAGTTGGTGGGGTAAAGGCCTACCA	180		
Sbjct 154	CCTCAGCTATCAGATGAGCCTAGGTCGGATTAGCTAGTTGGTGGGGTAAAGGCCTACCA	213		
Query 181	AGGCGACGATCCGTAACCTGGTCTGAGAGGATGATCAGTCACACTGGAACGAGACACGGT	240		
Sbjct 214	AGGCGACGATCCGTAACCTGGTCTGAGAGGATGATCAGTCACACTGGAACGAGACACGGT	273		
Query 241	CCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGAAAAGCCTGATCCA	300		
Sbjct 274	CCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGAAAAGCCTGATCCA	333		
Query 301	GCCATGCCGCGTGTGTGAAGAAGGTCTTCGGATTGTAAAGCACTTTAAGTTGGGAGGAAG	360		
Sbjct 334	GCCATGCCGCGTGTGTGAAGAAGGTCTTCGGATTGTAAAGCACTTTAAGTTGGGAGGAAG	393		
Query 361	GGCAGTAAGTTAATACCTTGCTGTTTTGACGTTACCAACAGAATAAGCACCGGCTAACTT	420		
Sbjct 394	GGCAGTAAGTTAATACCTTGCTGTTTTGACGTTACCAACAGAATAAGCACCGGCTAACTT	453		
Query 421	CGTGCCAGCAGCCGCGGTAATACGAAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAA	480		
Sbjct 454	CGTGCCAGCAGCCGCGGTAATACGAAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAA	513		
Query 481	AGCGCGCGTAGGTGGTTTTCAGCAAGTTGGATGTGAAATCCCCGGGCTCAACCTGGGAAC	540		
Sbjct 514	AGCGCGCGTAGGTGGTTTTCAGCAAGTTGGATGTGAAATCCCCGGGCTCAACCTGGGAAC	573		
Query 541	CATCCAAACTACTGAGCTAGAGTACGGTAGAGGGTGGTGGAAATTCCTGTGTAGCGGTG	600		
Sbjct 574	CATCCAAACTACTGAGCTAGAGTACGGTAGAGGGTGGTGGAAATTCCTGTGTAGCGGTG	633		
Query 601	AAATGCGTAGATATAGGAAGGAACACCAAGTGGCGAAGGCGACCACTGGACTGATACTGA	660		
Sbjct 634	AAATGCGTAGATATAGGAAGGAACACCAAGTGGCGAAGGCGACCACTGGACTGATACTGA	693		
Query 661	CACTGAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGT	720		
Sbjct 694	CACTGAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGT	753		
Query 721	AAACGATGTCGACTAGCCGTTGGGATCCTTGAGATCTTAGTGGCGCAGCTAACGCGATAA	780		
Sbjct 754	AAACGATGTCGACTAGCCGTTGGGATCCTTGAGATCTTAGTGGCGCAGCTAACGCGATAA	813		
Query 781	GTCGACCGCCTGGGGAGTACGGCCGCAAGGTTAAACTCAAATGAATTGACGGGGGCCG	840		
Sbjct 814	GTCGACCGCCTGGGGAGTACGGCCGCAAGGTTAAACTCAAATGAATTGACGGGGGCCG	873		
Query 841	CACAAGCGGTGGAGCATGTGGTTTAAATTCTAAGCAACGCGAAAAACCTTACCTGGGCCTT	900		
Sbjct 874	CACAAGCGGTGGAGCATGTGGTTTAAATTCTAAGCAACGCGAAAAACCTTACCTGGGCCTT	933		
Query 901	GACATGCTGAAAACCTTCCAAAAATGGATTGGGGCCTTC--GGAATC-AAACCCAGG--	955		
Sbjct 934	GACATGCTGAGAACTTCCAAAGATGGATTGGTGCCTCCGGGAACCAAAACACAGGGT	993		
Query 956	GCTGCA 961			
Sbjct 994	GCTGCA 999			



Leclercia adecarboxylata KT899848.1

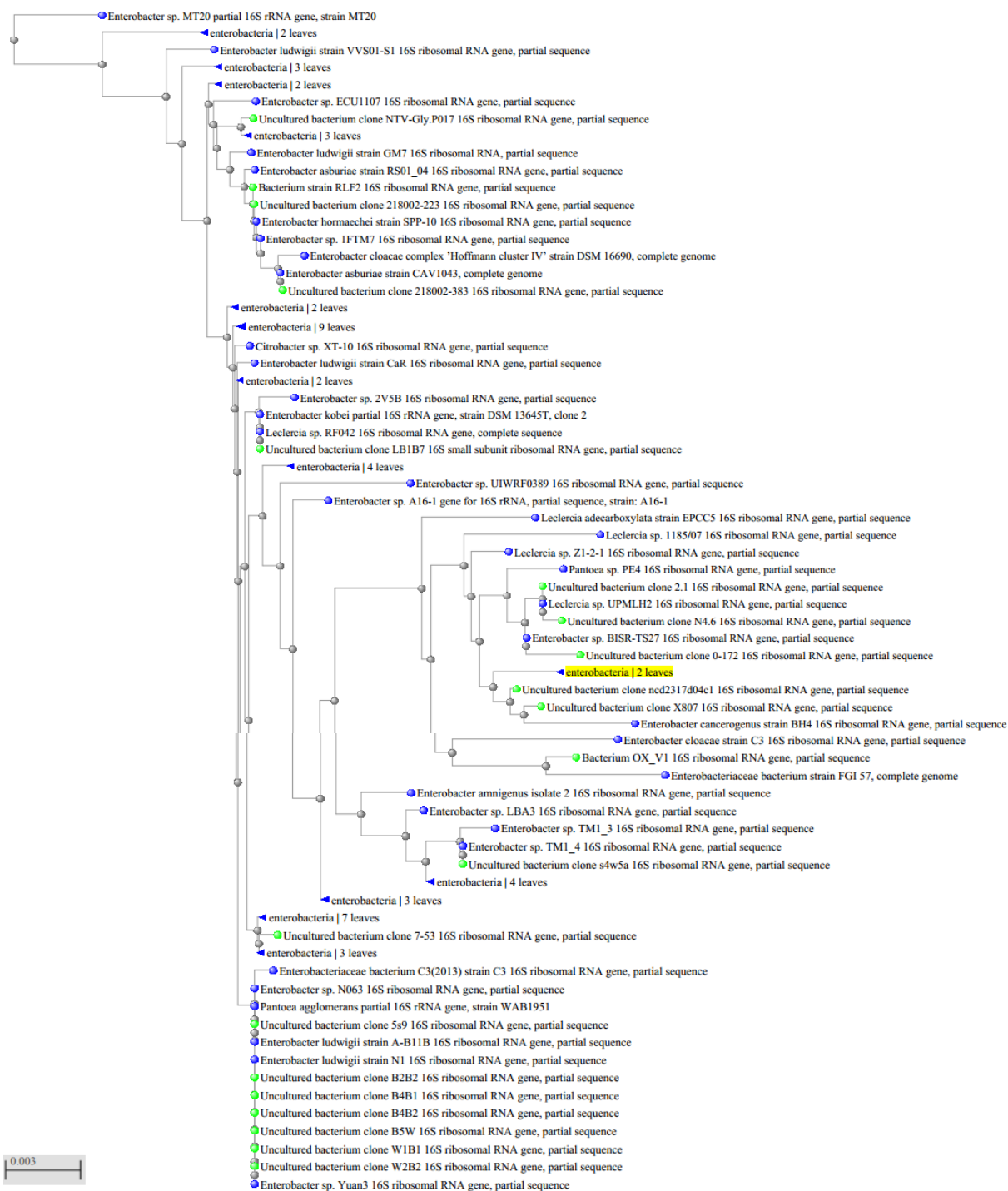
Leclercia adecarboxylata strain MCCB 331 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT899848.1](#) Length: 1323 Number of Matches: 1

Range 1: 1 to 905 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1597 bits(1770)	0.0	897/905(99%)	1/905(0%)	Plus/Plus	
Query 6	GACGAGTGGCGGACGGGTGAGTAATGTCTGGGAAACTGCCTGATGGAGGGGGATAACTAC				65
Sbjct 1	GACGAGTGGCGGACGGGTGAGTAATGTCTGGGAAACTGCCTGATGGAGGGGGATAACTAC				60
Query 66	TGGAAACGGTAGCTAATACCGCATAAYGTCGCAAGACCAAAGAGGGGGACCTTCGGGCCT				125
Sbjct 61	TGGAAACGGTAGCTAATACCGCATAATGTCTGCAAGACCAAAGAGGGGGACCTTCGGGCCT				120
Query 126	CTTGCCATCGGATGTGCCAGATGGGATTAGCTAGTAGGTGGGGTAATGGCTCACCTAGG				185
Sbjct 121	CTTGCCATCGGATGTGCCAGATGGGATTAGCTAGTAGGTGGGGTAATGGCTCACCTAGG				180
Query 186	CGACGATCCCTAGCTGGTCTGAGAGGATGACCAGCCACACTGGAAGTGAAGACACGGTCCA				245
Sbjct 181	CGACGATCCCTAGCTGGTCTGAGAGGATGACCAGCCACACTGGAAGTGAAGACACGGTCCA				240
Query 246	GACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCC				305
Sbjct 241	GACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACAATGGGCGCAAGCCTGATGCAGCC				300
Query 306	ATGCCGCGTGTATGAAGAAGGCCTTCGGGTTGTAAAGTACTTTTCAGCGAGGAGGAAGGCA				365
Sbjct 301	ATGCCGCGTGTATGAAGAAGGCCTTCGGGTTGTAAAGTACTTTTCAGCGAGGAGGAAGGCG				360
Query 366	TTGTGGTTAATAACCGCAGTGATTGACGTTACTCGCAGAAGAAGCACCGGCTAACTCCGT				425
Sbjct 361	TTGTGGTTAATAACCGCAGTGATTGACGTTACTCGCAGAAGAAGCACCGGCTAACTCCGT				420
Query 426	GCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGC				485
Sbjct 421	GCCAGCAGCCGCGGTAATACGGAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGC				480
Query 486	GCACGCAGGCGGTCTGTTAAGTCAGATGTGAAATCCCCGGGCTCAACCTGGGAACTGCAT				545
Sbjct 481	GCACGCAGGCGGTCTGTTAAGTCAGATGTGAAATCCCCGGGCTCAACCTGGGAACTGCAT				540
Query 546	TTGAAACTGGCAGGCTTGAGTCTTGTAGAGGGGGGTAGAATTCCAGGTGTAGCGGTGAAA				605
Sbjct 541	TTGAAACTGGCAGGCTTGAGTCTTGTAGAGGGGGGTAGAATTCCAGGTGTAGCGGTGAAA				600
Query 606	TGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCGGCCCCCTGGACAAAGACTGACGC				665
Sbjct 601	TGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCGGCCCCCTGGACAAAGACTGACGC				660
Query 666	TCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAA				725
Sbjct 661	TCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAA				720
Query 726	CGATGTGCACTTGGAGGTTGTGCCCTTGAGGCGTGGCTTCCGGAGCTAACGCGTTAAGTC				785
Sbjct 721	CGATGTGCACTTGGAGGTTGTGCCCTTGAGGCGTGGCTTCCGGAGCTAACGCGTTAAGTC				780
Query 786	GACCGCTGGGGAGTACGGCCGCAAGGTTAAACTCAAATGAATTGACGGGGGCCCCG-CA				844
Sbjct 781	GACCGCTGGGGAGTACGGCCGCAAGGTTAAACTCAAATGAATTGACGGGGGCCCCGCCA				840
Query 845	CAAGCGGGGGAGCATGTGGTTTAAATTCATGCAACGCGAAAAACCTTACCTACTCTTGAC				904
Sbjct 841	CAAGCGGTGGAGCATGTGGTTTAAATTCGATGCAACGCGAAGAACCTTACCTRCTCTTGAC				900
Query 905	ATCCA	909			
Sbjct 901	ATCCA	905			



Enterobacter asburiae EU239468.1

Enterobacter asburiae strain XJUH-5 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|EU239468.1](#) Length: 1446 Number of Matches: 1

Range 1: 504 to 1394 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1573 bits(1744)	0.0	884/891(99%)	1/891(0%)	Plus/Minus
Query 1	TTTGTCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTTCAC	60		
Sbjct 1394	TTTGTCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTTCAC	1335		
Query 61	CGTAGCATTCTGATCTACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGACT	120		
Sbjct 1334	CGTAGCATTCTGATCTACGATTACTAGCGATTCCGACTTCATGGAGTCGAGTTGCAGACT	1275		
Query 121	CCAATCCGGACTACGACGCACCTTTATGAGGTCCGCTTGCTCTCGCGAGGTCGCTTCTCTT	180		
Sbjct 1274	CCAATCCGGACTACGACGCACCTTTATGAGGTCCGCTTGCTCTCGCGAGGTCGCTTCTCTT	1215		
Query 181	TGTATGCGCCATTGTAGCACGTGTGTAGCCCTACTCGTAAGGGCCATGATGACTTGACGT	240		
Sbjct 1214	TGTATGCGCCATTGTAGCACGTGTGTAGCCCTACTCGTAAGGGCCATGATGACTTGACGT	1155		
Query 241	CATCCCCACCTTCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGGCCTAACCGC	300		
Sbjct 1154	CATCCCCACCTTCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCGGCCTAACCGC	1095		
Query 301	TGGCAACAAAGGATAAAGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTACAAACACG	360		
Sbjct 1094	TGGCAACAAAGGATAAAGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTACAAACACG	1035		
Query 361	AGCTGACGACAGCCATGCAGCACCTGTCTCAGAGTTCCCGAAGGCACCAAGCCATCTCTG	420		
Sbjct 1034	AGCTGACGACAGCCATGCAGCACCTGTCTCAGAGTTCCCGAAGGCACCAATCCATCTCTG	975		
Query 421	GCAAGTTCTCTGGATGTCAAGAGTAGGTAAGGTTCTTCGCGTTGCATCGAATTAAACCAC	480		
Sbjct 974	GAAAGTTCTCTGGATGTCAAGAGTAGGTAAGGTTCTTCGCGTTGCATCGAATTAAACCAC	915		
Query 481	ATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCAATTTGAGTTTTAACCTTGCGGCCGTA	540		
Sbjct 914	ATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCAATTTGAGTTTTAACCTTGCGGCCGTA	855		
Query 541	CTCCCCAGGCGGTCTGACTTAACGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACT	600		
Sbjct 854	CTCCCCAGGCGGTCTGACTTAACGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACT	795		
Query 601	CCAAGTCGACATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCAC	660		
Sbjct 794	CCAAGTCGACATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCAC	735		
Query 661	GCTTTTCGCACCTGAGCGTCAGTCTTTGTCCAGGGGGCCGCTTCGCCACCGGTATTTCCTC	720		
Sbjct 734	GCTTTTCGCACCTGAGCGTCAGTCTTTGTCCAGGGGGCCGCTTCGCCACCGGTATTTCCTC	675		
Query 721	CAGATCTCTACGCATTTACCGCTACACCTGGAATTCTACCCCCCTCTACAAGACTCAAG	780		
Sbjct 674	CAGATCTCTACGCATTTACCGCTACACCTGGAATTCTACCCCCCTCTACAAGACTCTAG	615		
Query 781	CCTGCCAGTTTCAAATGCAGTTCCC-NGTTGAGCCCGGGGATTTACATCTGACTTAACA	839		
Sbjct 614	CCTGCCAGTTTCAAATGCAGTTCCCAGGTTGAGCCCGGGGATTTACATCTGACTTGACA	555		
Query 840	GACCGCTGCGTGCCTTTACGCCCAGTAATTCGATTAAACGCTTGACCCC	890		
Sbjct 554	GACCGCTGCGTGCCTTTACGCCCAGTAATTCGATTAAACGCTTGACCCC	504		



Candidatus Roseomonas massiliae KT321690.1

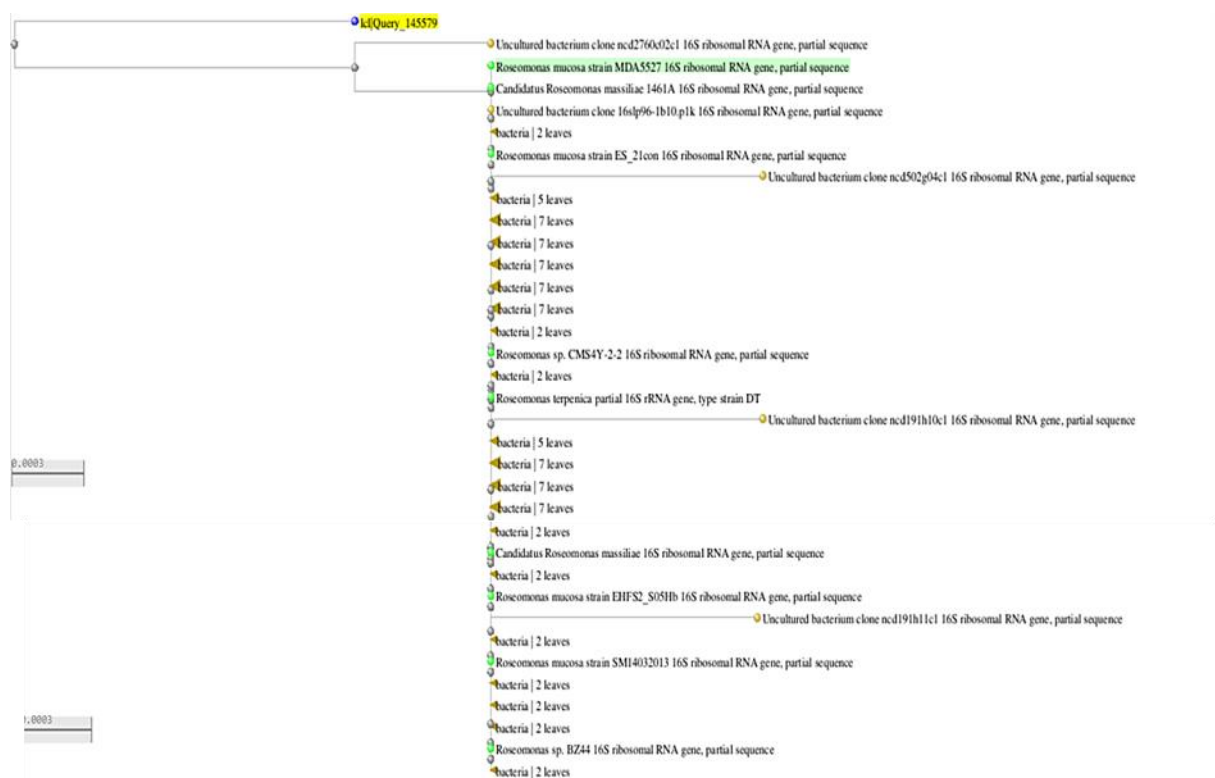
Candidatus Roseomonas massiliae 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT321690.1](#) Length: 1443 Number of Matches: 1

Range 1: 74 to 974 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1613 bits(1788)	0.0	898/901(99%)	0/901(0%)	Plus/Plus
Query 1	GGACGGGTGAGTAACGCGTAGGAACGTGTCCTGAGGTGGGGGACAACCCCGGGAAACTGG	60		
Sbjct 74	GGACGGGTGAGTAACGCGTAGGAACGTGTCCTGAGGTGGGGGACAACCCCGGGAAACTGG	133		
Query 61	GGCTAATACCGCATATGGGCTGAGGCCCAAAGCCGAGAGGCGCCTTTGGAGCGGCCTGCG	120		
Sbjct 134	GGCTAATACCGCATATGGGCTGAGGCCCAAAGCCGAGAGGCGCCTTTGGAGCGGCCTGCG	193		
Query 121	TCCGATTAGGTAGTTGGTGGGGTAAAGGCCTACCAAGCCTGCGATCGGTAGCTGGTCTGA	180		
Sbjct 194	TCCGATTAGGTAGTTGGTGGGGTAAAGGCCTACCAAGCCTGCGATCGGTAGCTGGTCTGA	253		
Query 181	GAGGACGACGACGACCACTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCAGT	240		
Sbjct 254	GAGGACGACGACGACCACTGGGACTGAGACACGGCCAGACTCCTACGGGAGGCAGCAGT	313		
Query 241	GGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCAATGCCGCGTGGGTGAAGAAGGT	300		
Sbjct 314	GGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCAATGCCGCGTGGGTGAAGAAGGT	373		
Query 301	CTTCGGATCGTAAAGCCCTTTTCGACGGGGACGATGATGACGGTACCCGTAGAAGAAGCCC	360		
Sbjct 374	CTTCGGATCGTAAAGCCCTTTTCGACGGGGACGATGATGACGGTACCCGTAGAAGAAGCCC	433		
Query 361	CGGCTAACTTCGTGCCAGCAGCCGCGGTAATACGAAGGGGGCTAGCGTTGCTCGGAATTA	420		
Sbjct 434	CGGCTAACTTCGTGCCAGCAGCCGCGGTAATACGAAGGGGGCTAGCGTTGCTCGGAATTA	493		
Query 421	CTGGGCGTAAAGGGCGCGTAGGCGGCGGCCAAGTCAGGCGTGAAATTCCTGGGCTCAAC	480		
Sbjct 494	CTGGGCGTAAAGGGCGCGTAGGCGGCGGCCAAGTCAGGCGTGAAATTCCTGGGCTCAAC	553		
Query 481	CTGGGGACTGCGCTTGATACTGGGTTGCTTGAGGATGGAAGAGGCTCGTGGAATTCCTAG	540		
Sbjct 554	CTGGGGACTGCGCTTGATACTGGGTTGCTTGAGGATGGAAGAGGCTCGTGGAATTCCTAG	613		
Query 541	TGTAGAGGTGAAATTCGTAGATATTGGGAAGAACACCGGTGGCGAAGGCGGCGAGCTGGT	600		
Sbjct 614	TGTAGAGGTGAAATTCGTAGATATTGGGAAGAACACCGGTGGCGAAGGCGGCGAGCTGGT	673		
Query 601	CCATTACTGACGCTGAGGCGCGACAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAG	660		
Sbjct 674	CCATTACTGACGCTGAGGCGCGACAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAG	733		
Query 661	TCCACGCCGTAAACGATGTGCGCTGGATGTTGGGGCCCATAGGGTCTCAGTGTCTAGGCC	720		
Sbjct 734	TCCACGCCGTAAACGATGTGCGCTGGATGTTGGGGCCCATAGGGTCTCAGTGTCTAGGCC	793		
Query 721	AACGCGGTAAAGCGCACCGCCTGGGGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGA	780		
Sbjct 794	AACGCGGTAAAGCGCACCGCCTGGGGAGTACGGCCGCAAGGTTGAAACTCAAAGGAATTGA	853		
Query 781	CGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAAATTCGAAGCAACGCGCANAACCTTA	840		
Sbjct 854	CGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAAATTCGAAGCAACGCGCAGAACCTTA	913		
Query 841	CCAGCCCTTGACATGGTACGACCGGTCCAAAAATGGACTTTCTAGCAATAGGCGTGAT	900		
Sbjct 914	CCAGCCCTTGACATGGTACGACCGGTCCAGAGATGGACTTTCTAGCAATAGGCGTGAT	973		
Query 901	G 901			
Sbjct 974	G 974			



Pseudomonas parafulva KT758848.1

Pseudomonas parafulva strain YAB-1 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KT758848.1|](#) Length: 1401 Number of Matches: 1

Range 1: 39 to 980 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1651 bits(1830)	0.0	933/942(99%)	2/942(0%)	Plus/Plus
Query 1	AGCGGCGGACGGGTGAGTAATGCCTAGGAATCTGCCTGGTAGTGGGGGACAACGTTTCGA	60		
Sbjct 39	AGCGGCGGACGGGTGAGTAATGCCTAGGAATCTGCCTGGTAGTGGGGGACAACGTTTCGA	98		
Query 61	AAGGAACGCTAATACCGCATACGTCCTACGGGAGAAAGCAGGGGACCTTCGGGCCTTGCG	120		
Sbjct 99	AAGGAACGCTAATACCGCATACGTCCTACGGGAGAAAGCAGGGGACCTTCGGGCCTTGCG	158		
Query 121	CTATCAGATGAGCCTAGGTCGGATTAGCTTGTGGTGAGGTAATGGCTCACCAAGGCGAC	180		
Sbjct 159	CTATCAGATGAGCCTAGGTCGGATTAGCTTGTGGTGAGGTAATGGCTCACCAAGGCGAC	218		
Query 181	GATCCGTAAGTGGTCTGAGAGGATGATCAGTCACACTGGAAGTGGAGACACGGTCCAGACT	240		
Sbjct 219	GATCCGTAAGTGGTCTGAGAGGATGATCAGTCACACTGGAAGTGGAGACACGGTCCAGACT	278		
Query 241	CCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCCATGC	300		
Sbjct 279	CCTACGGGAGGCAGCAGTGGGGAATATTGGACAATGGGCGAAAGCCTGATCCAGCCATGC	338		
Query 301	CGCGTGTGTGAAGAAGGTCTTCGGATTGTAAAGCACTTTAAGTTGGGAGGAAGGGTTGTA	360		
Sbjct 339	CGCGTGTGTGAAGAAGGTCTTCGGATTGTAAAGCACTTTAAGTTGGGAGGAAGGGTTGTA	398		
Query 361	GATTAATACTCTGCAATTTTGACGTTACCGACAGAATAAGCACCGGCTAACTCTGTGCCA	420		
Sbjct 399	GATTAATACTCTGCAATTTTGACGTTACCGACAGAATAAGCACCGGCTAACTCTGTGCCA	458		
Query 421	GCAGCCGCGGTAATACAGAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGCGCGC	480		
Sbjct 459	GCAGCCGCGGTAATACAGAGGGTGCAAGCGTTAATCGGAATTACTGGGCGTAAAGCGCGC	518		
Query 481	GTAGGTGGTTTGTAAAGTTGGATGTGAAAGCCCCGGGCTCAACCTGGGAAGTGCATCCAA	540		
Sbjct 519	GTAGGTGGTTTGTAAAGTTGGATGTGAAAGCCCCGGGCTCAACCTGGGAAGTGCATCCAA	578		
Query 541	AACTGGCAAGCTAGAGTACGGTAGAGGGTGGTGGAAATTCCTGTGTAGCGGTGAAATGCG	600		
Sbjct 579	AACTGGCAAGCTAGAGTACGGTAGAGGGTGGTGGAAATTCCTGTGTAGCGGTGAAATGCG	638		
Query 601	TAGATATAGGAAGGAACACCAAGTGGCGAAGGCGACCACCTGGACTGATACTGACACTGAG	660		
Sbjct 639	TAGATATAGGAAGGAACACCAAGTGGCGAAGGCGACCACCTGGACTGATACTGACACTGAG	698		
Query 661	GTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGAT	720		
Sbjct 699	GTGCGAAAGCGTGGGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGAT	758		
Query 721	GTCAACTAGCCGTTGGAATCCTTGAGATTTTAGTGGCGCAGCTAACGCATTAAGTTGACC	780		
Sbjct 759	GTCAACTAGCCGTTGGAATCCTTGAGATTTTAGTGGCGCAGCTAACGCATTAAGTTGACC	818		
Query 781	GCCTGGGGAGTACGGCCGCAAGGTT-AAACTCAAATGAATTGACGGGGGCCCGCACAAGC	839		
Sbjct 819	GCCTGGGGAGTACGGCCGCAAGGTTAAACTCAAATGAATTGACGGGGGCCCGCACAAGC	878		
Query 840	GGTGGAGCATGTGGTTTAATTCGAAACAACGCGAAAAACCTTACCAGGCCTTGACATGCA	899		
Sbjct 879	GGTGGAGCATGTGGTTTAATTCGAAACAACGCGAAGAACCTTACCAGGCCTTGACATGCA	938		
Query 900	AAAAACTTTCCAAAAATGGATTGGGGCCTTCGGG-ACTCTGA	940		
Sbjct 939	GAGAACTTTCCAGAGATGGATTGGTGCCTTCGGGAACCTCTGA	980		

	Pseudomonas sp. 4 16S ribosomal RNA gene, partial sequence	
	Pseudomonas parafulva strain AJ 2129 16S ribosomal RNA gene, partial sequence	
	Pseudomonas parafulva strain AJ 2129 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. F24 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone nc262606c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain EX4 16S ribosomal RNA gene, partial sequence	
	Pseudomonas putida strain CLF2014-1 16S ribosomal RNA gene, partial sequence	
	bacteria 2 leaves	
	ε-proteobacteria 4 leaves	
	Pseudomonas fulva strain 1Y1103 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone nbw621f12c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. DP108 16S ribosomal RNA gene, partial sequence	Pseudomonas fulva strain i7_MX 16S ribosomal RNA gene, partial sequence
	Pseudomonas sp. NCCP-562 gene for 16S ribosomal RNA, partial sequence	
	bacteria 2 leaves	
	Pseudomonas sp. NCCP-553 gene for 16S ribosomal RNA, partial sequence	
	Uncultured bacterium clone nb18d11 16S ribosomal RNA gene, partial sequence	
	ε-proteobacteria 4 leaves	
	Pseudomonas fulva gene for 16S rRNA, partial sequence, strain: NBRC 16638	
	Pseudomonas fulva 67 16S ribosomal RNA gene, partial sequence	Uncultured bacterium clone nc137a07c1 16S ribosomal RNA gene, partial sequence
	Endophytic bacterium SY807 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. NCCP-571 gene for 16S ribosomal RNA, partial sequence	
	Pseudomonas sp. NLW-1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain SW32 16S ribosomal RNA gene, partial sequence	Uncultured bacterium clone nc2661b11c1 16S ribosomal RNA gene, partial sequence
	Pseudomonas fulva strain MX35 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone 654955 16S ribosomal RNA gene, partial sequence	
	Pseudomonas parafulva strain NBRC 16636 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone nc42803d07c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain BSW6 16S ribosomal RNA gene, partial sequence	Pseudomonas sp. MII-135 16S rRNA gene, strain MII-135
	Pseudomonas sp. enrichment culture clone 8 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. 3zhy partial 16S rRNA gene, strain 3zhy	
	Uncultured bacterium clone nc42564f11c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. MG-2011-7-BJ partial 16S rRNA gene, strain 7, isolate BJ	
	Uncultured bacterium clone nb122b10 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. DP109B 16S ribosomal RNA gene, partial sequence	Pseudomonas fulva strain iP1_MX 16S ribosomal RNA gene, partial sequence
	Pseudomonas sp. NCCP-561 gene for 16S ribosomal RNA, partial sequence	
	Pseudomonas sp. 2B5B 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium clone nc437302c1 16S ribosomal RNA gene, partial sequence	Uncultured bacterium clone MWL-15 16S ribosomal RNA gene, partial sequence
	Pseudomonas sp. URM017WK12-II1 genome assembly YEL_emb1.gr, chromosome : 1	
	Pseudomonas fulva strain BSW3 16S ribosomal RNA gene, partial sequence	Pseudomonas parafulva strain CRS01-1, complete genome
	Pseudomonas sp. B-6 16S ribosomal RNA gene, partial sequence	Pseudomonas sp. LB-X-GYM-4-1 16S ribosomal RNA gene, partial sequence
	Pseudomonas sp. H10zhy partial 16S rRNA gene, strain H10zhy	Pseudomonas sp. LB-S-GYM-2 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nc42688b02c1 16S ribosomal RNA gene, partial sequence	
	ε-proteobacteria 6 leaves	
	Pseudomonas fulva strain SGRAJ09 16S ribosomal RNA gene, partial sequence	Uncultured bacterium clone nc4861g02c1 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone nc42598e06c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain BSW1 16S ribosomal RNA gene, partial sequence	Pseudomonas fulva strain QC07 16S ribosomal RNA gene, partial sequence
	Pseudomonas fulva 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva partial 16S rRNA gene, strain Z67zhy	
	Uncultured bacterium clone nc4908e02c1 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva gene for 16S rRNA, partial sequence, strain: NBRC 16639	
	Uncultured bacterium clone nbw643e01c1 16S ribosomal RNA gene, partial sequence	Pseudomonas fulva strain i19_MX 16S ribosomal RNA gene, partial sequence
	Bacterium AM0315 16S ribosomal RNA gene, partial sequence	
	Pseudomonas sp. NCCP-567 gene for 16S ribosomal RNA, partial sequence	
	Pseudomonas sp. 471-1 16S ribosomal RNA gene, partial sequence	Uncultured bacterium clone nc42670f06c1 16S ribosomal RNA gene, partial sequence
	Pseudomonas putida strain JT-K21 16S ribosomal RNA gene, partial sequence	
	Uncultured bacterium partial 16S rRNA gene, clone C407 74	
	ε-proteobacteria 4 leaves	
	ε-proteobacteria 3 leaves	
	Pseudomonas fulva strain BSW8 16S ribosomal RNA gene, partial sequence	
	Pseudomonas fulva strain CP-11 16S ribosomal RNA gene, partial sequence	
	Pseudomonas putida strain F29 16S ribosomal RNA gene, partial sequence	Uncultured bacterium clone nc42625d06c1 16S ribosomal RNA gene, partial sequence

Bacillus licheniformis KU314515.1

Bacillus licheniformis strain HS10 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KU314515.1|](#) Length: 1514 Number of Matches: 1

Range 1: 491 to 1441 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand	
1710 bits(1896)	0.0	950/951(99%)	0/951(0%)	Plus/Minus	
Query 1	CACCGACTTCGGGTGTTACAAACTCTCGTGGTGTGACGGGCGGTGTGTACAAGGCCCGGG	60			
Sbjct 1441					1382
Query 61	AACGTATTACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAGCTTCACGCAGTCG	120			
Sbjct 1381					1322
Query 121	AGTTGCAGACTGCGATCCGAACTGAGAACAGATTTGTGGGATTGGCTTAGCCTCGCGGCT	180			
Sbjct 1321					1262
Query 181	TCGCTGCCCTTTGTTCTGCCCATTTGTAGCACGTGTGTAGCCCAGGTCATAAGGGGCATGA	240			
Sbjct 1261					1202
Query 241	TGATTTGACGTCATCCCCACCTTCCTCCGGTTTGTACCGGCAGTCACCTTAGAGTGCCC	300			
Sbjct 1201					1142
Query 301	AACTGAATGCTGGCAACTAAGATCAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATC	360			
Sbjct 1141					1082
Query 361	TCACGACACGAGCTGACGACAACCATGCACCACCTGTCACTCTGCCCCGAAGGGGAAGC	420			
Sbjct 1081					1022
Query 421	CCTATCTCTAGGGTTGTCAGAGGATGTCAAGACCTGGTAAGGTTCTTCGCGTTGCTTCGA	480			
Sbjct 1021					962
Query 481	ATTAAACCACATGCTCCACCGCTTGTGCGGGCCCCGTCAATTCCTTTGAGTTTCAGTCT	540			
Sbjct 961					902
Query 541	TGCGACCGTACTCCCCAGGCGGAGTGCTTAATGCGTTTGCTGCAGCACTAAAGGGCGGAA	600			
Sbjct 901					842
Query 601	ACCTCTAACACTTAGCACTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGT	660			
Sbjct 841					782
Query 661	TCGCTCCCCACGCTTTCGCGCCTCAGCGTCAGTTACAGACCAGAGAGTCGCCTTCGCCAC	720			
Sbjct 781					722
Query 721	TGGTGTTCTCTCCACATCTCTACGCATTTACCGCTACACGTGGAATTCACCTCTCCTCTT	780			
Sbjct 721					662
Query 781	CTGCACTCAAGTTCCCCAGTTTCCAATGACCCTCCCCGGTTGAGCCGGGGGCTTTACAT	840			
Sbjct 661					602
Query 841	CAAACTTAAGAAACCGCCTGCGCGCGCTTTACGCCCAATAATTCCGGACAACGCTTGCCA	900			
Sbjct 601					542
Query 901	CCTACGTATTACCGCGGCTGCTGGCACGTAGTTAGCCGTGGCTTTCTGGTT	951			
Sbjct 541					491

● Bacillus sp. BAB-5485 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. RKJDB-0018 partial 16S rRNA gene, isolate RKJDB-0018
 ● Bacillus sp. RKNM-0105 partial 16S rRNA gene, isolate RKNM-0105
 ● Bacillus licheniformis strain KP050r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KP052r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain HS10 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain TSS15 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. BAB-5636 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain SIB_Zn_R4 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain FJAT-29133 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. S94(2016) 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. S37 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain VTB12 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain CQN-22 16S ribosomal RNA gene, partial sequence
 ● Bacillus sonorensis strain CQN-15 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain CQN-12 16S ribosomal RNA gene, partial sequence
 ● Bacillus sonorensis strain CQN-11 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain CQN-8 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain CY2-24 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain CY1-20 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain Y8 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KTNB0010 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain SMR8 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain SMR7 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain SMR6 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain EVR2 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain RA5UN 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain RA13UN 16S ribosomal RNA gene, partial sequence
 ● Uncultured bacterium clone 958-15 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. C-3-24 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. C-3-11 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. C-3-3 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B-3-27 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B-3-7 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. A-3-12 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. A-3-5 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. A-2-29 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. A-2-18 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. A-2-13 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. C-1-20 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. C-1-19 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. C-1-3 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B-1-40 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B-1-16 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B-1-15 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B31(2015) 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. B27(2015) 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain F1159 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain F1152 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain F147 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain F144 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain F142 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain I-A-E-34 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain JS-1 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain DMB31 16S ribosomal RNA gene, partial sequence
 ● Bacillus freudenreichii strain NCIM2463 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain NCIM2715 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain NCIM2471 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain NJ4-1 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. NG4-2 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. NL3-1 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. NZ3-2 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. NZ3-1 16S ribosomal RNA gene, partial sequence
 ● Bacillus flexus strain NZ2-2 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain 103D-012 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain N9 16S ribosomal RNA gene, partial sequence
 ● Bacterium ARB15 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain CY-012 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain IHBB 11006 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. BAB-5495 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. N156PBVB07_1492r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. N33621VB01_1492r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain A1 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. FJAT-22511 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis partial 16S rRNA gene, strain IRQBAS20
 ● Bacillus licheniformis partial 16S rRNA gene, strain IRQBAS19
 ● Bacillus licheniformis partial 16S rRNA gene, strain IRQBAS18
 ● Bacillus sp. NCIM 2131 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KAR76 16S ribosomal RNA gene, complete sequence
 ● Bacillus licheniformis WX-02 genome
 ● Bacillus sp. KP138r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP129r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP126r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KP120r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP097r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KP093r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KP091r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KP081r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP071r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP057r 16S ribosomal RNA gene, partial sequence
 ● Bacillus licheniformis strain KP056r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP055r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. KP053r 16S ribosomal RNA gene, partial sequence
 ● Bacillus sp. H15-1 16S ribosomal RNA gene, partial sequence

[id|Query_71993](#)

Pseudomonas aeruginosa **KF680991.1**

Pseudomonas aeruginosa strain ATHA23 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KF680991.1](#) Length: 1115 Number of Matches: 1

Range 1: 86 to 1065 [GenBank](#) [Graphics](#)

[▼ Next Match](#) [▲ Previous Match](#)

Score	Expect	Identities	Gaps	Strand
1763 bits(1954)	0.0	979/980(99%)	0/980(0%)	Plus/Minus
Query 1	TTCTGGAGCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTAT	60		
Sbjct 1065	TTCTGGAGCAACCCACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTAT	1006		
Query 61	TCACCGTGACATTCTGATTACGATTACTAGCGATTCCGACTTCACGCAGTCGAGTTGCA	120		
Sbjct 1005	TCACCGTGACATTCTGATTACGATTACTAGCGATTCCGACTTCACGCAGTCGAGTTGCA	946		
Query 121	GACTGCGATCCGGACTACGATCGGTTTTATGGGATTAGCTCCACCTCGCGGCTTGGCAAC	180		
Sbjct 945	GACTGCGATCCGGACTACGATCGGTTTTATGGGATTAGCTCCACCTCGCGGCTTGGCAAC	886		
Query 181	CCTTTGTACCGACCATTTGTAGCACGTGTGTAGCCCTGGCCGTAAGGGCCATGATGACTTG	240		
Sbjct 885	CCTTTGTACCGACCATTTGTAGCACGTGTGTAGCCCTGGCCGTAAGGGCCATGATGACTTG	826		
Query 241	ACGTCATCCCCACCTTCCTCCGTTTGTACCGGCAGTCTCCTTAGAGTGCCACCCGAG	300		
Sbjct 825	ACGTCATCCCCACCTTCCTCCGTTTGTACCGGCAGTCTCCTTAGAGTGCCACCCGAG	766		
Query 301	GTGCTGGTAACTAAGGACAAGGGTTGCGCTCGTTACGGGACTTAACCCAACATCTCACGA	360		
Sbjct 765	GTGCTGGTAACTAAGGACAAGGGTTGCGCTCGTTACGGGACTTAACCCAACATCTCACGA	706		
Query 361	CACGAGCTGACGACAGCCATGCAGCACCTGTGTCTGAGTTCCCGAAGGCACCAATCCATC	420		
Sbjct 705	CACGAGCTGACGACAGCCATGCAGCACCTGTGTCTGAGTTCCCGAAGGCACCAATCCATC	646		
Query 421	TCTGGAAAGTTCTCAGCATGTCAAGGCCAGGTAAGGTTCTTCGCGTTGCTTCGAATTAAA	480		
Sbjct 645	TCTGGAAAGTTCTCAGCATGTCAAGGCCAGGTAAGGTTCTTCGCGTTGCTTCGAATTAAA	586		
Query 481	CCACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCAATTTAGTTTTAACTTGC	540		
Sbjct 585	CCACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCAATTTAGTTTTAACTTGC	526		
Query 541	CGTACTCCCCAGGCGGTGCGACTTATCGCGTTAGCTGCGCCACTAAGATCTCAAGGATCCC	600		
Sbjct 525	CGTACTCCCCAGGCGGTGCGACTTATCGCGTTAGCTGCGCCACTAAGATCTCAAGGATCCC	466		
Query 601	AACGGCTAGTCGACATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCC	660		
Sbjct 465	AACGGCTAGTCGACATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCC	406		
Query 661	CCACGCTTTTCGCACCTCAGTGTGAGTATCAGTCCAGGTGGTTCGCTTCGCCACTGGTGT	720		
Sbjct 405	CCACGCTTTTCGCACCTCAGTGTGAGTATCAGTCCAGGTGGTTCGCTTCGCCACTGGTGT	346		
Query 721	CCTTCCTATATCTACGCATTTACCGCTACACAGGAAATTCACCACCCTCTACCGTACT	780		
Sbjct 345	CCTTCCTATATCTACGCATTTACCGCTACACAGGAAATTCACCACCCTCTACCGTACT	286		
Query 781	CTAGCTCAGTAGTTTTGGATGCAGTTCCAGGTTGAGCCCGGGGATTTCACATCCAAC	840		
Sbjct 285	CTAGCTCAGTAGTTTTGGATGCAGTTCCAGGTTGAGCCCGGGGATTTCACATCCAAC	226		
Query 841	GCTGAACCACCTACGCGCGCTTTACGCCAGTAATTCGATTAAACGCTTGACCCCTTCGT	900		
Sbjct 225	GCTGAACCACCTACGCGCGCTTTACGCCAGTAATTCGATTAAACGCTTGACCCCTTCGT	166		
Query 901	ATTACGCGGCTGCTGGCACGAAGTTAGCCGGGCTTATTCTGTTGGTAACGTCAAACAG	960		
Sbjct 165	ATTACGCGGCTGCTGGCACGAAGTTAGCCGGGCTTATTCTGTTGGTAACGTCAAACAG	106		
Query 961	CAAGGTATTAACTTACTGCC	980		
Sbjct 105	CAAGGTATTAACTTACTGCC	86		

- *Pseudomonas aeruginosa* strain HMT 7 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain HMT 2 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain HMT51 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain B7 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain A1 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain NBAl AFP-6 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain NBAl AFP-3 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain NBAl ND-4 IART-B 16S ribosomal RNA gene, partial sequence
- *Bacterium* C37 16S ribosomal RNA, partial sequence
- *Pseudomonas otitidis* strain FJYM11 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain C37 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain BM6 16S ribosomal RNA gene, partial sequence
- *Pseudomonas aeruginosa* strain ATHA23 16S ribosomal RNA gene, partial sequence
- *Bacterium* WU1 16S ribosomal RNA gene, partial sequence

Agrobacterium larrymoorei EF178437.1

Agrobacterium larrymoorei strain 2R46 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|EF178437.1](#) Length: 1414 Number of Matches: 1

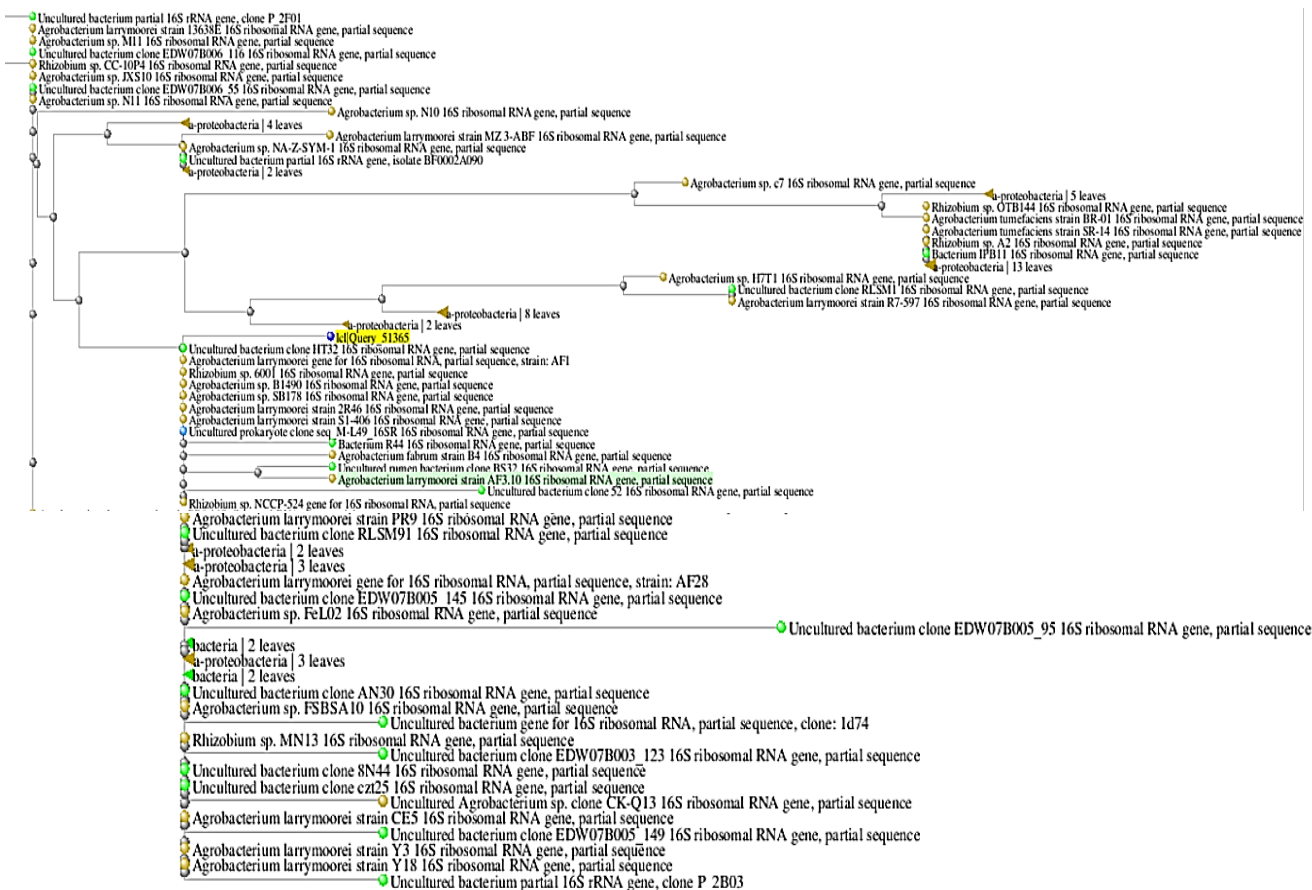
Range 1: 398 to 1329 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1676 bits(1858)	0.0	931/932(99%)	0/932(0%)	Plus/Minus

Query	1	GGGTAAAACCAACTCCCATGGTGTGACGGGCGGTGTGTACAAGGCCCGGGAACGTATTCA	60
Sbjct	1329		1270
Query	61	CCGCAGCATGCTGATCTGCGATTACTAGCGATTCCAATTTCATGCACTCGAGTTGCAGAG	120
Sbjct	1269		1210
Query	121	TGCAATCCGAACGTAGATGGCTTTTGGAGATTAGCTCGACATCGCTGTCTCGCTGCCAC	180
Sbjct	1209		1150
Query	181	TGTCACCACCATTGTAGCACGTGTGTAGCCAGCCCGTAAGGGCCATGAGGACTTGACGT	240
Sbjct	1149		1090
Query	241	CATCCCCACCTTCTCTCGGCTTATCACCAGCAGTCCCTTAGAGTGCCCAACCAAATGC	300
Sbjct	1089		1030
Query	301	TGGCAACTAAGGGCGAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCACGACACG	360
Sbjct	1029		970
Query	361	AGCTGACGACAGCCATGCAGCACCTGTTCTAGGGCCAGCCGAAGTGAAGGTCATCGTCTC	420
Sbjct	969		910
Query	421	CAATGACCATAACCCGAATGTCAAGAGCTGGTAAGGTTCTGCGCGTTGCTTCGAATTAAA	480
Sbjct	909		850
Query	481	CCACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTAAATCTTGCAG	540
Sbjct	849		790
Query	541	CGTACTCCCCAGGCGGAATGTTTAAATGCGTTAGCTGCGCCACCGAACAGTATACTGCCCG	600
Query	601	ACGGCTAACATTCATCGTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCC	660
Sbjct	729		670
Query	661	CACGCTTTCGCACCTCAGCGTCAGTAATGGACCAGTAAGCCGCCTTCGCCACTGGTGTTC	720
Sbjct	669		610
Query	721	CTGCGAATATCTACGAATTTACCTCTACACTCGCAATTCACCTACCTCTTCCATACTC	780
Sbjct	609		550
Query	781	AAGATACCCAGTATCAAAGGCAGTTCCAGAGTTGAGCTCTGGGATTTACCCCTGACTTA	840
Sbjct	549		490
Query	841	AATATCCGCCTACGTGCGCTTTACGCCAGTAATTCGAACAACGCTAGCCCCCTTCGTA	900
Sbjct	489		430
Query	901	TTACCGCGGCTGCTGGCACGAAGTTAGCCGGG	932
Sbjct	429		398

actnchi.nlm.nih.gov/Blast.cgi#dlnDwnl_127892477



Pantoea septica KF475883.1

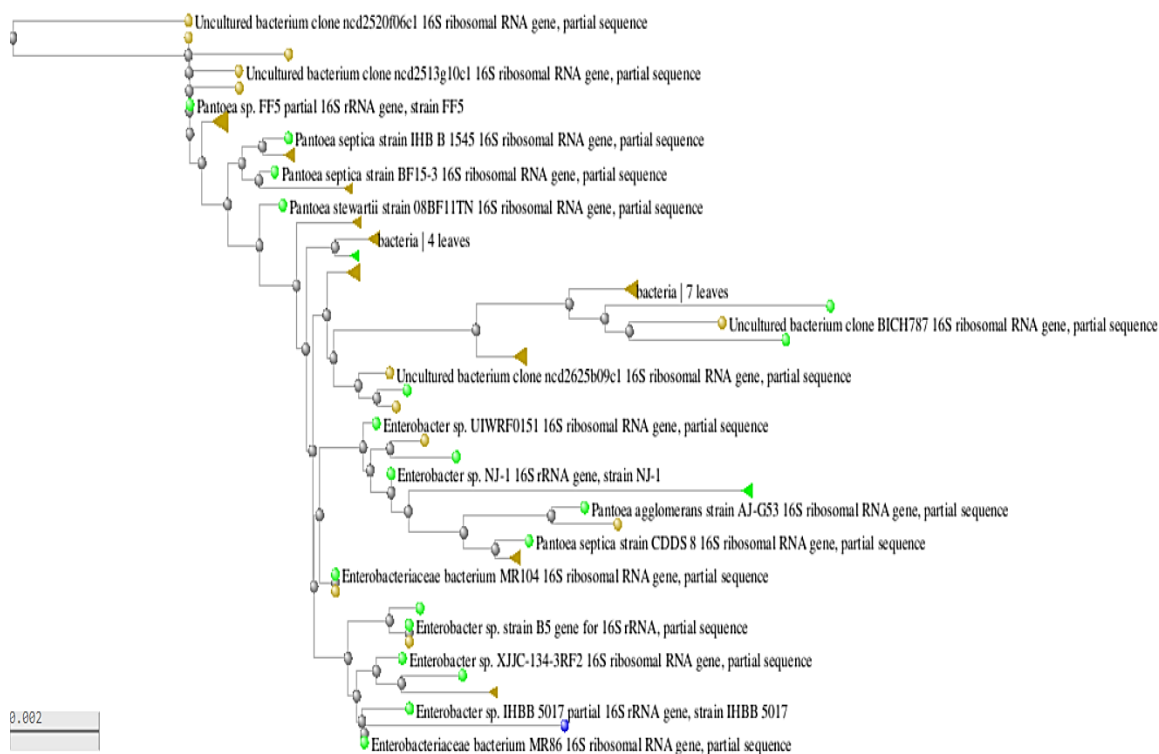
Pantoea septica strain IHB B 1545 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KF475883.1](#) Length: 1508 Number of Matches: 1

Range 1: 68 to 975 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
1599 bits(1772)	0.0	900/908(99%)	1/908(0%)	Plus/Plus
Query 1	ACAGAAGAGCTTGCTCTTTGGGTGGCGAGTGGCGGACGGGTGAGTAATGTCTGGGAAACT	60		
Sbjct 68	ACAGAAGAGCTTGCTCTTTGGGTGGCGAGTGGCGGACGGGTGAGTAATGTCTGGGAAACT	127		
Query 61	GCCCCGATGGAGGGGGATAACTACTGGAACGGTAGCTAATACCGCATAACGTCGCAAGAC	120		
Sbjct 128	GCCCCGATGGAGGGGGATAACTACTGGAACGGTAGCTAATACCGCATAACGTCGCAAGAC	187		
Query 121	CAAAGTGGGGGACCTTCGGGCCTCACACCATCGGATGTGCCAGATGGGATTAGCTAGTA	180		
Sbjct 188	CAAAGTGGGGGACCTTCGGGCCTCACACCATCGGATGTGCCAGATGGGATTAGCTAGTA	247		
Query 181	GGTGGGGTAACGGCTCACCTAGGCGACGATCCCTAGCTGGTCTGAGAGGATGACCAGCCA	240		
Sbjct 248	GGTGGGGTAACGGCTCACCTAGGCGACGATCCCTAGCTGGTCTGAGAGGATGACCAGCCA	307		
Query 241	CACCTGGAAGTGAAGACACGGTCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACA	300		
Sbjct 308	CACCTGGAAGTGAAGACACGGTCCAGACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACA	367		
Query 301	ATGGGCGCAAGCCTGATGCAGCCATGCCGCGTGTATGAAGAAGGCCCTTCGGGTTGTAAAG	360		
Sbjct 368	ATGGGCGCAAGCCTGATGCAGCCATGCCGCGTGTATGAAGAAGGCCCTTCGGGTTGTAAAG	427		
Query 361	TACTTTTACGCGGGGAGGAAGGCGACGCGGTTAATAACCGCGTCGATTGACGTTACCCGCA	420		
Sbjct 428	TACTTTTACGCGGGGAGGAAGGCGACGCGGTTAATAGCCGCGTCGATTGACGTTACCCGCA	487		
Query 421	GAAGAAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGAGGGTGCAAGCGTTA	480		
Sbjct 488	GAAGAAGCACCGGCTAACTCCGTGCCAGCAGCCGCGGTAATACGAGGGTGCAAGCGTTA	547		
Query 481	ATCGGAATTACTGGGCGTAAAGCGCACGCAGGCGGTCTGTCAAGTCGGATGTGAAATCCC	540		
Sbjct 548	ATCGGAATTACTGGGCGTAAAGCGCACGCAGGCGGTCTGTCAAGTCGGATGTGAAATCCC	607		
Query 541	CGGGCTTAACCTGGGAACGTCATTGCAAACTGGCAGGCTAGAGTCTTGTAGAGGGGGGTA	600		
Sbjct 608	CGGGCTTAACCTGGGAACGTCATTGCAAACTGGCAGGCTAGAGTCTTGTAGAGGGGGGTA	667		
Query 601	GAATTCCAGGTGTAGCGGTGAAATGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCG	660		
Sbjct 668	GAATTCCAGGTGTAGCGGTGAAATGCGTAGAGATCTGGAGGAATACCGGTGGCGAAGGCG	727		
Query 661	GCCCCCTGGACAAAGACTGACGCTCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGAT	720		
Sbjct 728	GCCCCCTGGACAAAGACTGACGCTCAGGTGCGAAAGCGTGGGGAGCAAACAGGATTAGAT	787		
Query 721	ACCCTGGTAGTCCACGCCGTAAACGATGTCGACTTGGAGGCTGTTCCCTGAGGAGTGGC	780		
Sbjct 788	ACCCTGGTAGTCCACGCCGTAAACGATGTCGACTTGGAGGCTGTTCCCTGAGGAGTGGC	847		
Query 781	TTCCGGAGCTAACGCGTTAAGTCGACCGCCTGGGGAGTACGGGCGCAAGGTTAAACTCA	840		
Sbjct 848	TTCCGGAGCTAACGCGTTAAGTCGACCGCCTGGGGAGTACGGGCGCAAGGTTAAACTCA	907		
Query 841	AATGAATTGACGGGGGCCGCCAAGCGGGGAGCATGTGGTTTAAATTCGATGC-ACGCG	899		
Sbjct 908	AATGAATTGACGGGGGCCGCCAAGCGGTGGAGCATGTGGTTTAAATTCGATGCAACGCG	967		
Query 900	AAAAACCT 907			
Sbjct 968	AAGAACCT 975			



***Stenotrophomonas rhizophila* KP050794.1**

Stenotrophomonas rhizophila strain HT12-MRL 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KP318062.1|](#) Length: 1295 Number of Matches: 1

Range 1: 59 to 491 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
760 bits(842)	0.0	429/433(99%)	1/433(0%)	Plus/Plus
Query 37	GGGTGAGGAA-ACATCGTTGTCTACCTTTTCGTGGGGGATAACGTAGGGAACTTACGCT	95		
Sbjct 59	GGGTGAGGAATACATCGGAATCTACCTTTTCGTGGGGGATAACGTAGGGAACTTACGCT	118		
Query 96	AATACCGCATACGACCTTCGGGTGAAAGCAGGGGACCTTCGGGCCTTGCGCGGATAGATG	155		
Sbjct 119	AATACCGCATACGACCTTCGGGTGAAAGCAGGGGACCTTCGGGCCTTGCGCGGATAGATG	178		
Query 156	AGCCGATGTCGGATTAGCTAGTTGGCGGGGTAAAGGCCACCAAGGCGACGATCCGTAGC	215		
Sbjct 179	AGCCGATGTCGGATTAGCTAGTTGGCGGGGTAAAGGCCACCAAGGCGACGATCCGTAGC	238		
Query 216	TGGTCTGAGAGGATGATCAGCCACACTGGAACGTAGACACGGTCCAGACTCCTACGGGAG	275		
Sbjct 239	TGGTCTGAGAGGATGATCAGCCACACTGGAACGTAGACACGGTCCAGACTCCTACGGGAG	298		
Query 276	GCAGCAGTGGGGAATATTGGACAATGGGCGCAAGCCTGATCCAGCCATACCGCGTGGGTG	335		
Sbjct 299	GCAGCAGTGGGGAATATTGGACAATGGGCGCAAGCCTGATCCAGCCATACCGCGTGGGTG	358		
Query 336	AAGAAGGCCTTCGGGTGTAAAGCCCTTTTGTGGGAAAGAAAAGCAGTCGATTAATACT	395		
Sbjct 359	AAGAAGGCCTTCGGGTGTAAAGCCCTTTTGTGGGAAAGAAAAGCAGTCGATTAATACT	418		
Query 396	CGGTTGTTCTGACGGTACCCAAAGAATAAGCACCGGCTAACTTCGTGCCAGCAGCCGCGG	455		
Sbjct 419	CGGTTGTTCTGACGGTACCCAAAGAATAAGCACCGGCTAACTTCGTGCCAGCAGCCGCGG	478		
Query 456	TAATACGAAGGGT	468		
Sbjct 479	TAATACGAAGGGT	491		



Citrobacter freundii CP007557

Citrobacter freundii CFNIH1, complete genome

Sequence ID: [gb|CP007557.1|](#) Length: 5099034 Number of Matches: 1

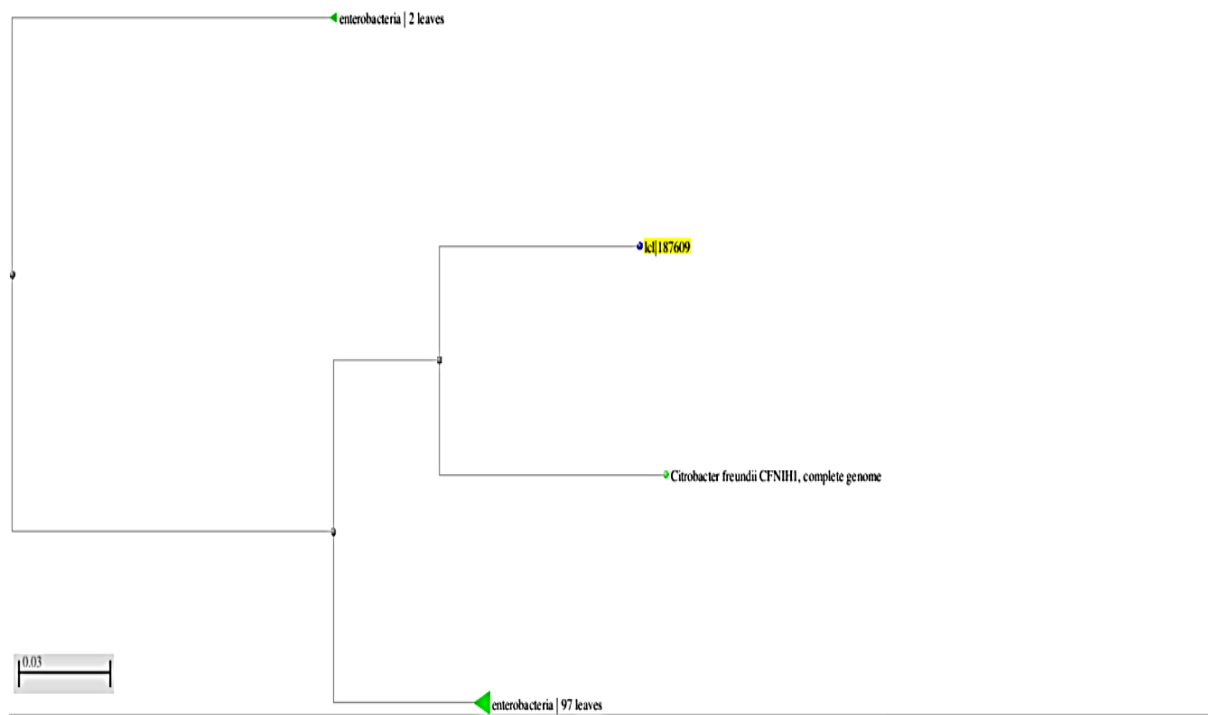
Range 1: 5039222 to 5039628 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
477 bits(528)	6e-131	353/408(87%)	3/408(0%)	Plus/Plus

Features: [hypothetical protein](#)
[phospho-2-dehydro-3-deoxyheptonate aldolase](#)

Query	1	TTTTTGCATGATGGTGATCCTGTTTAGCTCGTTTGGCATAGTTGATCCTCAGCGAGGAGG	60
Sbjct	5039222	TTTTTGCATGATGGTGATCCTGTTTAGCTCGTTTGGCATAGTTGATCCTCAGCGAGGAGG	5039281
Query	61	AAATAACGATACCATAACAGGTAAAGATTCAATCCACAATCCGTAAATTTAATTTACA	120
Sbjct	5039282	AAATAACGATATCACAACAGGTAAAGATTCAATCCACAATCCGTAAATTTAATTTACA	5039341
Query	121	CAGTGTATTTTAAAGGCAAAATAGCCCTT-ATAAATGTACACTTAAATTTACACCA-CG	178
Sbjct	5039342	TCGAGCTATTA-ATCACTAAATAAGGCCTACATGAGTGTATAGTTAAATTTACACAATCC	5039400
Query	179	AATTTTCAGATCCGCTATGCTTaaaaaaaCAAGGGAGCACAGGCAATGAAGCAACTCATC	238
Sbjct	5039401	TGATATCAGATCCACTATGCTTAAAAAACGCGGGAGCACAGACAATGAAGCAACTCATC	5039460
Query	239	AGCATCTTATTTCTATTCCTTCTTAGCGGATGTCAGATAGATCCCTACACCCATGCCCT	298
Sbjct	5039461	AGCACCTTATTAATACTACTTCTTAGCGGATGTCAGATAGACCCCTATACTCAGGCCCT	5039520
Query	299	ACCTGGACTGGTACCGACTGGTATGACGCCGGGATACAAGATGCCATTCGGGGCTATGCC	358
Sbjct	5039521	ACCTGGACTGGCACCAGCTGGTACGATGCCGGCATAACGATGCCATTCAGGCTATGCG	5039580
Query	359	GTTAAAGATAATGAAACTCTTGCCGACAATTCAATGATCCCGAAGTC	406
Sbjct	5039581	GTTAAAGATAATGAAATCTTGCCGACAATTACAATGATCCAGAAGTC	5039628



Pseudomonas frederiksbergensis EU373369.1

Pseudomonas frederiksbergensis strain PR18 16S ribosomal RNA gene, partial sequence

Sequence ID: [gb|KJ870030.1](#) Length: 1449 Number of Matches: 1

Range 1: 832 to 1350 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
919 bits(1018)	0.0	515/519(99%)	0/519(0%)	Plus/Minus
Query 6	GTGTGTAATAATGCGCGGGAACGTATTACCGCGACATTCTGATTTCGCGATTACTAGCGAT	65		
Sbjct 1350	GTGTGTACAAGGCCCGGGAACGTATTACCGCGACATTCTGATTTCGCGATTACTAGCGAT	1291		
Query 66	TCCGACTTCACGCAGTCGAGTTGCAGACTGCGATCCGGACTACGATCGGTTTTATGGGAT	125		
Sbjct 1290	TCCGACTTCACGCAGTCGAGTTGCAGACTGCGATCCGGACTACGATCGGTTTTCTGGGAT	1231		
Query 126	TAGCTCCACCTCGCGGCTTGGCAACCCCTCTGTACCGACCATTGTAGCACGTGTGTAGCCC	185		
Sbjct 1230	TAGCTCCACCTCGCGGCTTGGCAACCCCTCTGTACCGACCATTGTAGCACGTGTGTAGCCC	1171		
Query 186	AGGCCGTAAGGGCCATGATGACTTGACGTCATCCCCACCTTCTCCGGTTTGTCAACGGC	245		
Sbjct 1170	AGGCCGTAAGGGCCATGATGACTTGACGTCATCCCCACCTTCTCCGGTTTGTCAACGGC	1111		
Query 246	AGTCTCCTTAGAGTGCCCAACATTACGTGCTGGTAACTAAGGACAAGGGTTGCGCTCGTT	305		
Sbjct 1110	AGTCTCCTTAGAGTGCCCAACATTACGTGCTGGTAACTAAGGACAAGGGTTGCGCTCGTT	1051		
Query 306	ACGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAGCCATGCAGCACCTGTCTC	365		
Sbjct 1050	ACGGGACTTAACCCAACATCTCACGACACGAGCTGACGACAGCCATGCAGCACCTGTCTC	991		
Query 366	AATGTTCCCGAAGGCACCAATCCATCTCTGGAAAGTTCATTGGATGTCAAGGCCTGGTAA	425		
Sbjct 990	AATGTTCCCGAAGGCACCAATCCATCTCTGGAAAGTTCATTGGATGTCAAGGCCTGGTAA	931		
Query 426	GTTTCTTCGCGTTGCTTCGAATTAACACATGCTCCACCGCTTGTGCGGGCCCCCGTCA	485		
Sbjct 930	GTTTCTTCGCGTTGCTTCGAATTAACACATGCTCCACCGCTTGTGCGGGCCCCCGTCA	871		
Query 486	ATTCATTGAGTTTTTAACCTTGCGGCCGTACTCCCCCAG	524		
Sbjct 870	ATTCATTGAGTTTTTAACCTTGCGGCCGTACTCCCCCAG	832		

klQuery_233167	
Pseudomonas sp. HNRI3 16S ribosomal RNA gene, partial sequence	Pseudomonas vancoverensis strain BD44 16S ribosomal RNA gene, partial sequence
	Uncultured bacterium clone B12 16S ribosomal RNA gene, partial sequence
	g-proteobacteria 4 leaves
	Pseudomonas putida strain DNR504 16S ribosomal RNA gene, partial sequence
Pseudomonas sp. s1p23 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s2p12 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s2p21 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s2p23 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s3p21 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. s4p22 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. BSP27 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. PT19 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. SCALX0301 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. Gwa3-10 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. P32/2013 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. QW11 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone CD36 16S ribosomal RNA gene, partial sequence	
Pseudomonas chlororaphis partial 16S rRNA gene, isolate ToZa7	
Uncultured Pseudomonas sp. clone BIPSS22-01 16S ribosomal RNA gene, partial sequence	
Pseudomonas cedrina strain LCQ-5 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. RA-20 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. S10114 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. SB442 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. XBBSA2 16S ribosomal RNA gene, partial sequence	
Pseudomonas extremorientalis strain EB-171 16S ribosomal RNA gene, partial sequence	
Pseudomonas vancoverensis strain Amp46 16S ribosomal RNA gene, partial sequence	
Pseudomonas extremorientalis strain NSPMB02 16S ribosomal RNA gene, partial sequence	
Pseudomonas extremorientalis strain NSPMB02 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. P2-1 16S ribosomal RNA gene, partial sequence	
Pseudomonas chlororaphis strain 7.3B 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. C34 16S ribosomal RNA gene, partial sequence	
Endophytic bacterium 10 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IN2CAET02BYJ10 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE05CD4X 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE04EFT0G 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE07IAZG1 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE07IDFY 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE07IKPDF 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone IE34LQE08ITYDU 16S ribosomal RNA gene, partial sequence	
Uncultured bacterium clone LY62 16S ribosomal RNA gene, partial sequence	
Uncultured bacterium clone HZ37 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. QJX-1 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. AD157 16S ribosomal RNA gene, partial sequence	
Pseudomonas azotiformans strain BG D 16S ribosomal RNA gene, partial sequence	
Pseudomonas umongensis strain PVR02 16S ribosomal RNA gene, partial sequence	
Pseudomonas azotiformans strain Sn16 16S ribosomal RNA gene, partial sequence	
Pseudomonas azotiformans strain Sn22 16S ribosomal RNA gene, partial sequence	
Pseudomonas azotiformans strain Sn47 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. DR 1-03 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. GR 7-06 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. NR 6-08 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. UT 3-02 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone RHDTWG191 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone RHDTWG205 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone RHDTWG209 16S ribosomal RNA gene, partial sequence	
Uncultured Pseudomonas sp. clone RHDTWG214 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. COB-2P 16S ribosomal RNA gene, partial sequence	
Rhodococcus sp. COB-3P 16S ribosomal RNA gene, partial sequence	
Pseudomonas marginalis strain CTE722-C 16S ribosomal RNA gene, partial sequence	
Pseudomonas sp. ACP_01 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-R15_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-R17_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-R43_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-R48_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-R49_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S13_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S15_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S19_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S22_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S25_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S29_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S3_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S30_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S31_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S35_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S37_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S47_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S5_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S56_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_M-S9_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-R16_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-R29_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-R34_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-R47_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-S43_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-S46_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_T-S9_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_C04_Z1224_Z014717a_W-X2_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_C05_Z1224_Z014718a_W-X3_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_C06_Z1224_Z014719a_W-X6_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_TH08_Z118570_Z079335a_W-X1_16SR 16S ribosomal RNA gene, partial sequence	
Uncultured prokaryote clone seq_TH09_Z118570_Z079336a_W-X9_16SR 16S ribosomal RNA gene, partial sequence	
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